

Non-targeted analysis approaches: A perspective from the USA

P. Lee Ferguson

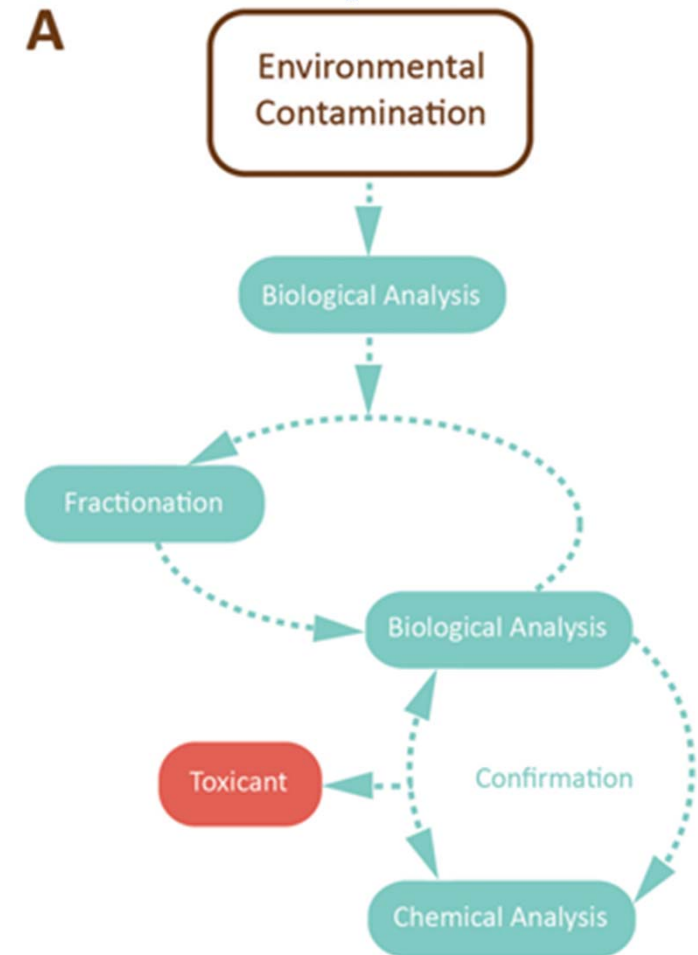
Department of Civil & Environmental Engineering and Nicholas School
of the Environment, Duke University, Durham, NC

lee.ferguson@duke.edu



Evolution of non-targeted analysis approaches within the US scientific community

- Toxicity Identification and Evaluation (TIE) – Early attempt at EDA
- Identification of “causative stressors” driven by observed sediment (usually) toxicity.
- Recent attempts at *in silico* emerging contaminant “prioritization” provide a roadmap (e.g. Howard and Muir).
- No systematic US framework (e.g. NORMAN) exists for non-targeted analysis.



R. M. Burgess, K. T. Ho, W. Brack, M. Lamoree, *Environ. Toxicol. Chem.* 2013, 32. 1935-1945

What have we learned from sediment TIE experiments over the years?

Environmental Toxicology and Chemistry, Vol. 32, No. 11, pp. 2424–2432, 2013
 © 2013 SETAC
 Printed in the USA

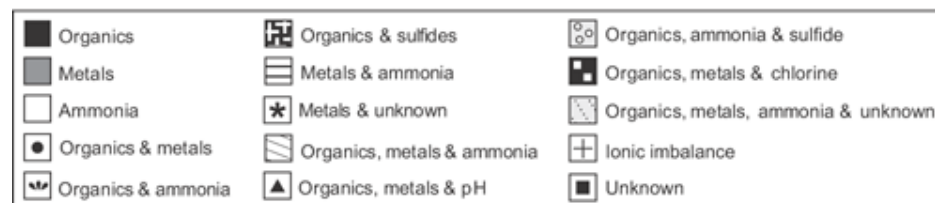
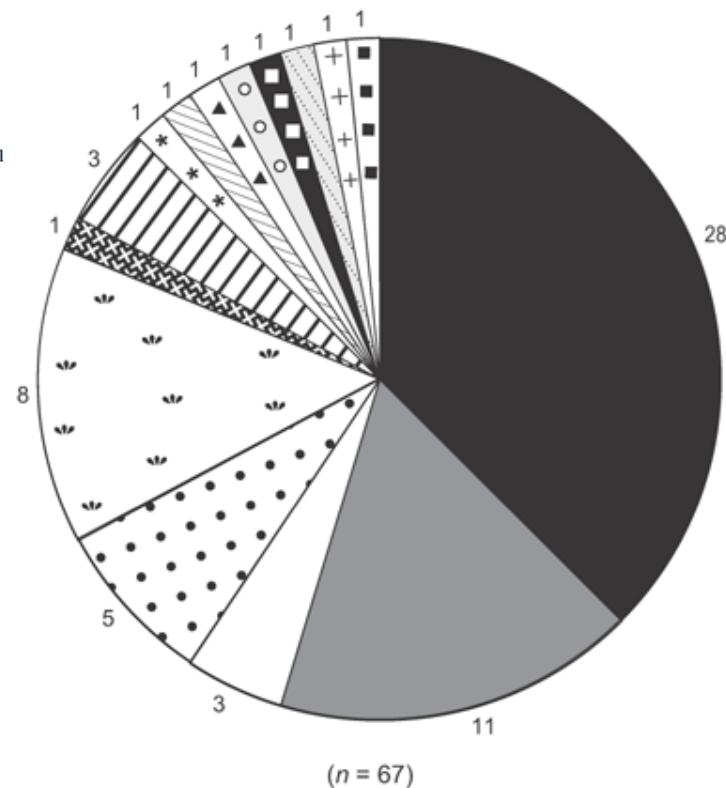
Critical Review

WHAT'S CAUSING TOXICITY IN SEDIMENTS? RESULTS OF 20 YEARS OF TOXICITY IDENTIFICATION AND EVALUATIONS

KAY T. HO* and ROBERT M. BURGESS

National Health and Environmental Effects Research Laboratory, Atlantic Ecology Division, Office of Research and Development, US Environmental Protection Agency, Narragansett, Rhode Island, USA

- Nonionic organics accounted for the largest fraction of observed toxicity.
- Whole-sediment TIE implicated nonionic organics in 90% of cases.
- “Molecular” identity of causative stressor was not always determined.



The “Howard and Muir” studies: Providing a roadmap for rational non-targeted analysis

Environ. Sci. Technol. 2006, 40, 7157–7166

Are There Other Persistent Organic Pollutants? A Challenge for Environmental Chemists[†]

DEREK C. G. MUIR*[†] AND PHILIP H. HOWARD[‡]
Water Science and Technology Directorate, Environment Canada, Burlington, Ontario, Canada, and Syracuse Research Corporation, Environmental Science Center, North Syracuse, New York

Environ. Sci. Technol. 2010, 44, 2277–2285

Identifying New Persistent and Bioaccumulative Organics Among Chemicals in Commerce

PHILIP H. HOWARD*[†] AND
DEREK C. G. MUIR[‡]
SRC, Environmental Science Center, 6502 Round Pond Road, North Syracuse, New York, and Aquatic Ecosystem Protection Research Division, Environment Canada, 867 Lakeshore Road, Burlington, Ontario

ENVIRONMENTAL
Science & Technology

Identifying New Persistent and Bioaccumulative Organics Among Chemicals in Commerce II: Pharmaceuticals

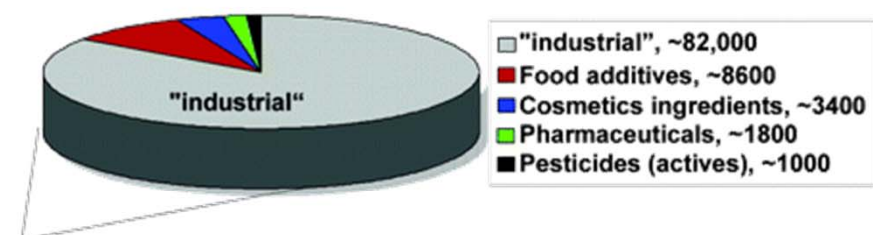
Philip H. Howard*[†] and Derek C. G. Muir[‡]

[†]SRC, Inc. Chemical, Biological, and Environmental Center (CBEC), 7502 Round Pond Road, North Syracuse, New York, and

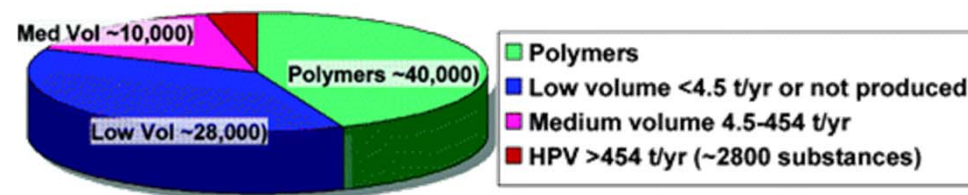
[‡]Aquatic Ecosystem Protection Research Division, Environment Canada, 867 Lakeshore Road, Burlington, Ontario

[dx.doi.org/10.1021/es201196x](https://doi.org/10.1021/es201196x) | *Environ. Sci. Technol.* 2011, 45, 6938–6946

A. Breakdown of the Chemicals in commerce – USA



B. “Industrial” Chemicals in commerce – US TSCA inventory



ENVIRONMENTAL
Science & Technology

Identifying New Persistent and Bioaccumulative Organics Among Chemicals in Commerce. III: Byproducts, Impurities, and Transformation Products

Philip H. Howard*[†] and Derek C. G. Muir[‡]

[†]SRC, Inc. Defense and Environmental Solutions (DES), 7502 Round Pond Road, North Syracuse, New York 13212, United States

[‡]Environment Canada, Aquatic Contaminants Research Division, 867 Lakeshore Road, Burlington, Ontario, Canada

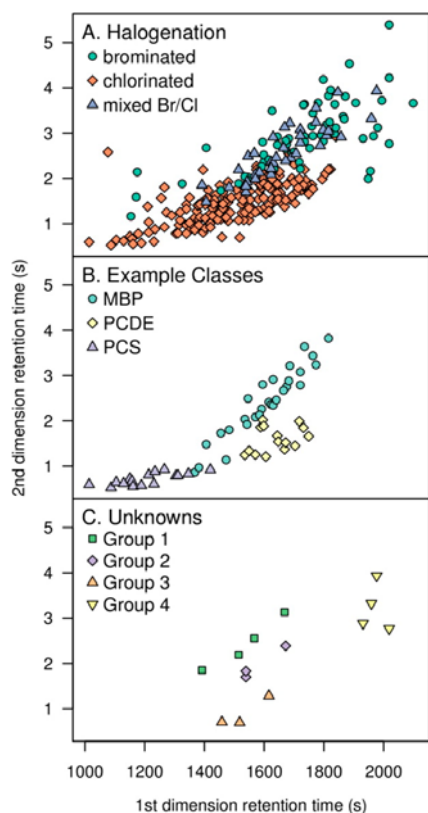
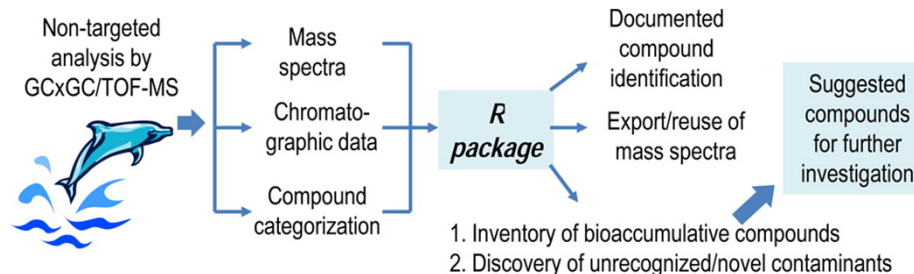
[dx.doi.org/10.1021/es40040751](https://doi.org/10.1021/es40040751) | *Environ. Sci. Technol.* 2013, 47, 5259–5266

Putting this roadmap to the test: Non-targeted analysis of POPs in dolphins

Nontargeted Comprehensive Two-Dimensional Gas Chromatography/Time-of-Flight Mass Spectrometry Method and Software for Inventorying Persistent and Bioaccumulative Contaminants in Marine Environments

Eunha Hoh,^{*,†} Nathan G. Dodder,^{*,‡} Steven J. Lehotay,[§] Kristin C. Pangallo,^{||} Christopher M. Reddy,[⊥] and Keith A. Maruya[‡]

dx.doi.org/10.1021/es301139q | Environ. Sci. Technol. 2012, 46, 8001–8008



- 271 Unique compounds were identified, all but one were halogenated.
- 2D GC separation allowed clustering by “compound class”.
- Many compounds were likely natural products (e.g. methylbipyrroles).
- Halogenated natural products were present at concentrations in blubber similar to PBDE congeners.

Table 3. Concentrations (ng/g of lipid mass) of Selected Halogenated Natural Products Compared to the Six Major PBDE Congeners

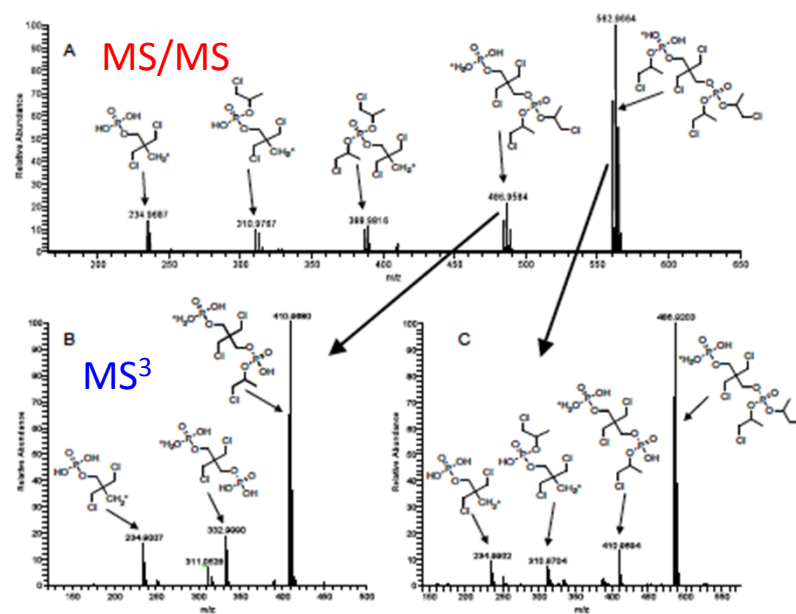
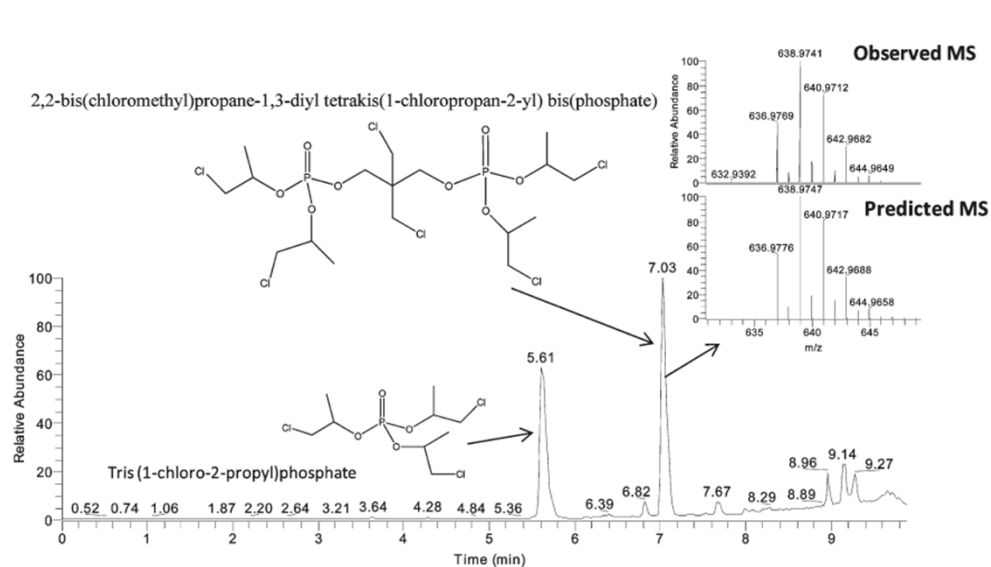
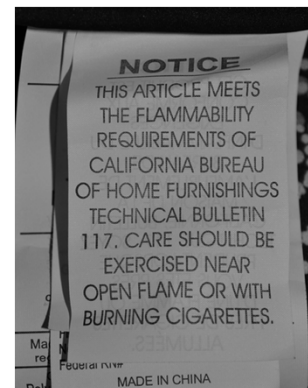
compd	concn	compd	concn
MBP-Cl ₇	85	2,2'-diMeO-BB-80	12.8
MBP-HBr ₃ Cl	1110	PBHD (3Br)	18
MBP-HBr ₆	478	PBHD (4Br)	246
MBP-Br ₆ Cl	1.62	BDE-28	8.6
MBP-Br ₇	0.504	BDE-47	727
DMBP-Br ₄ Cl ₂	124	BDE-100	241
DMBP-Br ₃ Cl	16.5	BDE-99	123
DMBP-Br ₆	30.9	BDE-154	103
2'-MeO-BDE68	47	BDE-153	51.3
6-MeO-BDE47	103		

What can you do when the compound is not present in any chemical registry?

Identification of Flame Retardants in Polyurethane Foam Collected from Baby Products

Heather M. Stapleton,^{*,†} Susan Klosterhaus,[‡] Alex Keller,[†] P. Lee Ferguson,[†] Saskia van Bergen,[§] Ellen Cooper,[†] Thomas F. Webster,^{||} and Arlene Blum[⊥]

[dx.doi.org/10.1021/es2007462](https://doi.org/10.1021/es2007462) | *Environ. Sci. Technol.* 2011, 45, 5323–5331



Example approaches from my laboratory:

1. **Effects-directed analysis:** Identifying toxic components of aircraft deicing/anti-icing fluids (ADAF)
2. **Activity-directed analysis:** Receptor affinity extraction for identifying estrogenic compounds associated with water reuse
3. **Fate-directed analysis:** Non-targeted analysis of micropollutant fate in wastewater treatment

Example approaches from my laboratory:

1. **Effects-directed analysis:** Identifying toxic components of aircraft deicing/anti-icing fluids (ADAF)
2. **Activity-directed analysis:** Receptor affinity extraction for identifying estrogenic compounds associated with water reuse
3. **Fate-directed analysis:** Non-targeted analysis of micropollutant fate in wastewater treatment



Type IV Anti-icing fluid

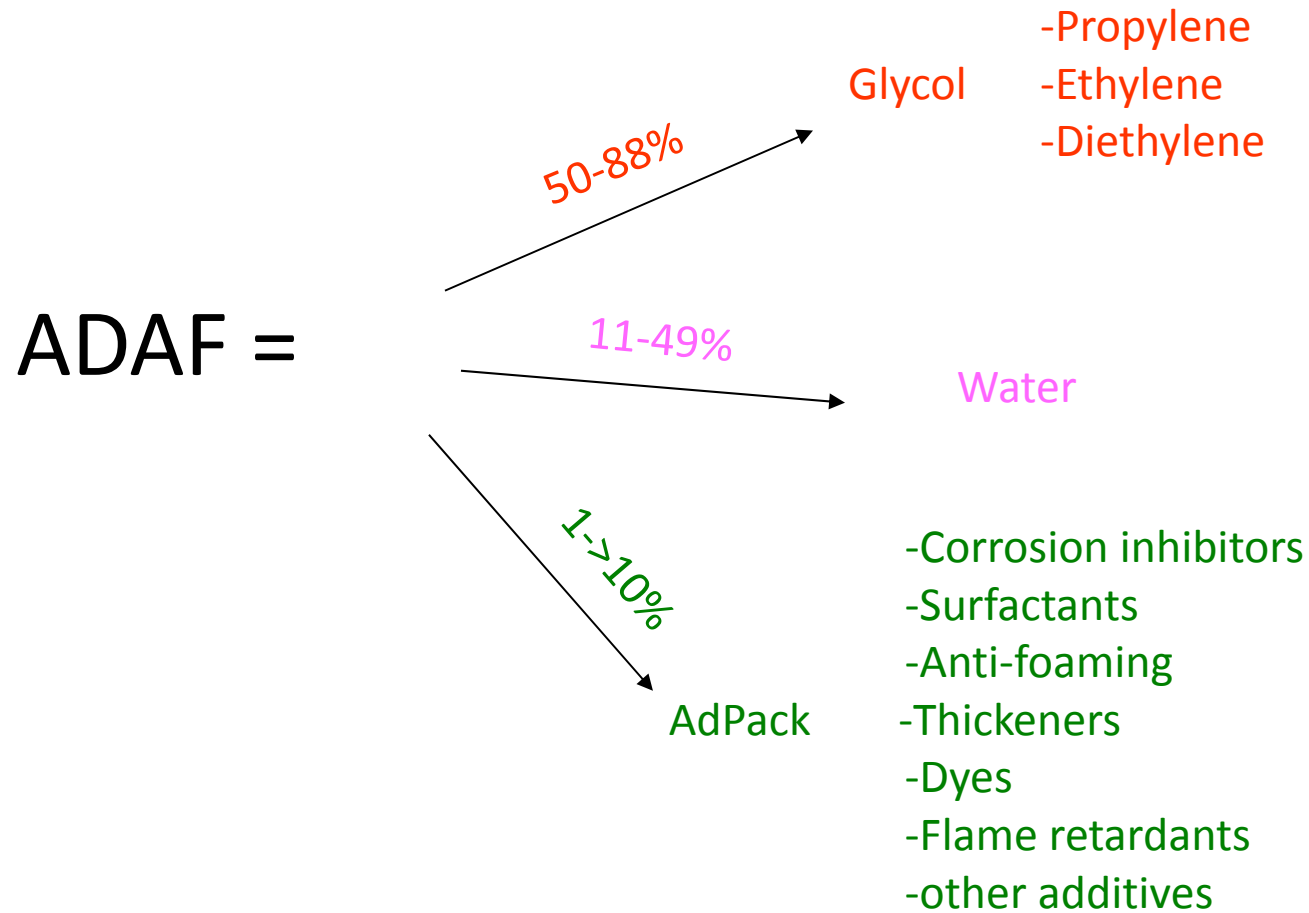


Type I Deicing fluid



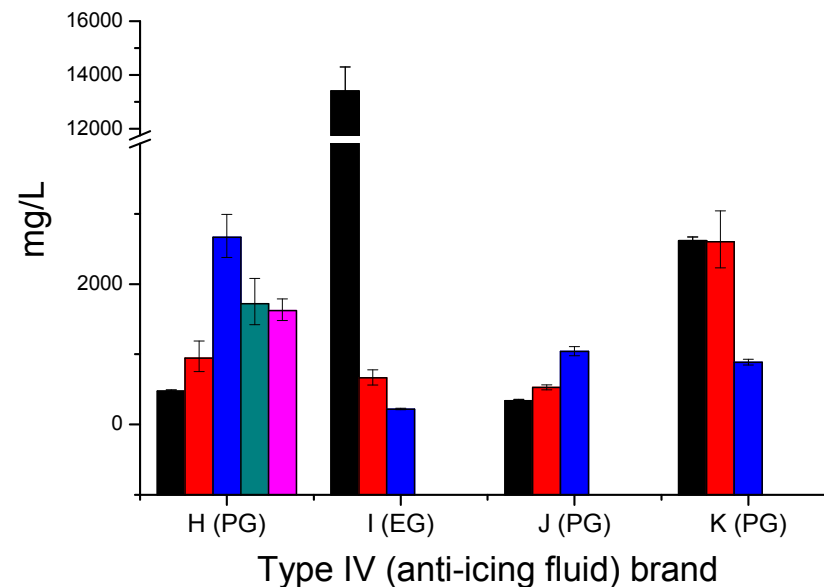
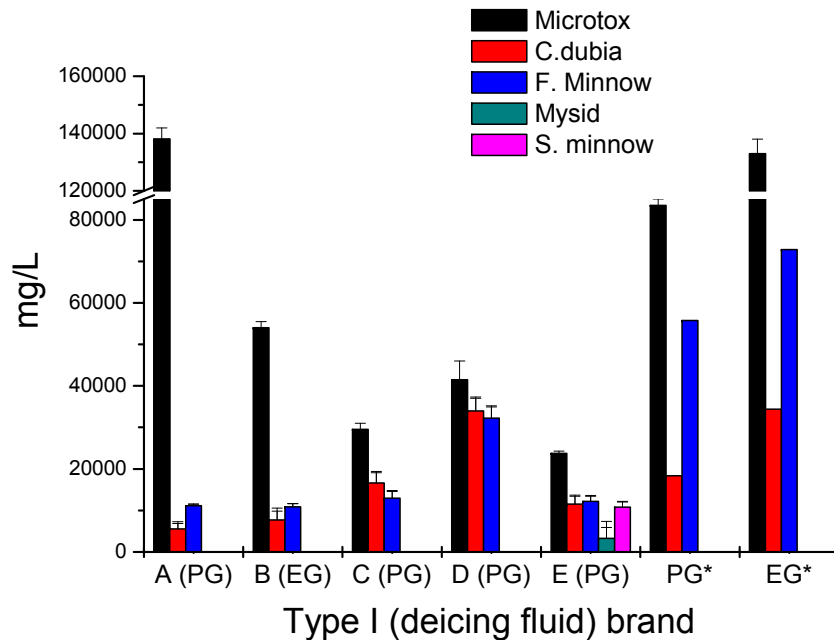
Pavement deicing fluid

Aircraft deicer and anti-icer fluid (ADAF) formulations: What are they?

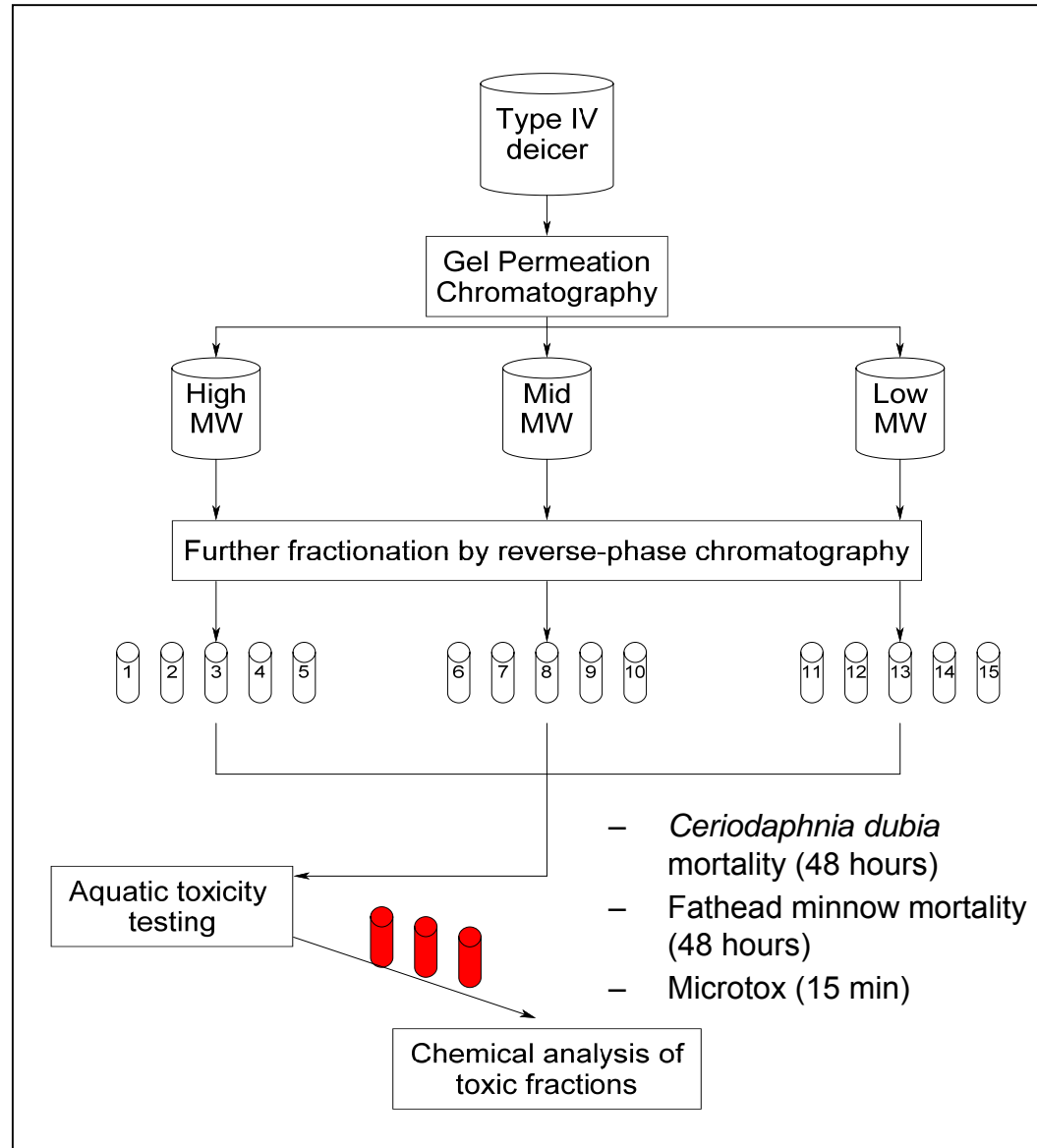


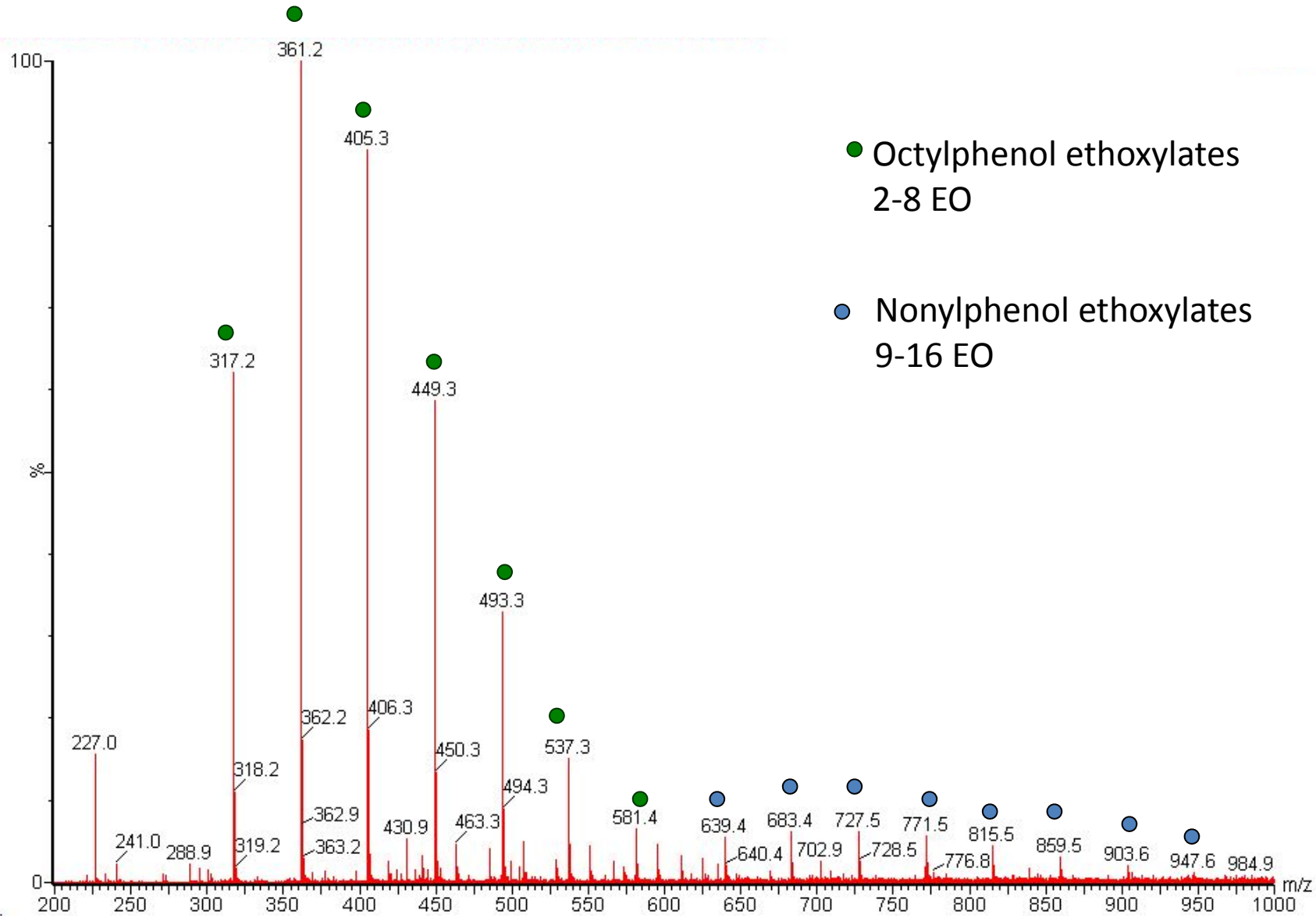
Aircraft anti-icing fluids are more toxic to aquatic organisms than deicing fluids

Toxicities varied within classes, and freezing point depressants did not account for the majority of toxicity in most cases.



Anti-Icer (Type IV) EDA Approach





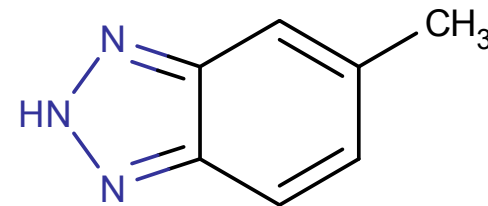
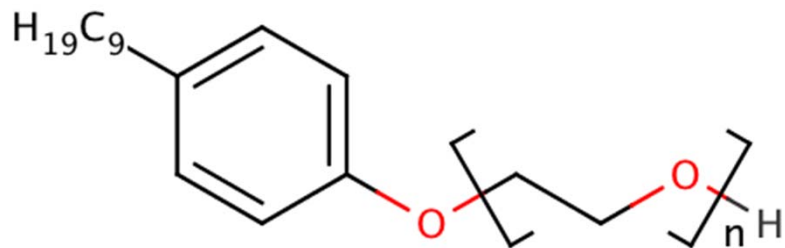
Mass Spectrum, *Chromatogram*, *Peak List*

Summary of EDA findings for Type IV ADAF

- Ethoxylated surfactants were identified in toxic fractions from all 4 fluids
- Tolyltriazole was identified in 1 toxic fraction

Deicer	Fraction #	Compounds Identified in Fraction
Product K	5	OPEO 2-9* NPEO 4-24* C ₁₆ EO 3-6*
	8	methyl-1H-benzotriazole*
	10	OPEO 2-5
Product J	5	C ₁₀ EO 5-16* C ₁₂ EO 2-19* C ₁₄ EO 2-17*
Product I	5	NPEO 2-18* C ₁₃ EO 15-19* C ₁₆ EO 3-13*
Product H	5	C ₁₂ EO 5-19 C ₁₄ EO 4-18
	10	C ₁₂ EO 2-6

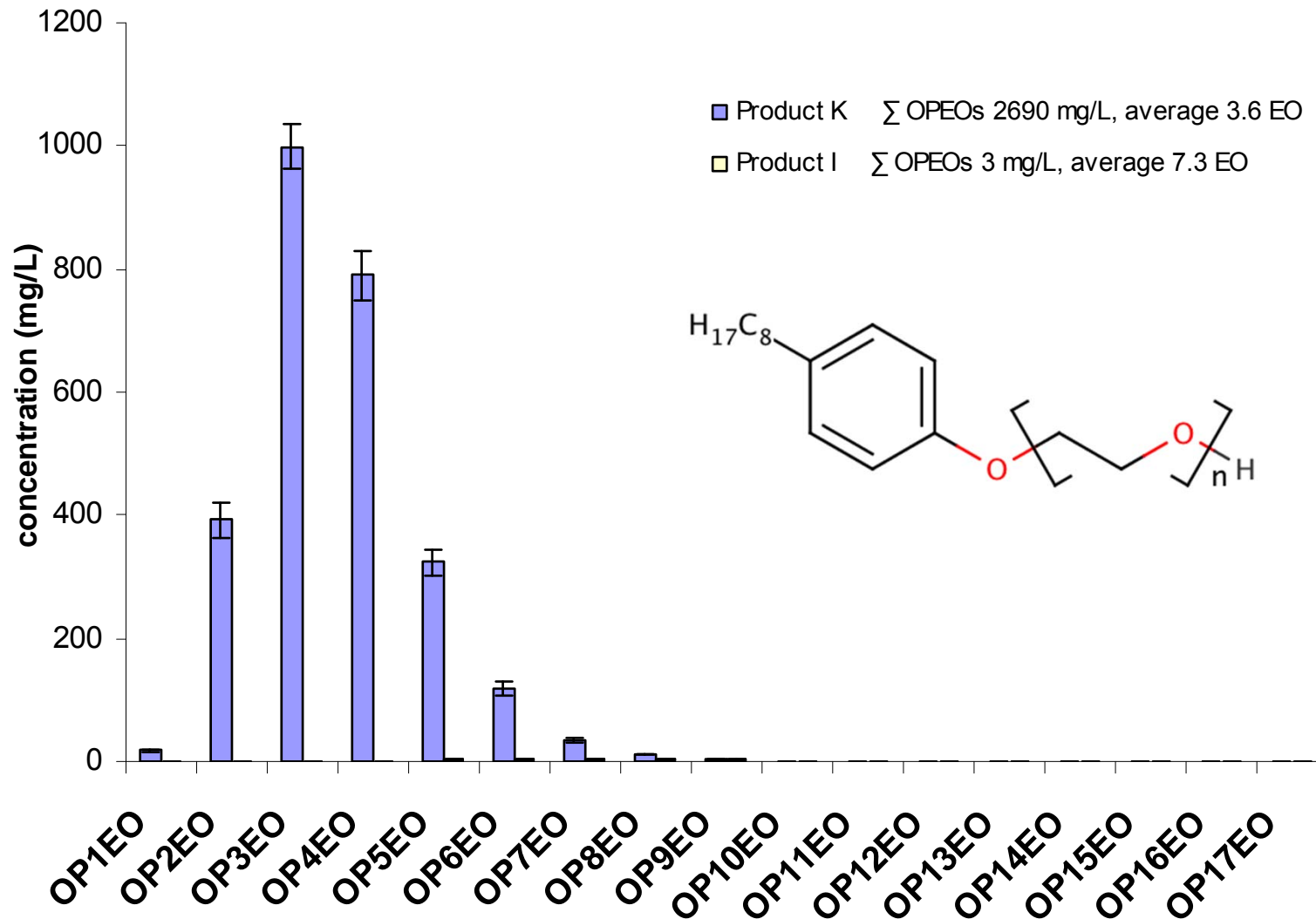
*Compounds identified with high mass accuracy(<5ppm)



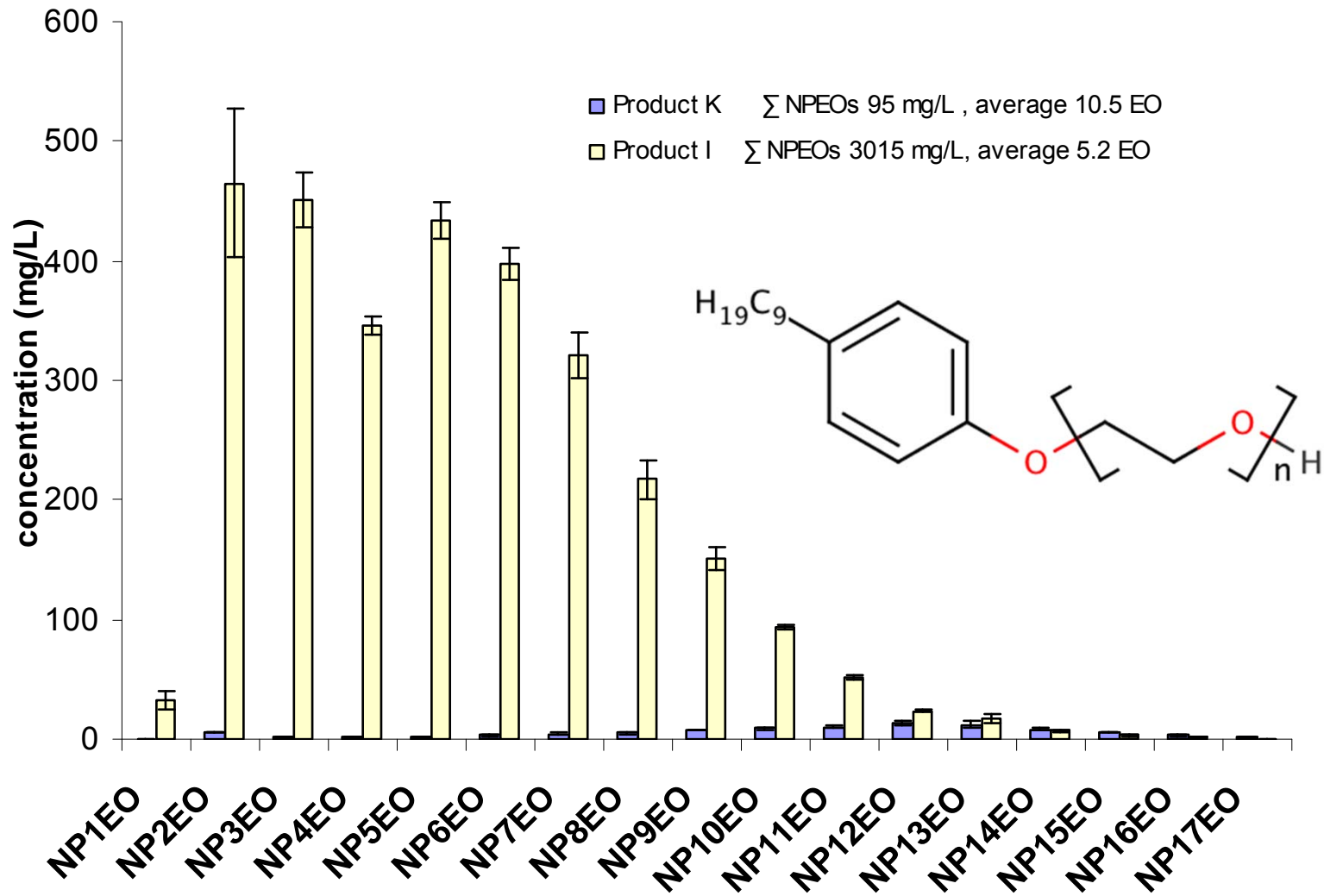
Targeted analysis revealed ethoxylated surfactants in all Type IV deicer fluids

Anti-icing fluid	Surfactants Identified	Estimated average EO number	Relative spectral abundance
Product H	C ₁₀ EO 2-19	6.13	2
	C ₁₁ EO 2-18	8.13	1
	C ₁₃ EO 1-17	6.08	3
	C ₁₅ EO 1-17	9.53	5
	C ₁₆ EO 1-5	2.85	1
Product I	OPEO 5-13	7.33	1
	C ₁₃ EO 1-19*	7.69	0.3
	C ₁₆ EO 1-16*	7.16	2
Product J	C ₁₀ EO 2-16*	7.07	2
	C ₁₁ EO 4-18	9.20	1
	C ₁₅ EO 1-17	9.49	3
	C ₁₆ EO 1-9	3.23	1
Product K	NPEO 4-24*	10.48	7
	C ₁₀ EO 3-18	9.86	1
	C ₁₂ EO 1-17	5.73	1
	C ₁₅ EO 1-17	6.61	3
	C ₁₆ EO 1-8*	4.58	0.1

Octylphenol ethoxylate concentrations in Product K and Product I



Nonylphenol ethoxylate concentrations in Product K and Product I



EDA validation: reformulated “mock” Product I

Formulation	Water	Ethylene Glycol (59%)	Polyacrylic Acid Thickener (2.7 g/L)	NPEO Surfactant (3.0 g/L)	4,5 MeBT Corrosion Inhibitor (0.163 g/L)	EC ₅₀ and LC ₅₀ Values (mg/L)		
						Microtox	<i>C. dubia</i>	Fathead Minnow
Product I	✓	✓	✓	✓	✓	14,063	506	253
Mock #1	✓	✓	✓	—	—	>51,500	>3,000	>3,000
Mock #2	✓	✓	✓	✓	—	29,167	1,576	1,237
Mock #3	✓	✓	✓	—	✓	36,750	>3,140	>3,140
Mock #4	✓	✓	✓	✓	✓	7,737	725	290

Checkmark indicates presence of component in mock formulation.

EDA validation: reformulated “mock” Product K

Formulation	Water	Propylene Glycol (46.6%)	Acid Thickener (2.37 g/L) Polyacrylic	Surfactant (3.0 g/L) OPEO	4,5 MeBT Corrosion Inhibitor (1.16 g/L)	EC ₅₀ and LC ₅₀ Values (mg/L)		
						Microtox	<i>C. dubia</i>	Fathead Minnow
Product K	✓	✓	✓	✓	✓	2,620	2,600	888
Mock #1	✓	✓	✓	—	—	>11,160	>11,160	>11,160
Mock #2	✓	✓	✓	✓	—	2,807	1,452	1,140
Mock #3	✓	✓	✓	—	✓	4,313	>11,500	>11,500
Mock #4	✓	✓	✓	✓	✓	2,250	1,061	576

Checkmark indicates presence of component in mock formulation.

Deicer toxicity: After the dust settled...

- Type IV aircraft anti-icing fluids were much more toxic to aquatic organisms than were Type I deicers.
- Toxic fractions identified through EDA of Type IV deicer fluids contained polyethoxylated surfactants.
- Quantitative analysis of Type IV deicers revealed:
 - Alkylphenol ethoxylate surfactants (3.0 g/L)
 - Alcohol ethoxylate surfactants (0.3 – 0.8 g/L)
 - Benzotriazole-based corrosion inhibitors (~0.1 – 0.8 g/L)
- Toxicity testing of reformulated “mock” Type IV deicer fluids validated EDA results
- “Green” Deicer fluids will require re-examination of surfactant components.

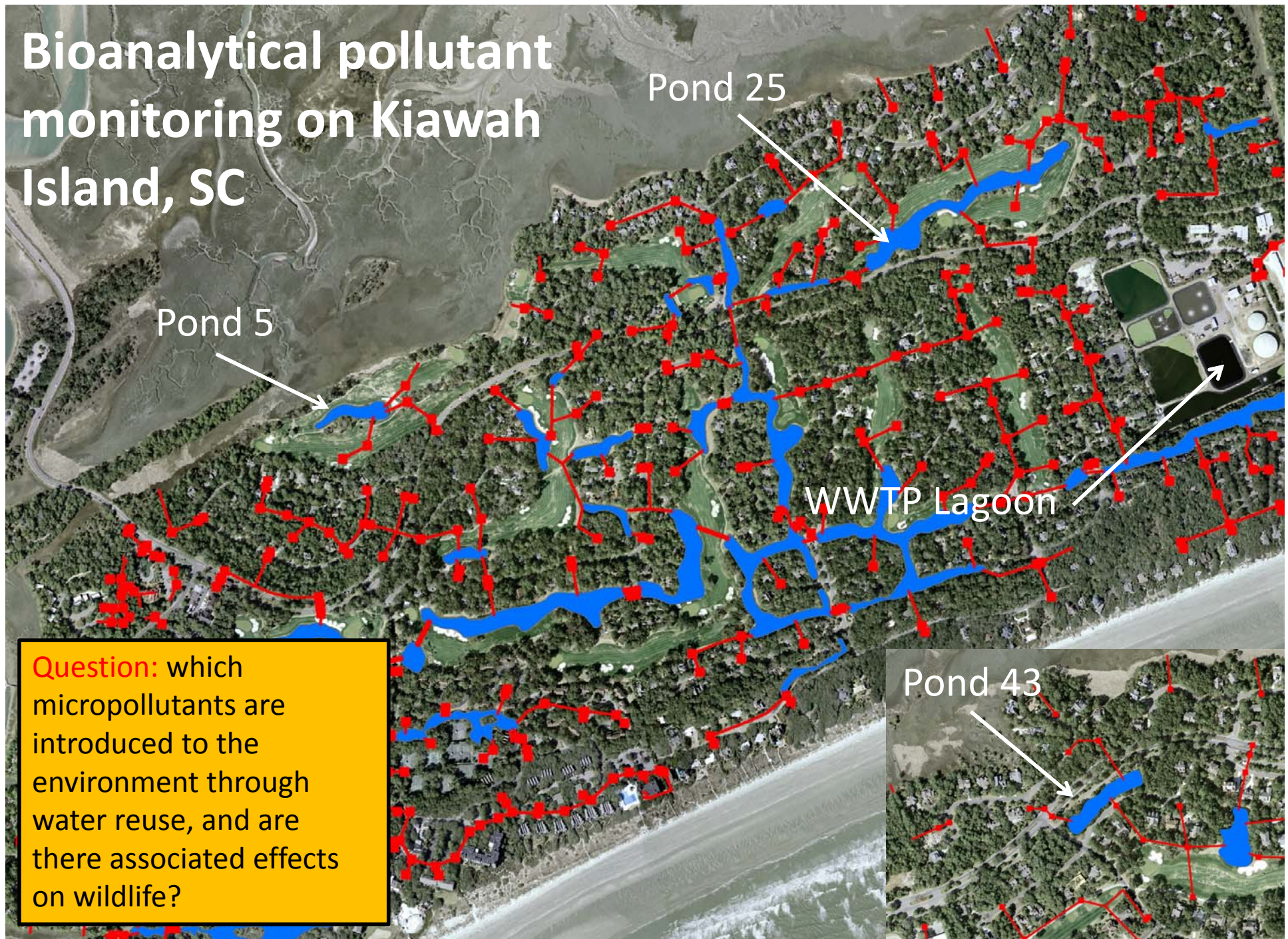
Example approaches from my laboratory:

1. Effects-directed analysis: Identifying toxic components of aircraft deicing/anti-icing fluids (ADAF)
2. **Activity-directed analysis**: Receptor affinity extraction for identifying estrogenic compounds associated with water reuse
3. Fate-directed analysis: Non-targeted analysis of micropollutant fate in wastewater treatment

Water reuse in turf-management: wastewater treatment or contaminant source?



Bioanalytical pollutant monitoring on Kiawah Island, SC



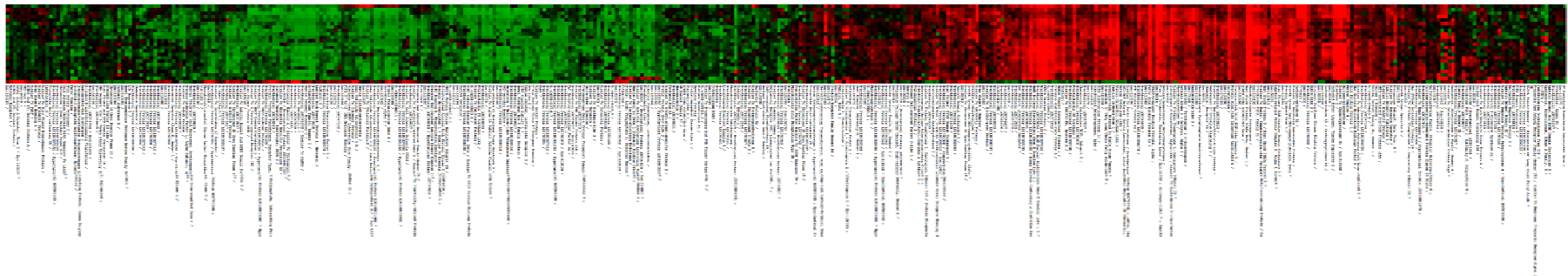
Male Fathead Minnow Exposures



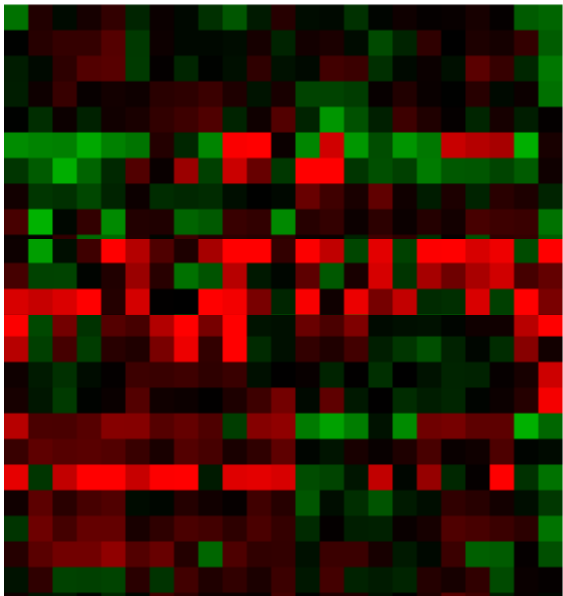
Mini Mobile Units for
“safer” in-situ fish
exposure (Alan
Kolok)



Microarray analysis of hepatic gene expression in Male Fathead Minnows After 1 Week Exposure



P43 C C C C W W W W P25 P25 P43 P43 P43 P5 P5 P5 P5 P25 P25 P43 P25



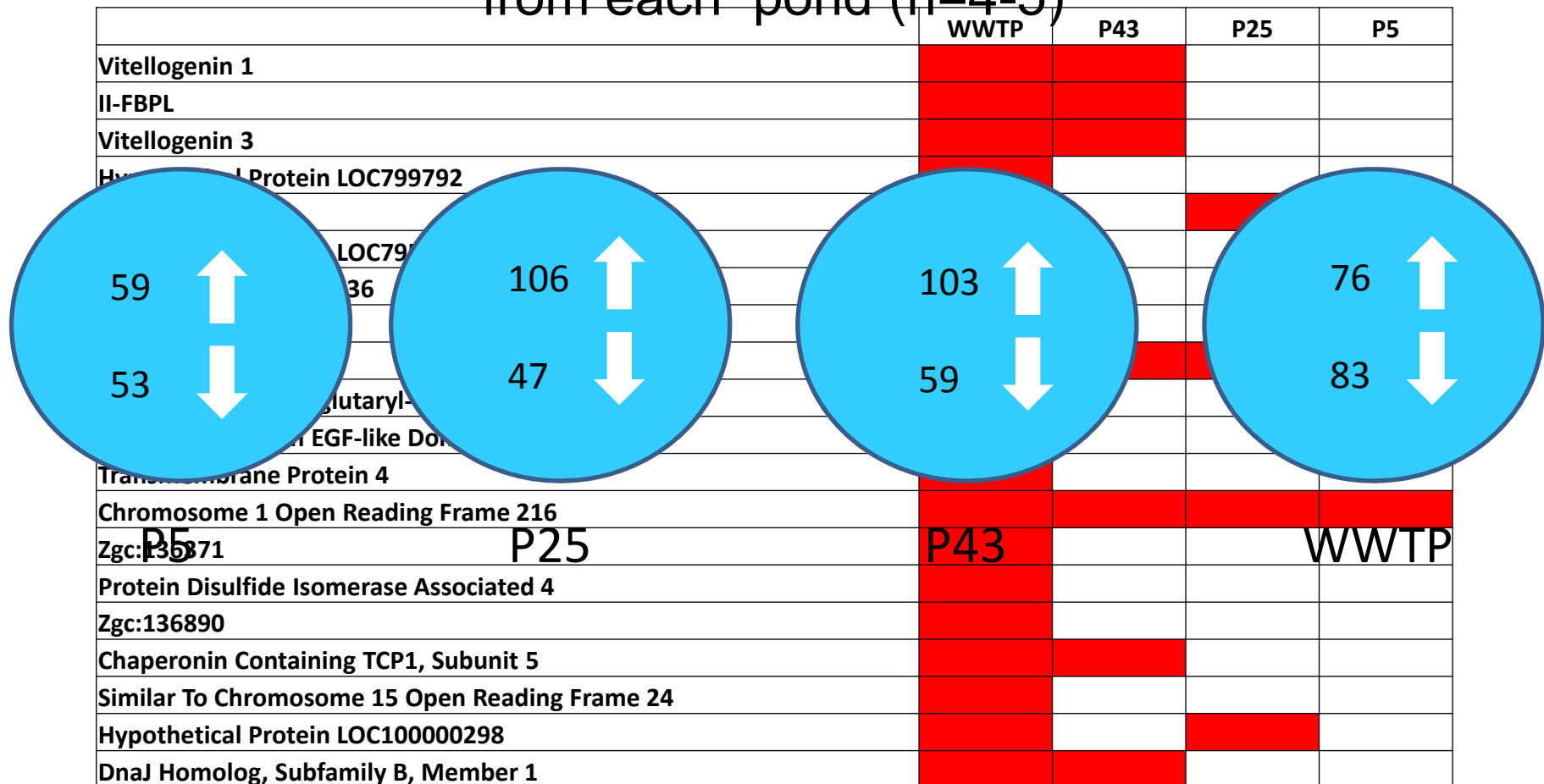
UV Radiation Resistance Associated Gene /
 Zac:110777 /
 Crvstallin, Gamma S1 /
 Hypothetical LOC561667 /
 Ankyrin Repeat And MYND Domain Containing 2
 Angiogenin, Ribonuclease, RNase A Family, 5
 II-FBPL /
 Iodothyronine 5-deiodinase Type III /
 Progesterin And AdipoQ Receptor Family Member
 Angiogenin, Ribonuclease A Family, Member 4
 Similar To Interferon-inducible Protein Gi
 Zac:158870 /
 Similar To Vitellogenin 1 / Vitellogenin 6
 Vitellogenin 3, Phosvitinless /
 Tubulin, Beta 5 /
 Zac:92294 /
 Cytokine Receptor Family Member B4 /
 Zac:112175 /
 Zac:103599 /
 Cbp/p300-interacting Transactivator, With C
 Zac:103418 /
 Serine Hydroxymethyltransferase 1 / Hypothe
 Tripartite Motif-containing 25 /

15K fathead minnow microarray
 ~ 3700 genes significantly
 expressed
 differently between ponds
 ~ 400 genes significantly
 expressed
 differently between ponds > 2X

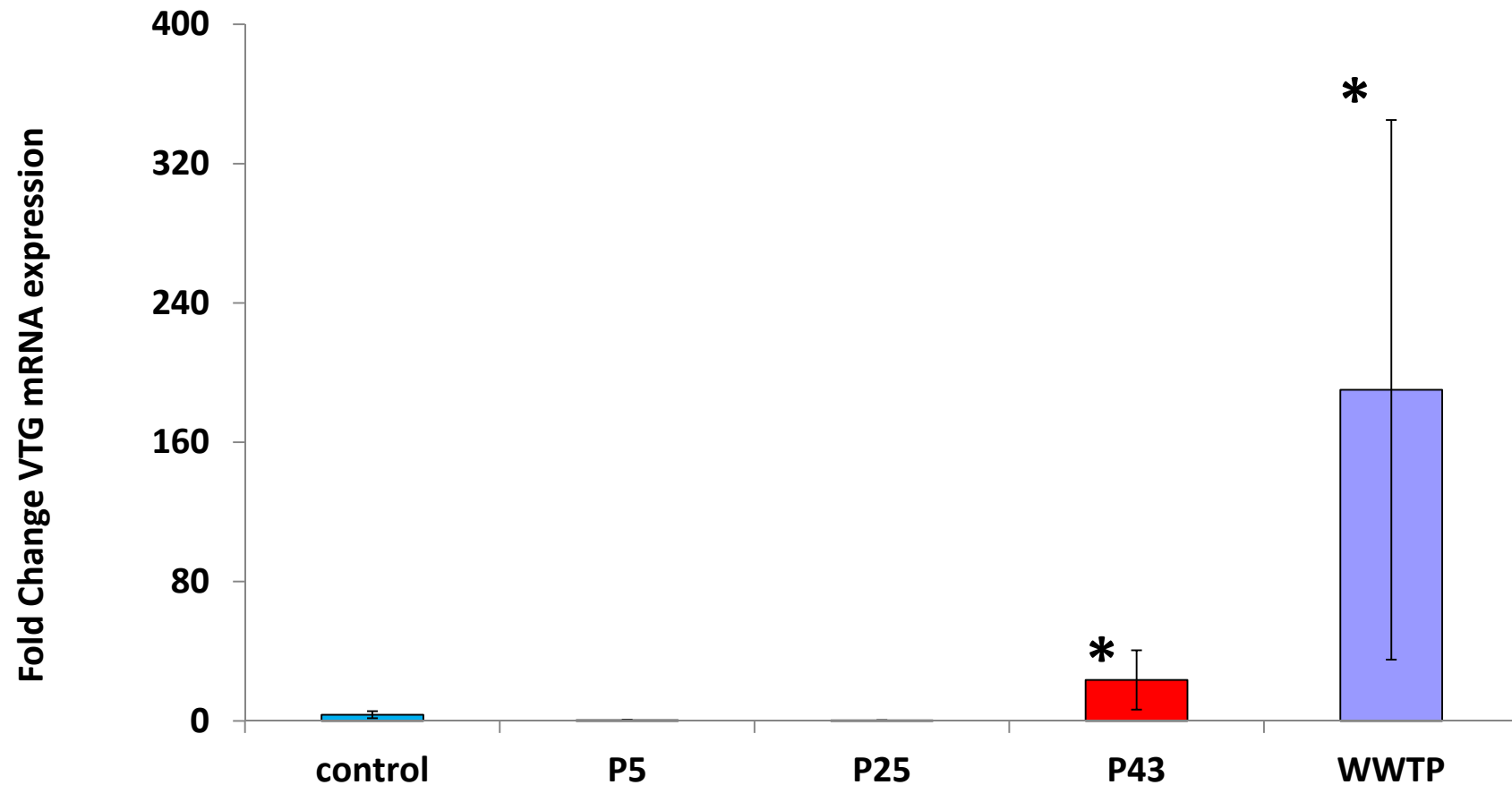
■ Upregulated
■ Downregulated

Microarray analysis of hepatic gene expression in Male Fathead Minnows After 1 Week Exposure

Number of genes significantly up- and down-regulated $\geq 2X$ in fish from each pond (n=4-5)

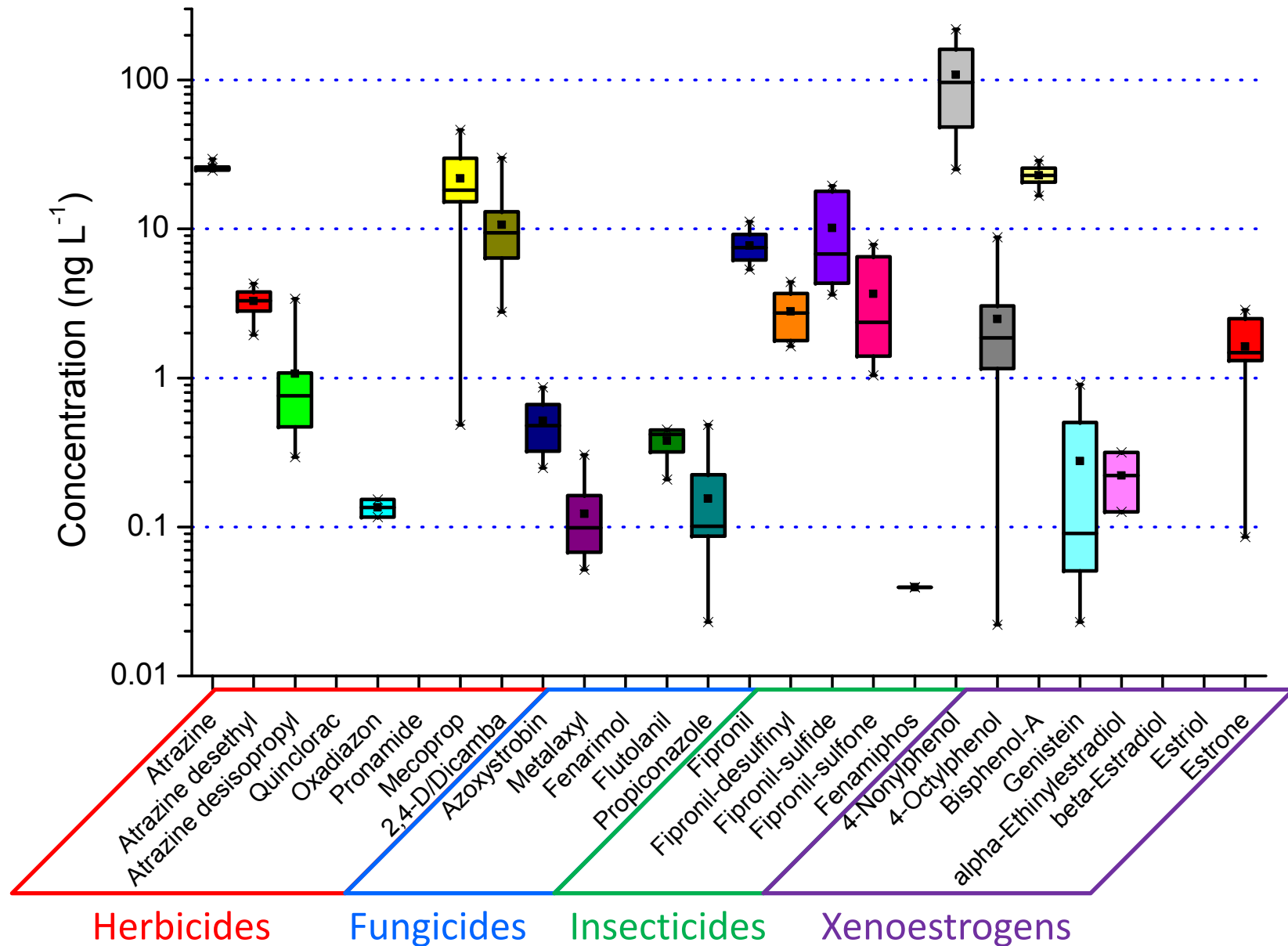


Hepatic Vitellogenin mRNA Expression in Male Fathead Minnows After 1 Week Exposure



* Indicate statistically significant values compared to control using one-way ANOVA ($p < 0.05$)

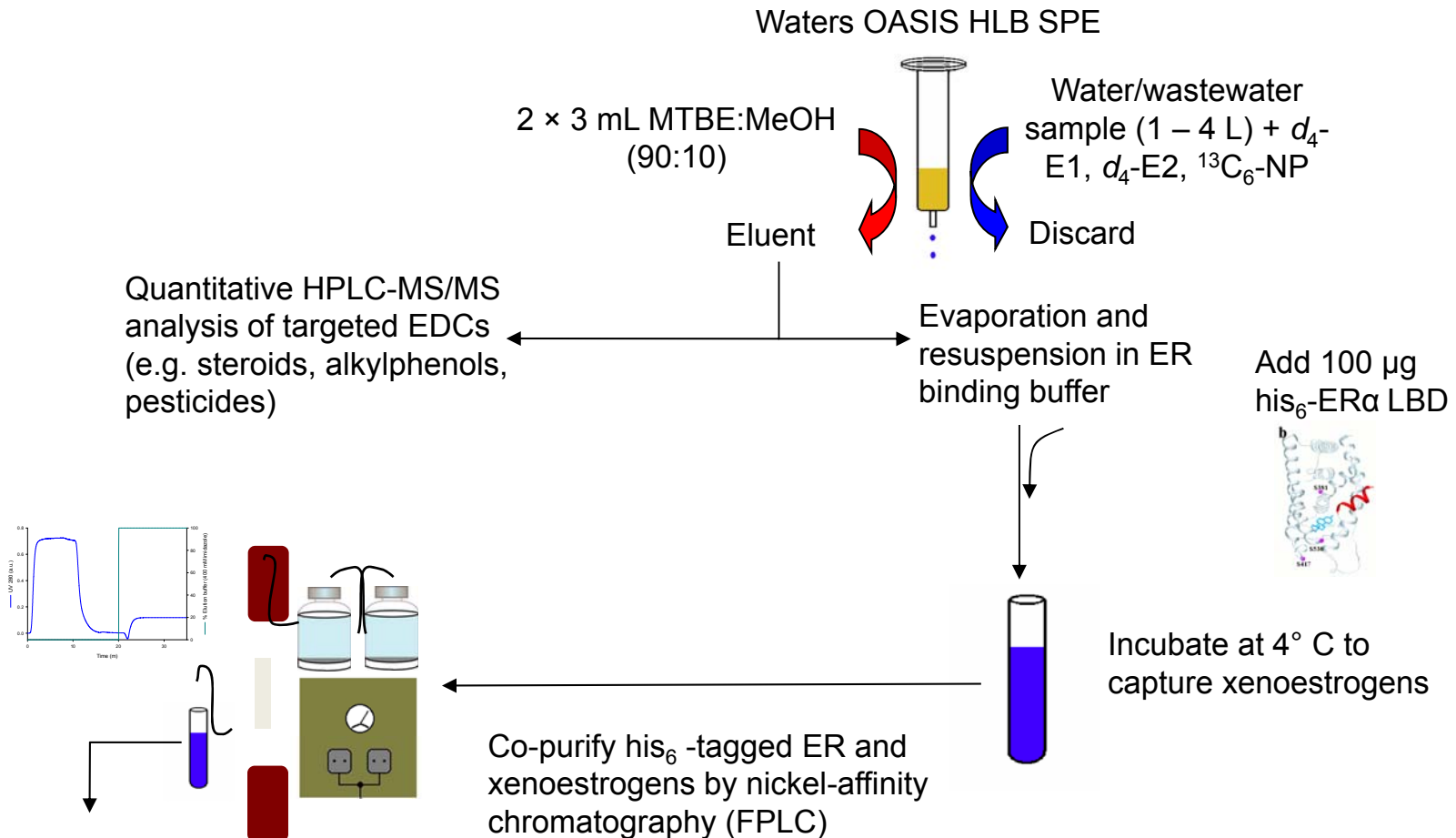
Micropollutants in WWTP Lagoon



Yes, but what ELSE might be contributing to estrogenicity in the reclaimed water??

- **Receptor affinity extraction:** Similar in concept to immunoaffinity chromatography – relies on high specificity/selectivity molecular interaction to isolate target analytes from a mixture prior to analysis
- Recombinant protein engineering:
 - ER α ligand binding domain triple Cys \rightarrow Ser mutant
 - Fusion of thioredoxin to ER enhances solubility
 - His₆ tag allows subsequent purification
- Proteins are cloned, expressed in bacterial vectors, and purified chromatographically

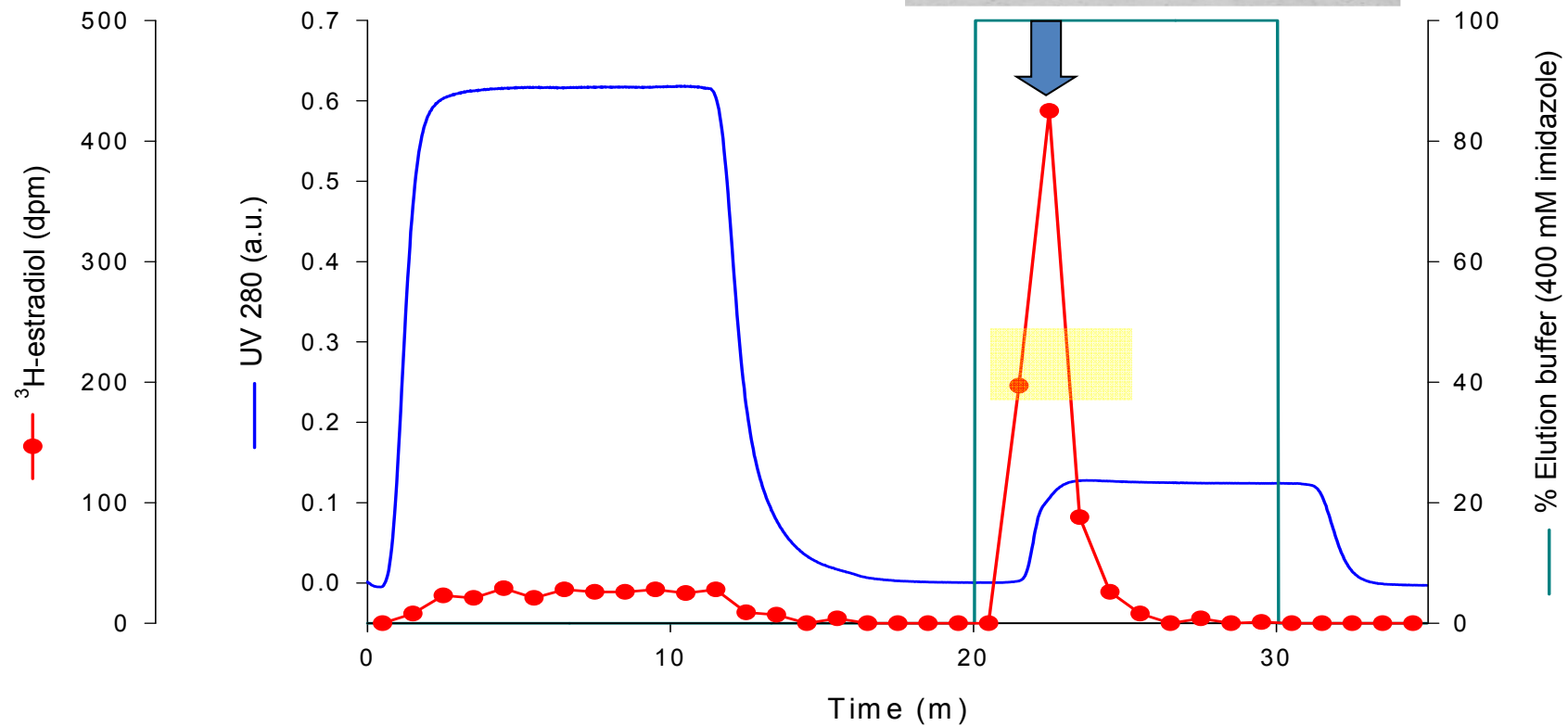
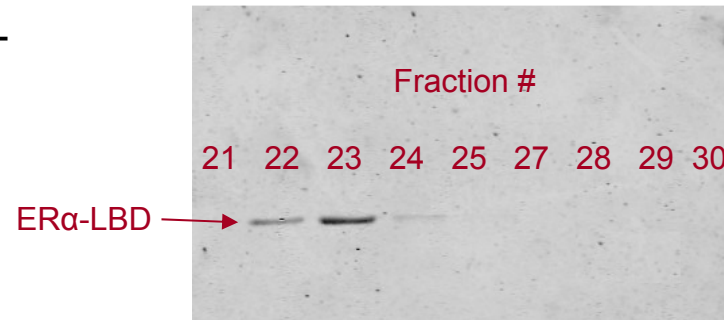
Estrogen receptor-affinity isolation for activity-directed analysis



Analysis of receptor-active EDCs in eluent by UHPLC-Orbitrap MS/MS or triple-quadrupole MS

Purification of ER-bound xenoestrogens from wastewater

ER α -LBD and radioligand tracer (^3H -17 β -estradiol) are selectively retained from wastewater extract by Ni $^{2+}$ agarose, and can be co-eluted with 400 mM imidazole



HRMS-enabled screening using published databases and curated/literature MS/MS data

Question: which compounds within a curated list are present in this sample?

Three databases →

1. 745 High production volume pharmaceuticals predicted to be persistent and or bioaccumulative (Howard & Muir, 2011)
2. 610 High production volume chemicals in commerce predicted to be persistent and or bioaccumulative (Howard & Muir, 2010)
3. 1006 pesticides & pharmaceuticals (ThermoFisher Scientific Environmental & Food Safety database)

ENVIRONMENTAL
Science & Technology

ARTICLE

pubs.acs.org/est

Identifying New Persistent and Bioaccumulative Organics Among Chemicals in Commerce II: Pharmaceuticals

Philip H. Howard^{*†} and Derek C. G. Muir[‡]

[†]SRC, Inc. Chemical, Biological, and Environmental Center (CBEC), 7502 Round Pond Road, North Syracuse, New York, and

[‡]Aquatic Ecosystem Protection Research Division, Environment Canada, 867 Lakeshore Road, Burlington, Ontario

Identifying New Persistent and Bioaccumulative Organics Among Chemicals in Commerce

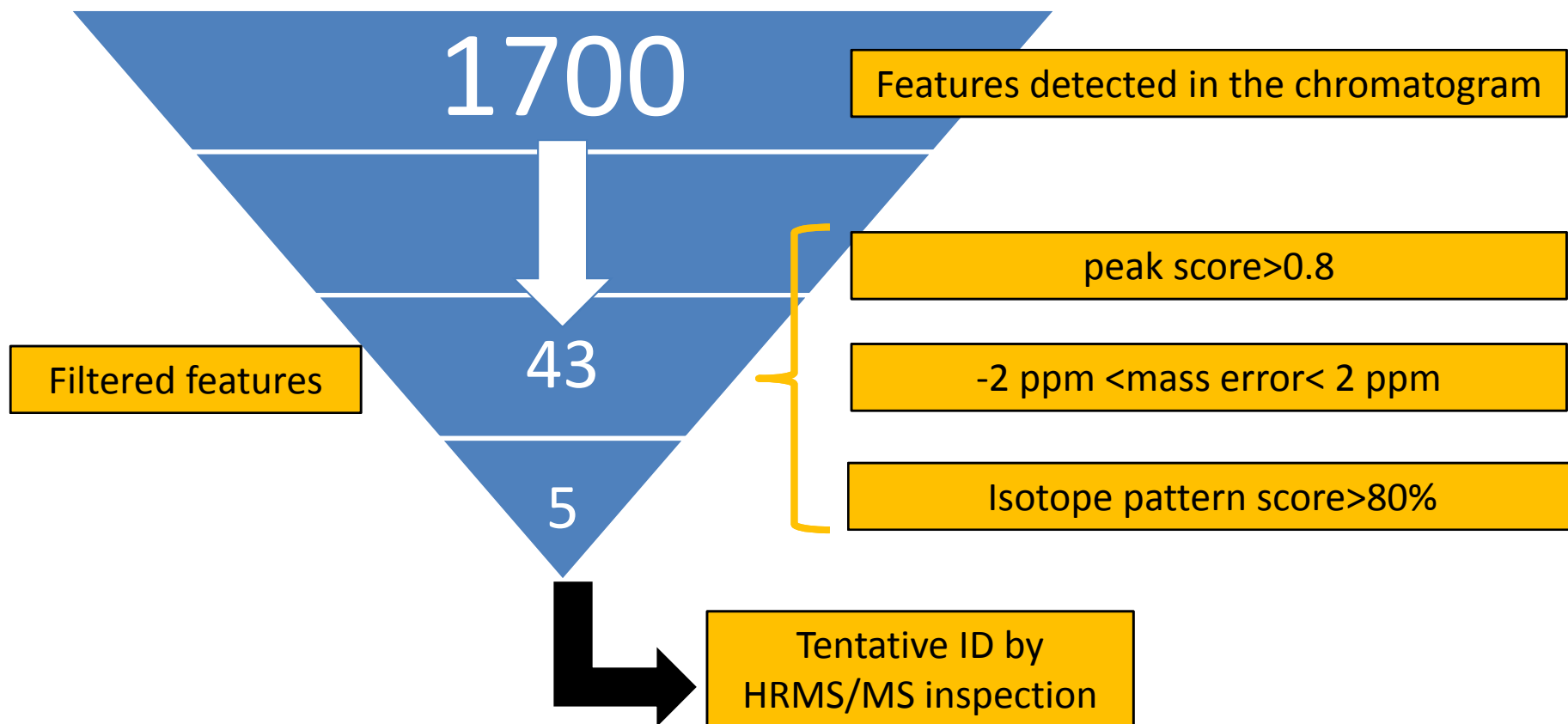
PHILIP H. HOWARD^{*†} AND
DEREK C. G. MUIR[‡]

SRC, Environmental Science Center, 6502 Round Pond Road, North Syracuse, New York, and Aquatic Ecosystem Protection Research Division, Environment Canada, 867 Lakeshore Road, Burlington, Ontario

Received November 6, 2009. Revised manuscript received January 19, 2010. Accepted January 22, 2010.

Thermo
SCIENTIFIC

Data filtering and tentative compound identification



Estrogen-Like Activity of Perfluoroalkyl Acids *In Vivo* and Interaction with Human and Rainbow Trout Estrogen Receptors *In Vitro*Abby D. Benninghoff,*¹ William H. Bisson,[†] Daniel C. Koch,[‡] David J. Ehresman,[§] Siva K. Kolluri,[‡] and David E. Williams[‡]^{*}Department of Animal, Dairy, and Veterinary Sciences and the Graduate Program in Toxicology, Utah State University, 4815 Old Main Hill, Logan, Utah 84322-4815; [†]Pharmaceutical Biochemistry Group, School of Pharmaceutical Sciences, University of Geneva, Geneva, Switzerland; [‡]Department of Environmental and Molecular Toxicology, Oregon State University, 1007 Agricultural and Life Sciences Building, Corvallis, Oregon 97331; and [§]3M Medical Department, Corporate Toxicology, 3M Center 220-2E-02, Saint Paul, Minnesota 55133¹To whom correspondence should be addressed at Animal, Dairy, and Veterinary Sciences, Utah State University, 4815 Old Main Hill, Logan, UT 84322-4815. Fax: (435) 979-2118. E-mail: abby.benninghoff@usu.edu.

Received August 26, 2010; accepted December 06, 2010

The objectives of this study were to determine the structural characteristics of perfluoroalkyl acids (PFAAs) that confer estrogen-like activity *in vivo* using juvenile rainbow trout (*Oncorhynchus mykiss*) as an animal model and to determine whether these chemicals interact directly with the estrogen receptor (ER) using *in vitro* and *in silico* species comparison approaches. Perfluorooctanoic (PFOA), perfluorononanoic (PFNA), perfluorodecanoic (PFDA), and perfluoroundecanoic (PFUnDA) acids were all potent inducers of the estrogen-responsive biomarker protein vitellogenin (Vtg) *in vivo*, although at fairly high dietary exposures. A structure-activity relationship for PFAAs was observed, where eight to ten fluorinated carbons and a carboxylic acid end group were optimal for maximal Vtg induction. These *in vivo* findings were corroborated by *in vitro* mechanistic assays for trout and human ER. All PFAAs tested weakly bound to trout liver ER with half maximal inhibitory concentration (IC₅₀) values of 15.2–289 μM. Additionally, PFOA, PFNA, PFDA, PFUnDA, and perfluorooctane sulfonate (PFOS) significantly enhanced human ERα-dependent transcriptional activation at concentrations ranging from 10–1000 nM. Finally, we employed an *in silico* computational model based upon the crystal structure for the human ERα ligand-binding domain complexed with E2 to structurally investigate binding of these putative ligands to human, mouse, and trout ERα. PFOA, PFNA, PFDA, and PFOS all efficiently docked with ERα from different species and formed a hydrogen bond at residue Arg394/398/407 (human/mouse/trout) in a manner similar to the environmental estrogens bisphenol A and nonylphenol. Overall, these data support the contention that several PFAAs are weak environmental xenoestrogens of potential concern.

Key Words: perfluoroalkyl acid; estrogen; perfluorooctanoic acid; perfluorooctane sulfonate; vitellogenin; molecular docking.

The widespread industrial and commercial use of polyfluorinated chemicals (PFCs) as surfactants and surface protectors for paper and textile coatings, polishes, food

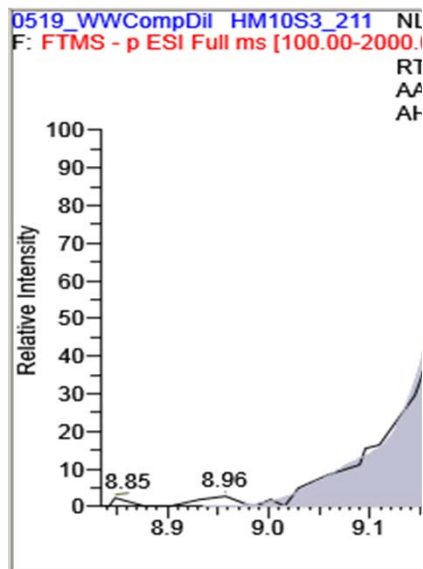
packaging, and fire-retardant foams has led to the pervasive presence of these chemicals in the environment, wildlife, and humans (see reviews by Calafat *et al.*, 2007; Houde *et al.*, 2006). The general structure of PFCs resembles that of fatty acids in that each compound has a hydrophobic polyfluorinated carbon tail of varying length and a functional end group, which provides the basis for classification (Fig. 1). Perfluoroalkyl acids (PFAAs) and fluorotelomers comprise the two major structural groups of PFCs. Human blood levels of perfluorooctanoic acid (PFOA) and perfluorooctane sulfonate (PFOS), the two most commonly studied PFCs, are about 4 and 20 ppb, respectively, although national survey data suggest that these levels have decreased in recent years (Calafat *et al.*, 2007). Levels of PFOS and PFOA in wildlife tend to be higher, in the range of tens to thousands ppb (Kannan *et al.*, 2002). Other PFCs commonly detected in biological samples include perfluorononanoic acid (PFNA), perfluorodecanoic acid (PFDA), perfluoroundecanoic acid (PFUnDA), and perfluorododecanoic acid (PFDoDA).

The toxicology and toxicokinetics of PFOA, as an example PFC, have been thoroughly reviewed by Kennedy *et al.* (2004). PFOA does not accumulate in fatty tissues because of its dual lipophobic and hydrophobic chemical properties but instead binds to blood proteins and is distributed primarily to liver, plasma, and kidney. The measured biological half-life of PFOA varies among species ranging from hours in female rat to days in dogs or rainbow trout (Hanhijarvi *et al.*, 1988; Martin *et al.*, 2003). The estimated half-life of PFOA in humans is nearly 4 years, pointing to a lower capacity for elimination of the compound compared with other species (Olsen *et al.*, 2007). Finally, PFOA is not metabolized or defluorinated *in vivo*, although some fluorotelomer compounds may be metabolized to PFAAs in rodents (Martin *et al.*, 2005; Nabb *et al.*, 2007).

PFOA and other PFCs are members of a large group of chemicals called peroxisome proliferators (PPs), which

Peak ID

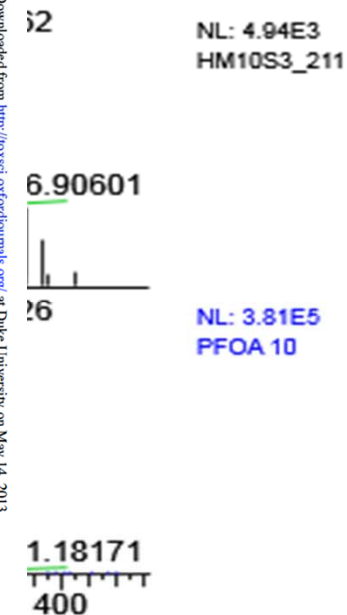
Peak 211(-412.966819.21)



Identification of perfluorooctanoic acid (PFOA)

MS

core

Downloaded from <http://toxsci.oxfordjournals.org/> at Duke University on May 14, 2013

Xenoestrogen extracts

SHORT COMMUNICATION

Efavirenz directly modulates the oestrogen receptor and induces breast cancer cell growth

MJ Sikora,¹ JM Rae,^{1,2} MD Johnson³ and Z Desta⁴

¹Department of Pharmacology, ²Department of Internal Medicine, University of Michigan Medical Center, Ann Arbor, MI, USA, ³Lombardi Cancer Center and Department of Oncology, Georgetown University Medical Center, Washington, DC, USA and ⁴Division of Clinical Pharmacology, Department of Medicine, Indiana University, Indianapolis, IN, USA

Objectives

Efavirenz-based HIV therapy is associated with breast hypertrophy and gynaecomastia. Here, we tested the hypothesis that efavirenz induces gynaecomastia through direct binding and modulation of the oestrogen receptor (ER).

Methods

To determine the effect of efavirenz on growth, the oestrogen-dependent, ER-positive breast cancer cell lines MCF-7, T47D and ZR-75-1 were treated with efavirenz under oestrogen-free conditions in the presence or absence of the anti-oestrogen ICI 182,780. Cells treated with 17 β -oestradiol in the absence or presence of ICI 182,780 served as positive and negative controls, respectively. Cellular growth was assayed using the crystal violet staining method and an *in vitro* receptor binding assay was used to measure the ER binding affinity of efavirenz.

Results

Efavirenz induced growth in MCF-7 cells with an estimated effective concentration for half-maximal growth (EC₅₀) of 15.7 μ M. This growth was reversed by ICI 182,780. Further, efavirenz binds directly to the ER [inhibitory concentration for half maximal binding (IC₅₀) of ~ 52 μ M] at a roughly 1000-fold higher concentration than observed with 17 β -oestradiol.

Conclusions

Our data suggest that efavirenz-induced gynaecomastia may be caused, at least in part, by drug-induced ER activation in breast tissues.

Keywords: efavirenz, gynaecomastia, highly active antiretroviral therapy, oestrogen receptor, oestrogens

Accepted 12 January 2010

Introduction

The introduction of highly active antiretroviral therapy (HAART) multi-drug combination regimens has considerably improved the prognosis of patients infected with HIV by reducing AIDS-related morbidity and mortality [1]. However, chronic treatment with these regimens is associated with multiple adverse effects, nonadherence and eventually therapy failure [2]. Treatment regimens containing the nonnucleoside reverse transcriptase inhibitor efavirenz are preferred in treatment-naïve patients

and are widely used in other settings [3]. While efavirenz is generally well tolerated, concentration-dependent side effects that impact drug adherence and promote resistance have been documented [4]. Common adverse effects of efavirenz include central nervous system symptoms, occurring in up to 50% of patients [5], but other less common adverse effects have also been reported. An increasing number of reports suggest that the use of HAART, in particular efavirenz-based therapy, is associated with breast hypertrophy or gynaecomastia [6-11]. While mechanisms underlying efavirenz-induced gynaecomastia are not well understood, a number of hypotheses exist, including a direct oestrogenic effect, induction of an immune response, or altered steroid hormone metabolism by cytochrome P450 enzymes. To our knowledge, none of

Correspondence: Dr Zeruesenay Desta, Division of Clinical Pharmacology, Department of Medicine, Indiana University School of Medicine, 1001 West 10th Street, WD Myers Bldg., W7123, Indianapolis, IN 46202, USA. E-mail: zdesta@iupui.edu

tor affinity screening

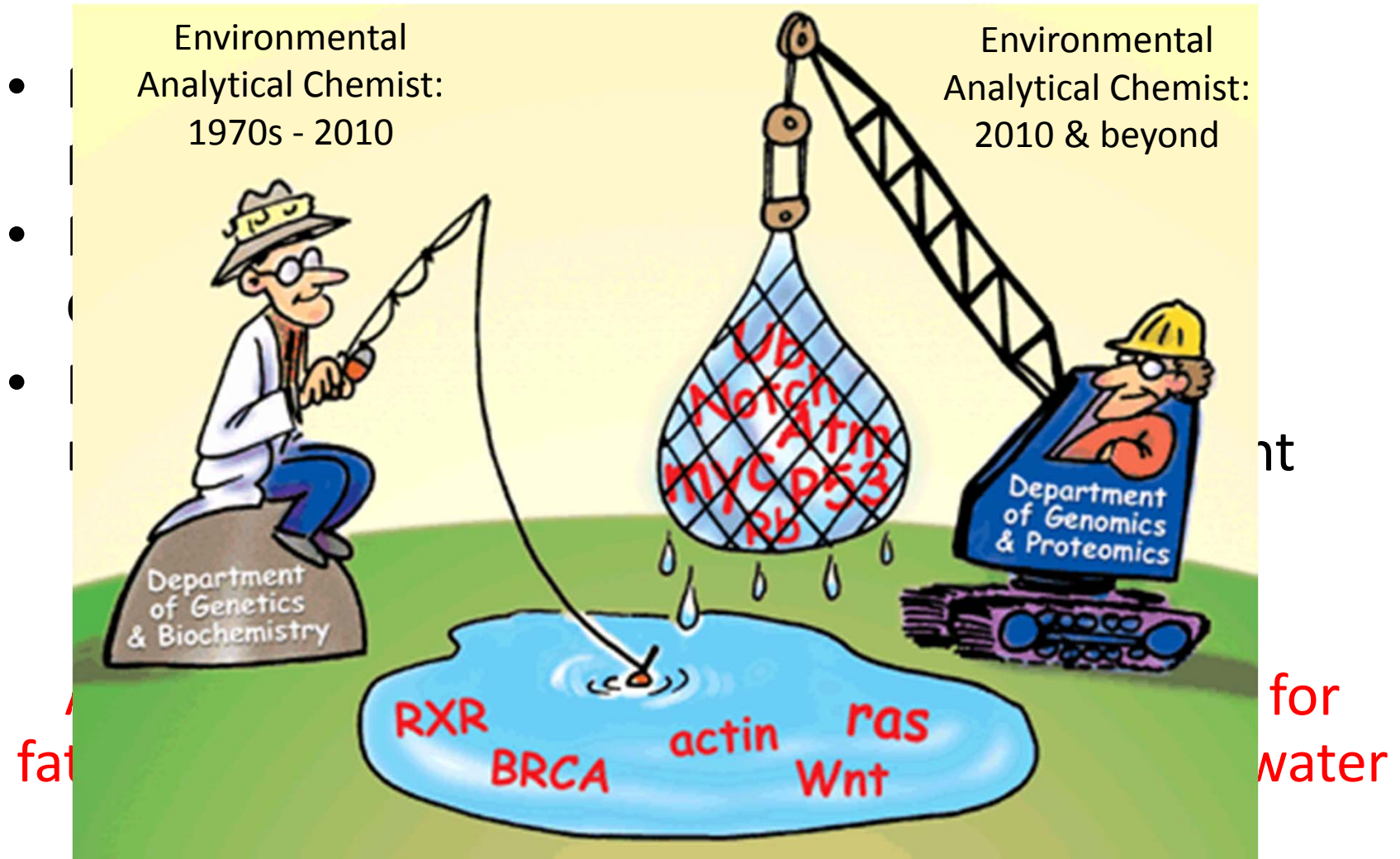
Name	Class
17 β -estradiol	Endogenous steroid
Estrone	Endogenous steroid
Celecoxib	NSAID Pharmaceutical
PFOA	Fluorinated surfactant
Efavirenz	Antiviral pharmaceutical

Pattern	Confirmed & Quantified?
00	0.70 ng/L
05	11.5 ng/L
00	no
00	no
00	no

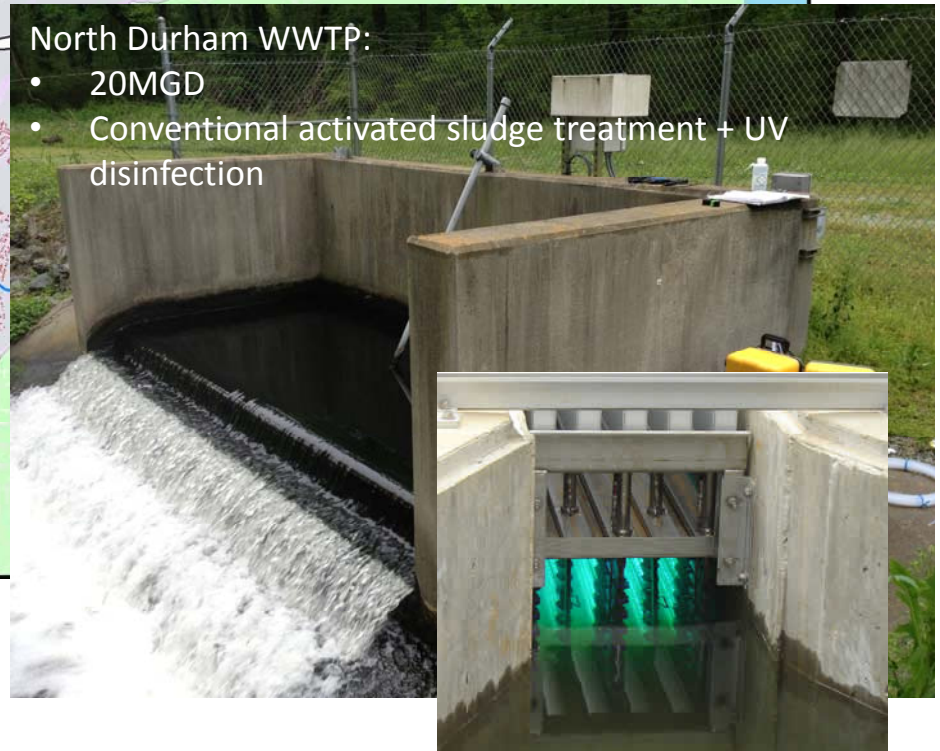
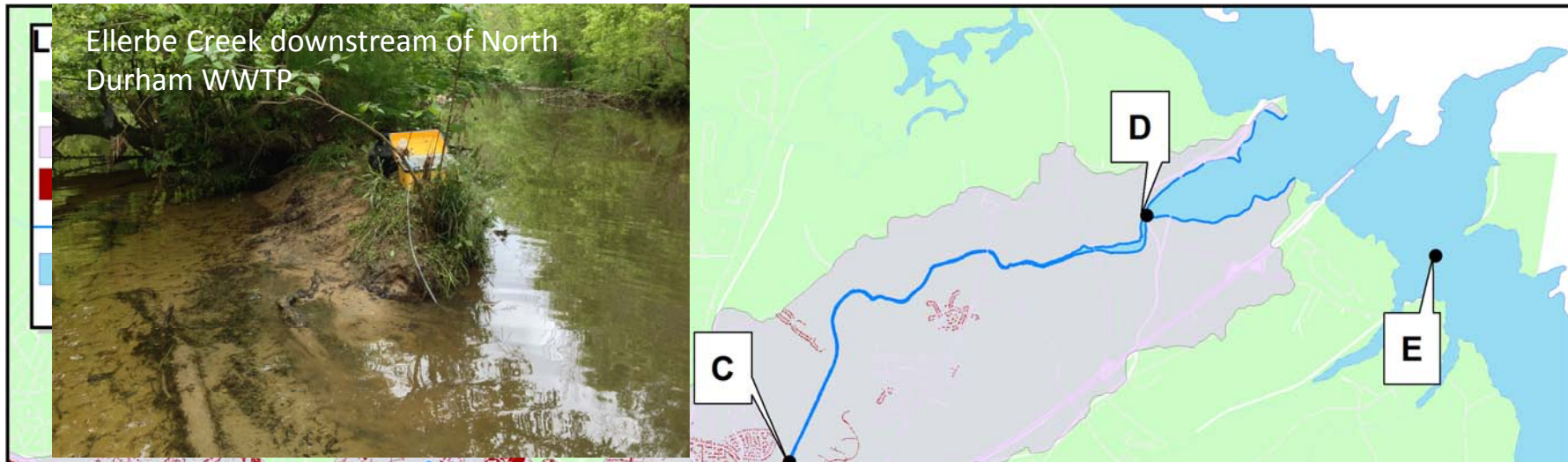
Example approaches from my laboratory:

1. Effects-directed analysis: Identifying toxic components of aircraft deicing/anti-icing fluids (ADAF)
2. Activity-directed analysis: Receptor affinity extraction for identifying estrogenic compounds associated with water reuse
3. Fate-directed analysis: Non-targeted analysis of micropollutant fate in wastewater treatment

Wastewater is a significant source of emerging contaminants to the aquatic environment



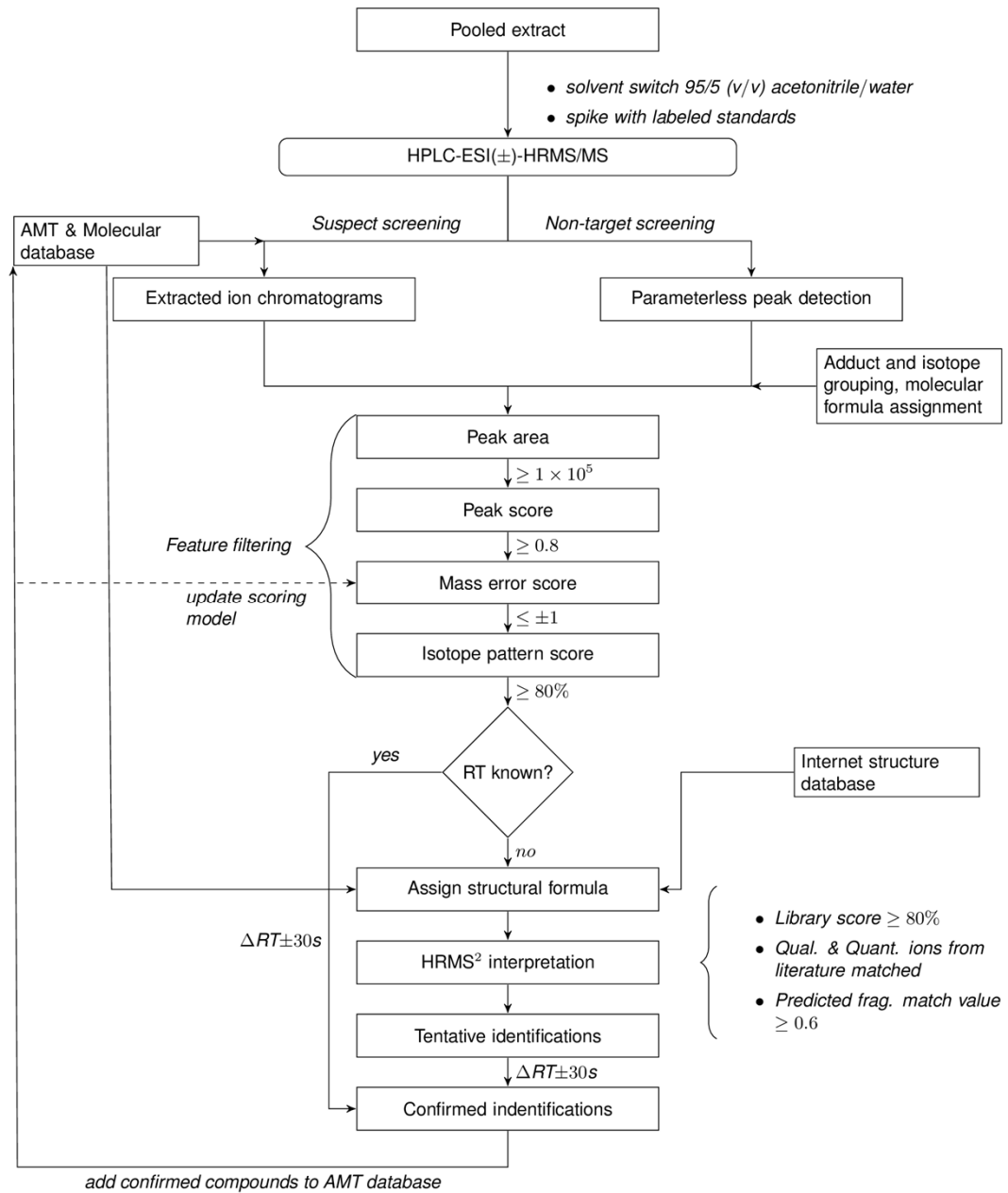
Study site and sampling



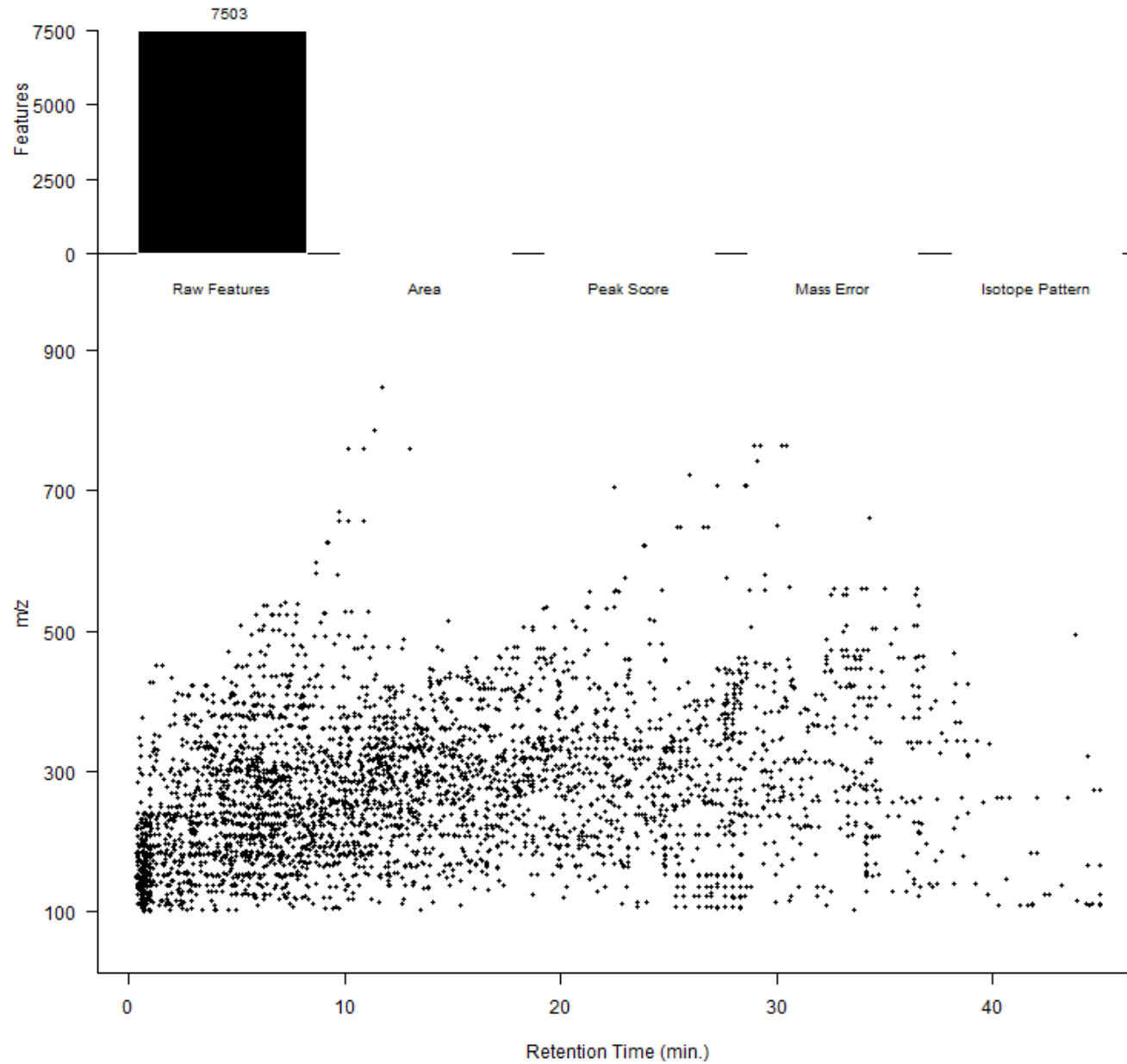
Data analysis workflow

Two suspect databases:

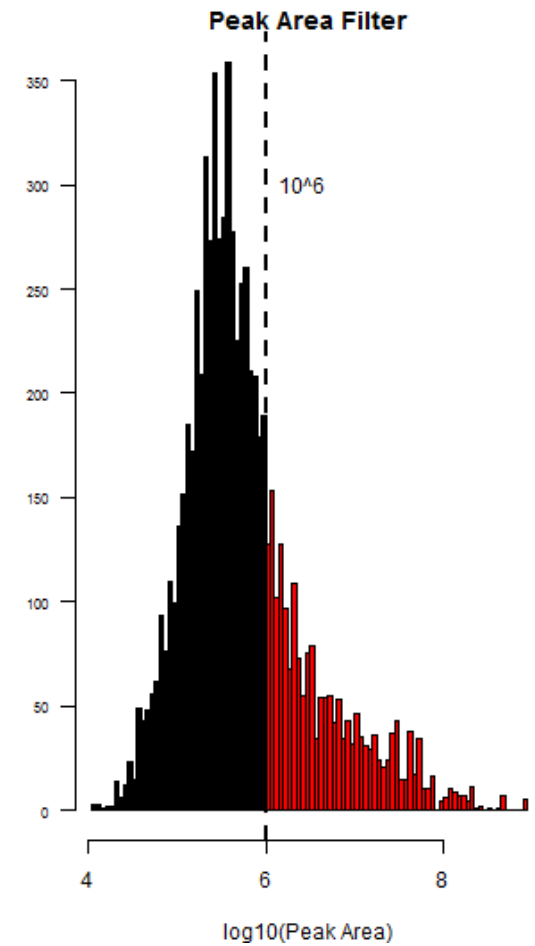
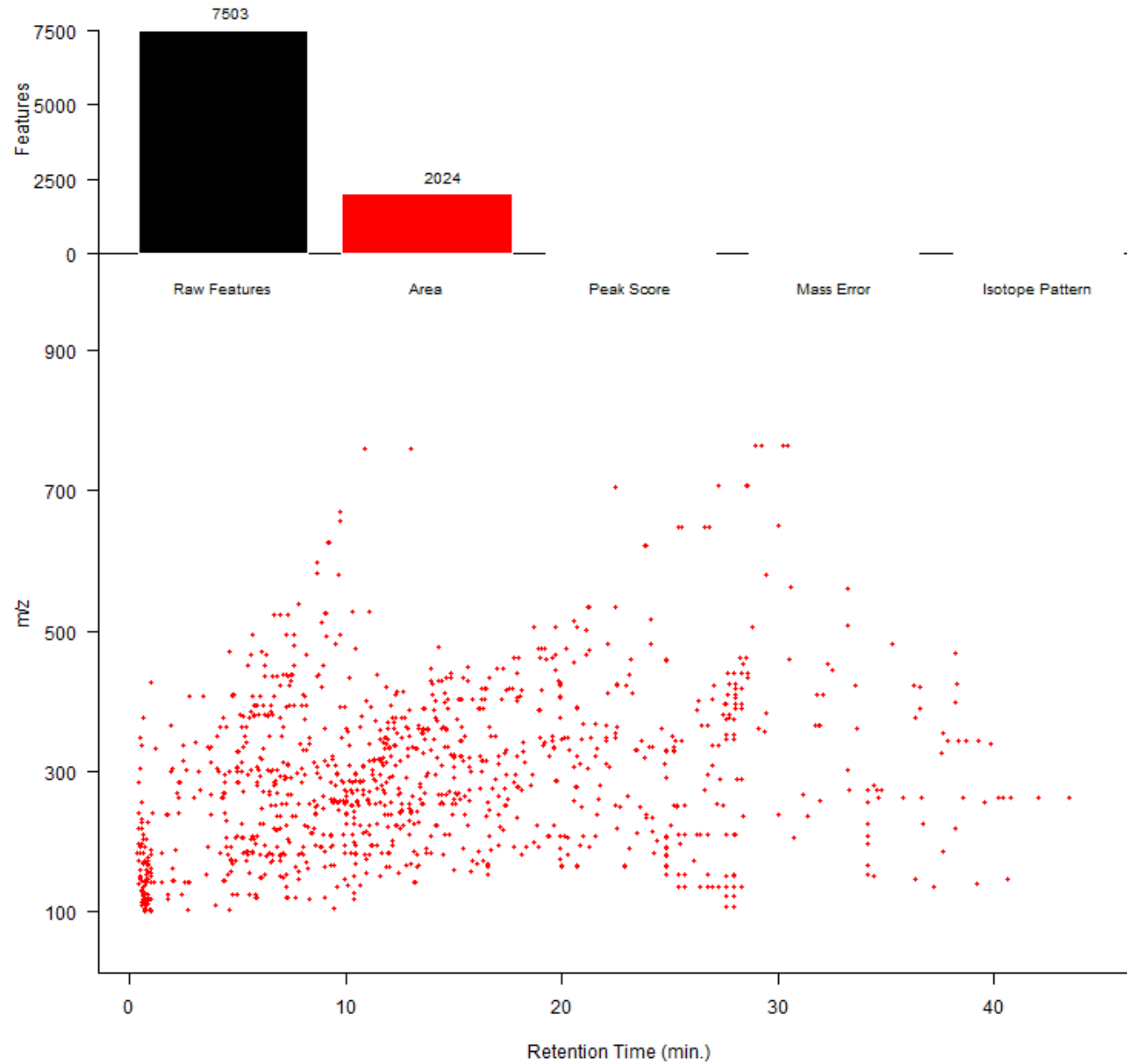
1. Thermo EFS database (1,004 substances with MS/MS library)
2. Compiled from literature including PBT, occurrence, and pharm. analytics (4,475 substances)



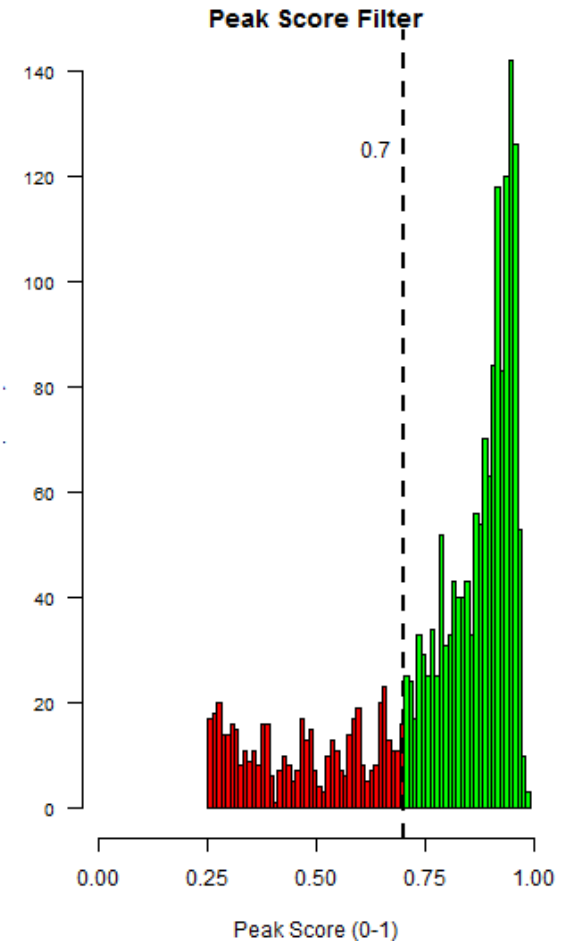
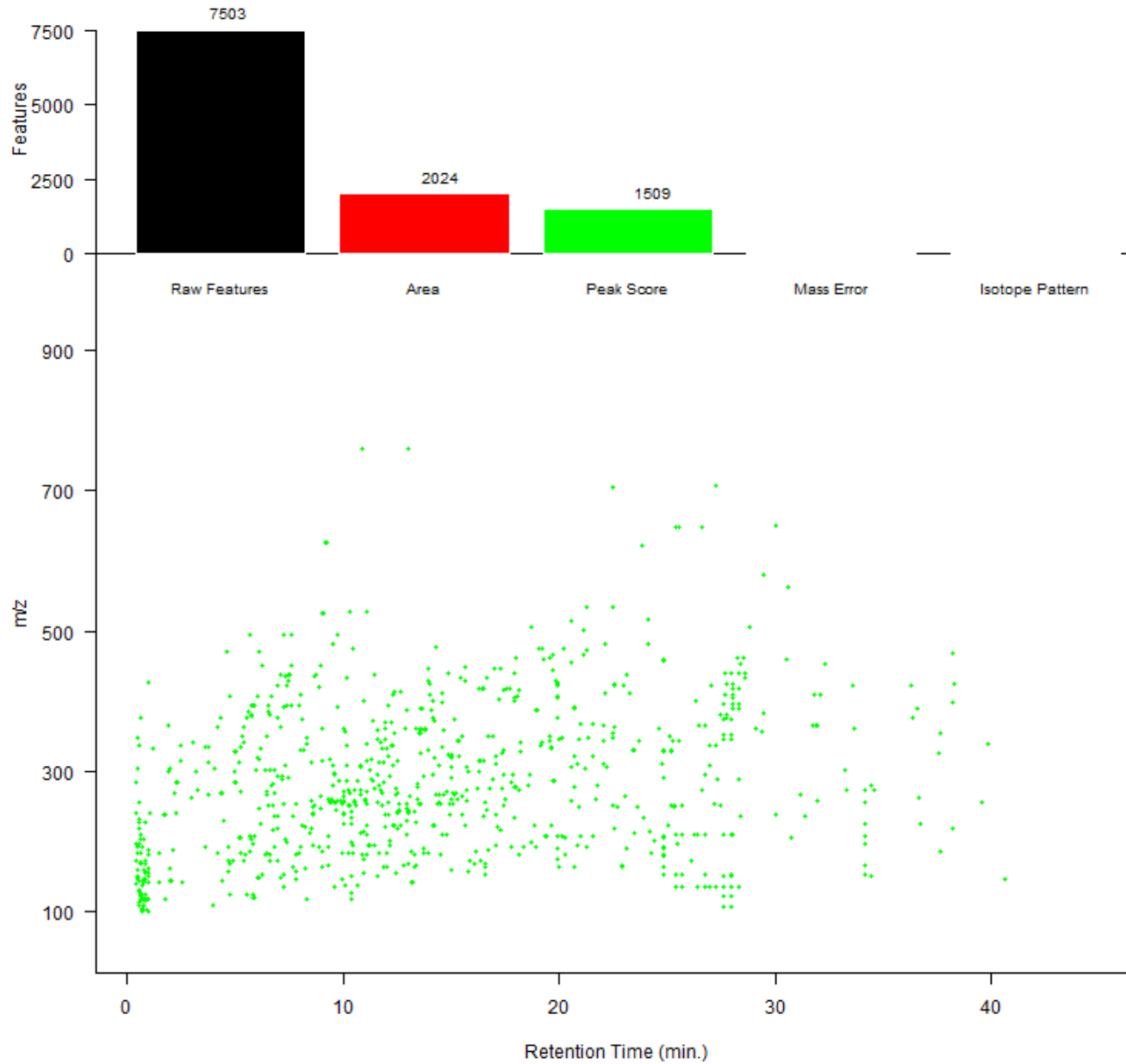
Feature filtering: Results



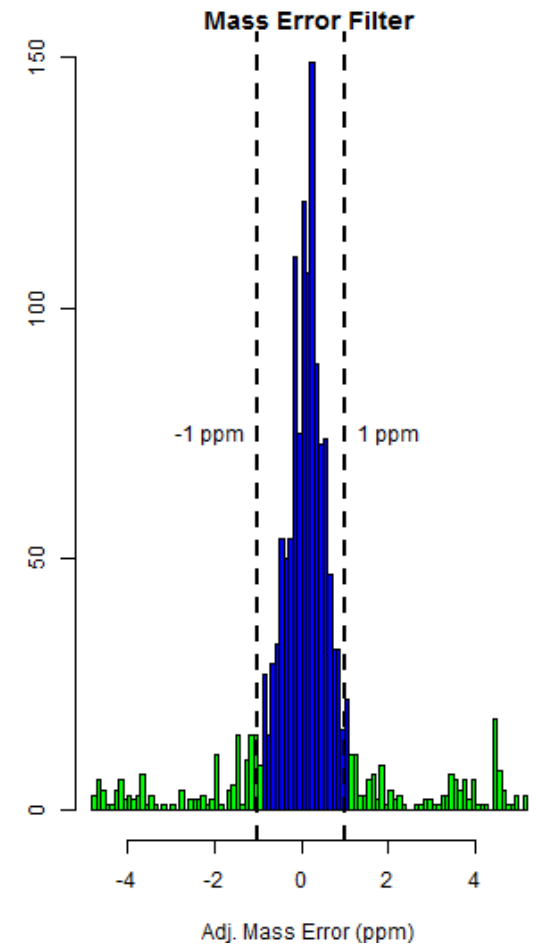
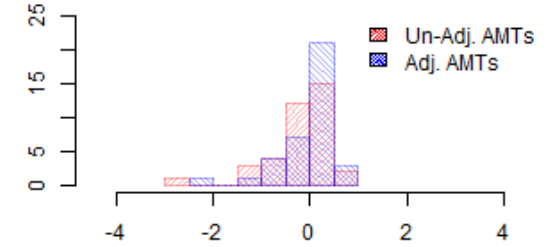
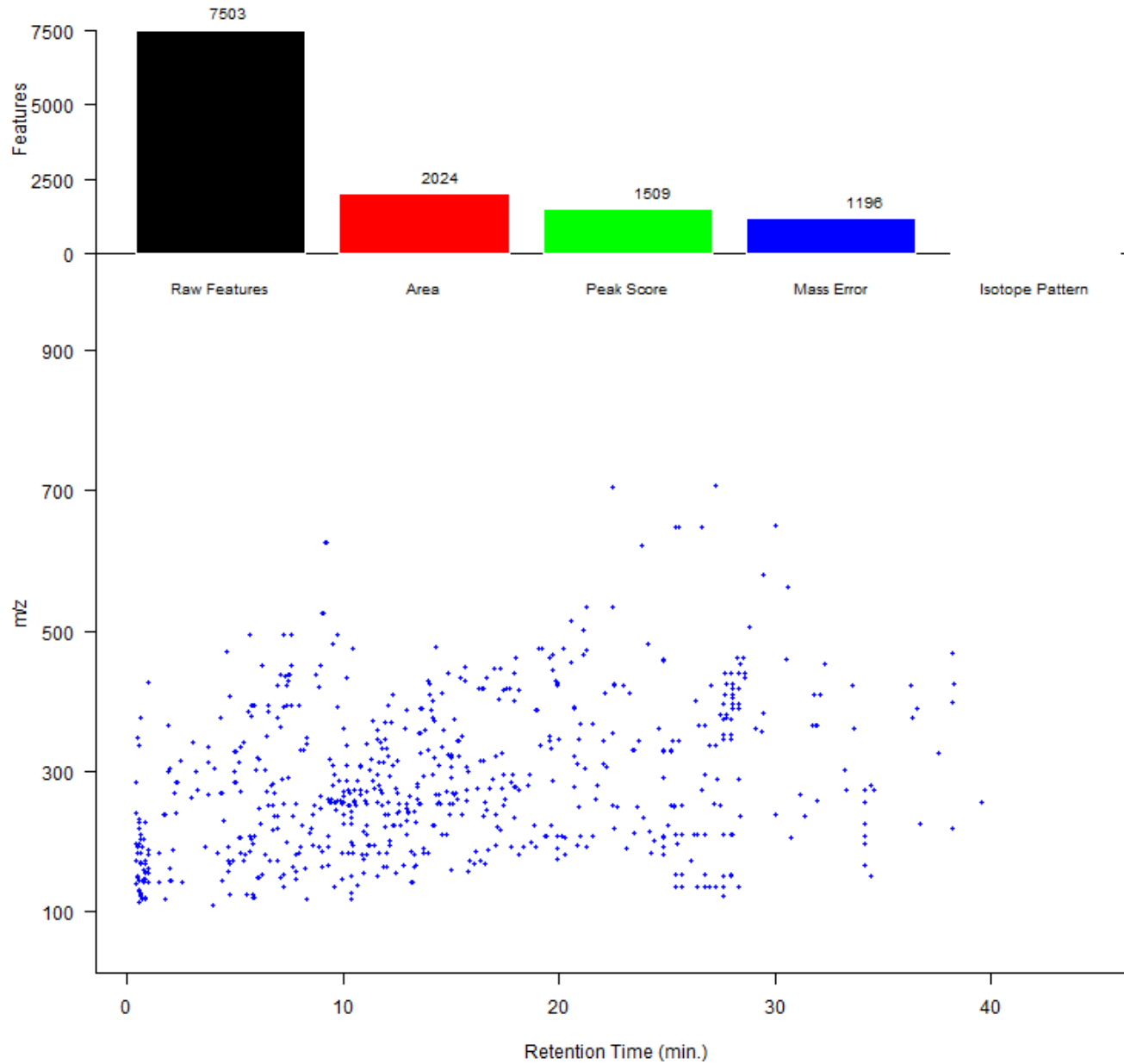
Feature filtering: Results



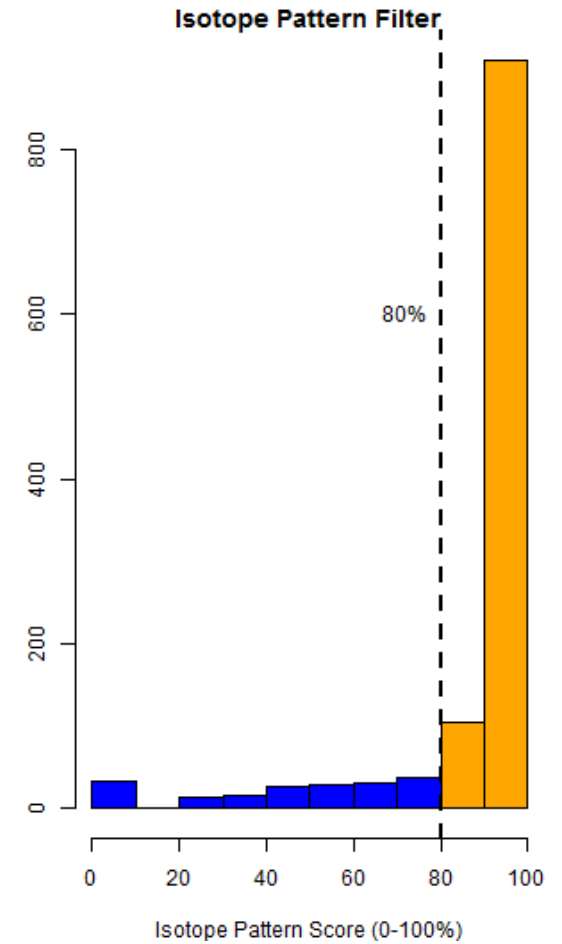
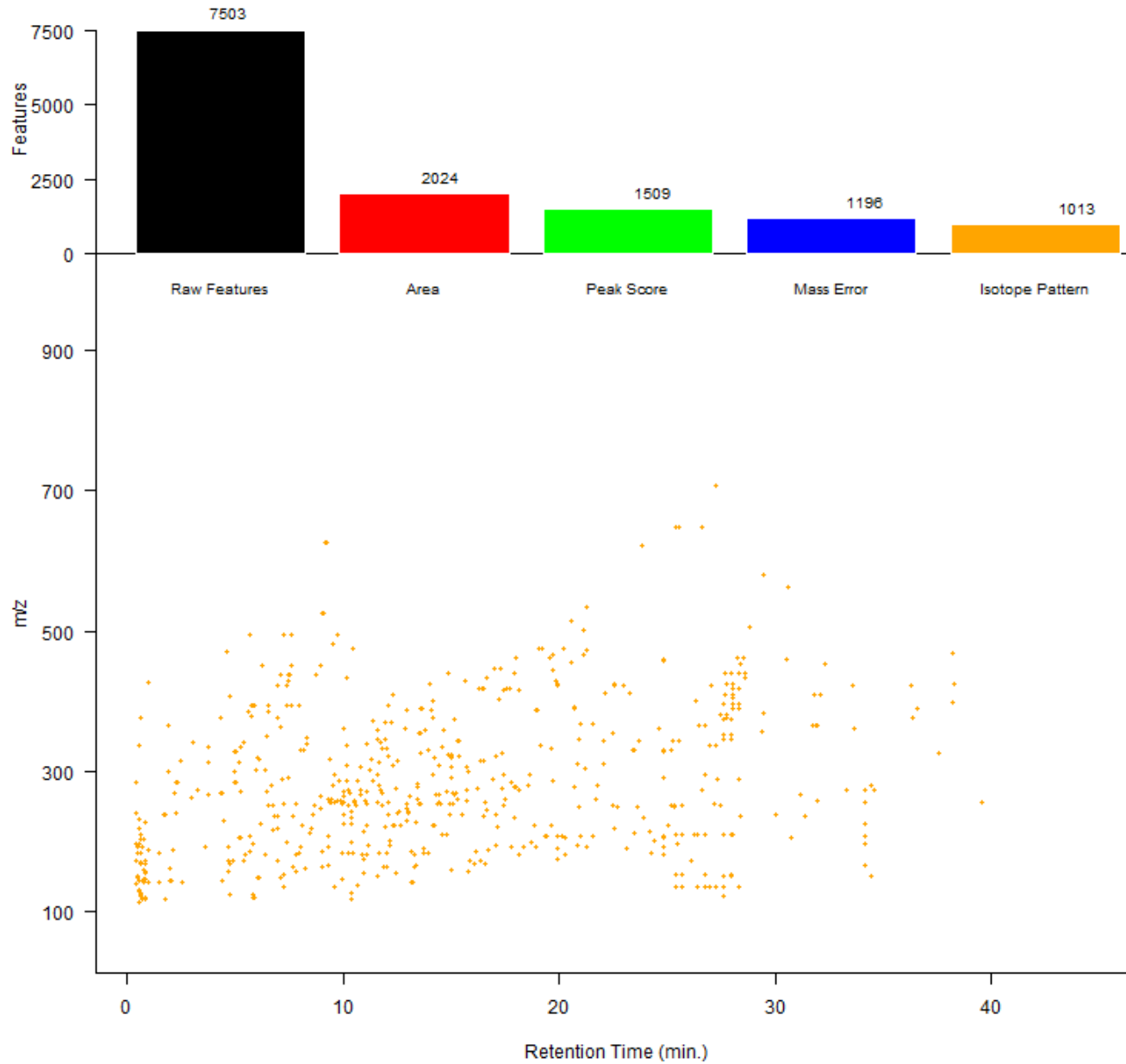
Feature filtering: Results



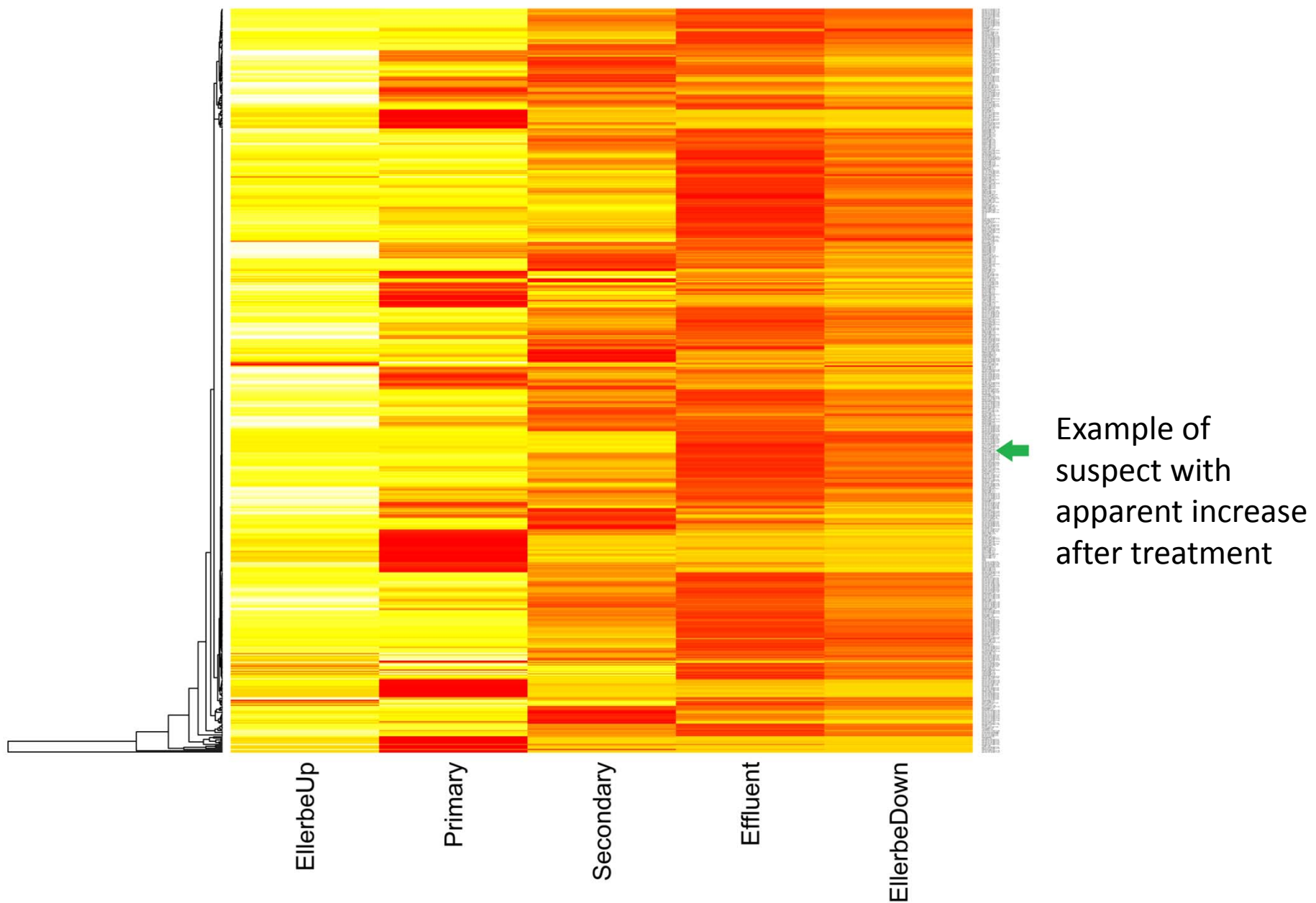
Feature filtering: Results



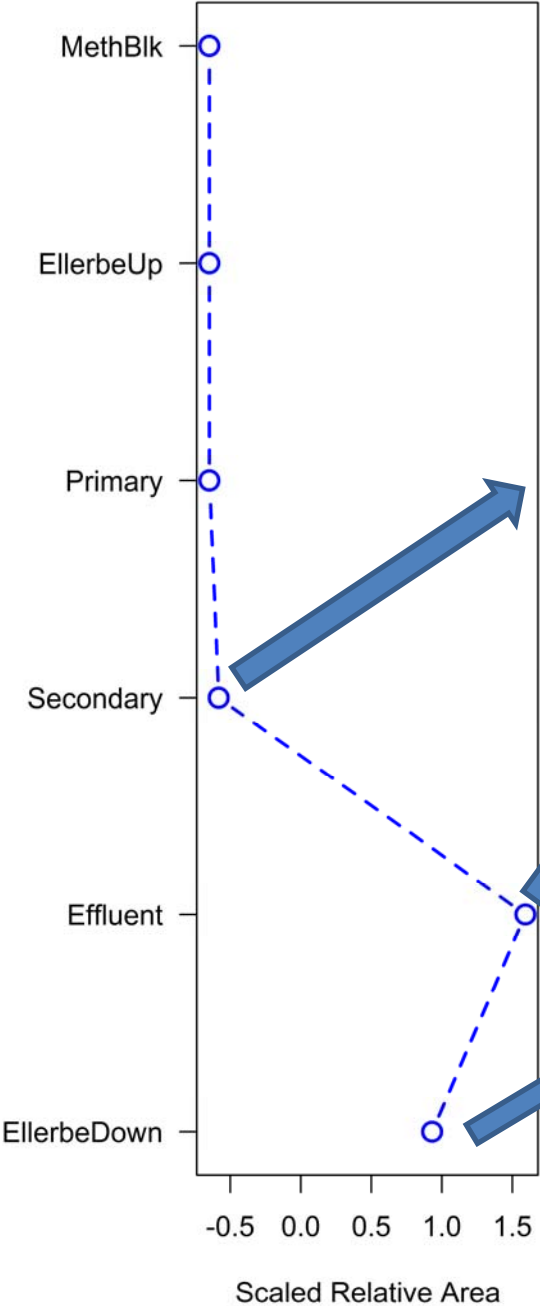
Feature filtering: Results



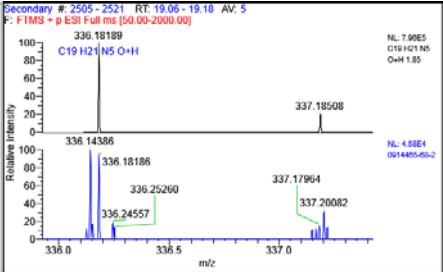
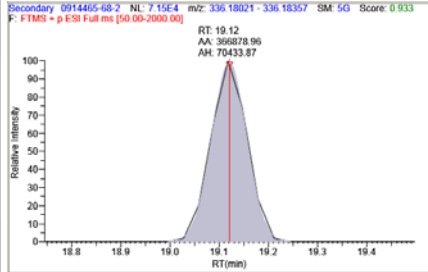
Clustering of suspect hits enables pattern-dependent analysis



Detailed inspection reveals strong increase after UV disinfection

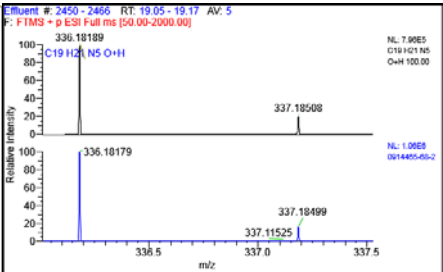
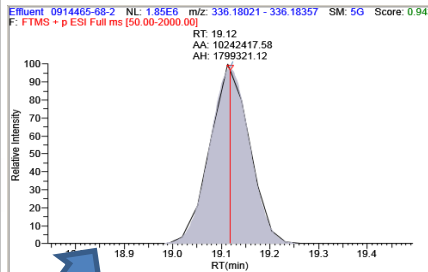


disinfection



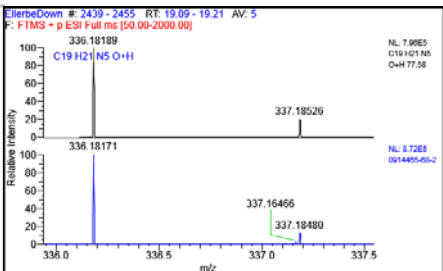
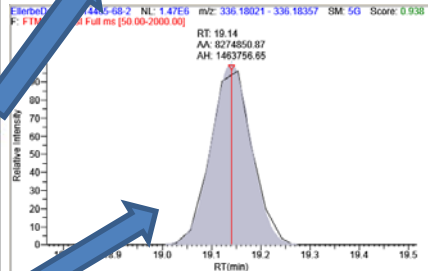
Secondary

- RT : 19.12 min
- Mass error: -0.37
- Peak area: 3.6e5



Effluent

- RT : 19.12 min
- Mass error: -0.19
- Peak area: 1.0e7



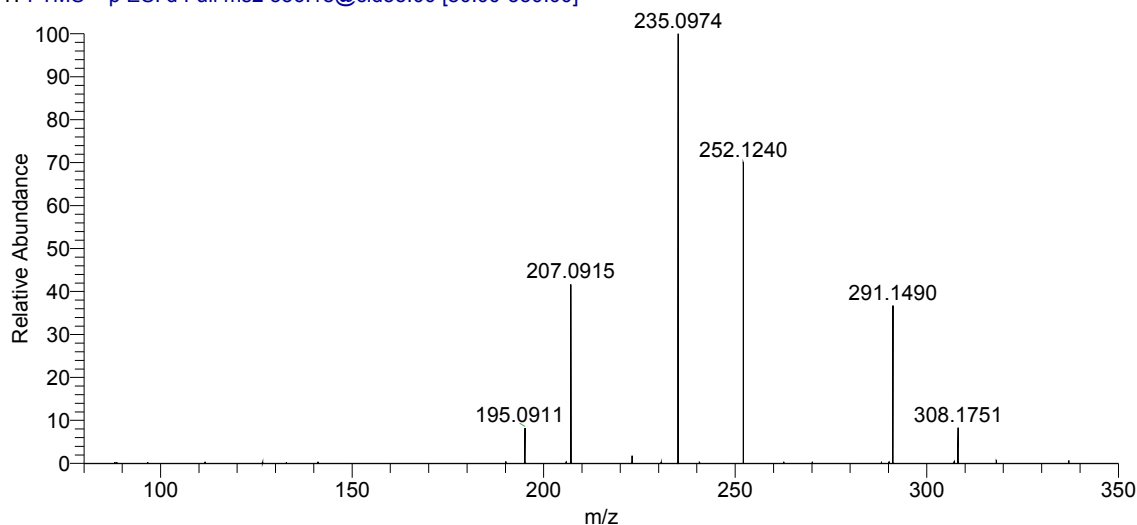
Ellerbe Down

- RT : 19.14 min
- Mass error: -0.64
- Peak area: 8.3e6

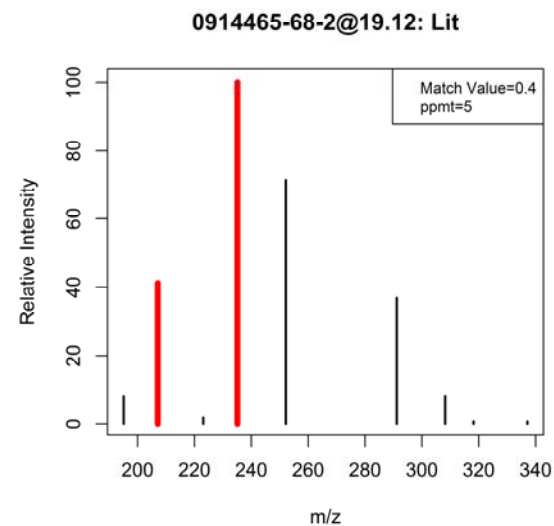
Tentative ID: Valsartan transformation product

Observed HR/AM MS²:

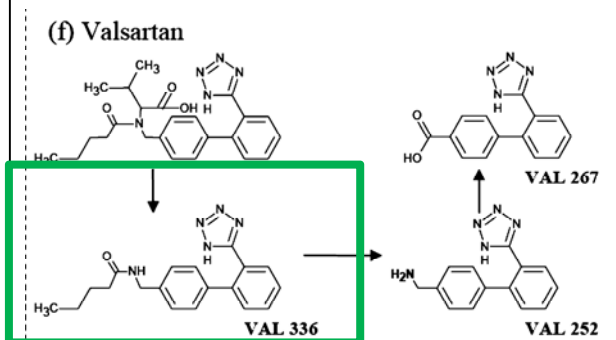
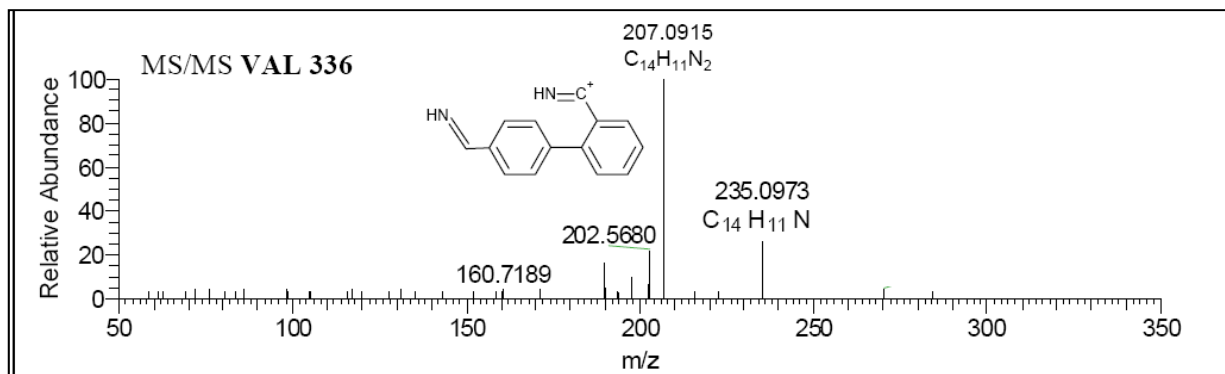
Effluent #2459 RT: 19.12 AV: 1 NL: 4.61E5
T: FTMS + p ESI d Full ms2 336.18@cid35.00 [80.00-350.00]



Match Value:



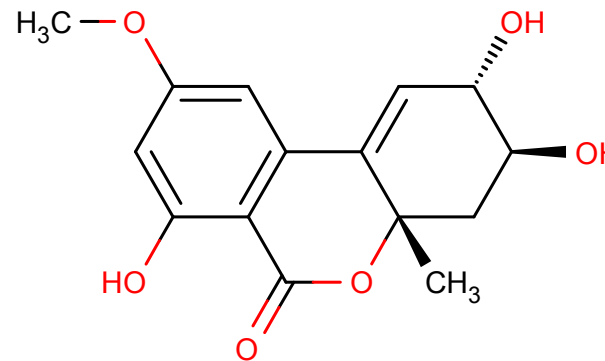
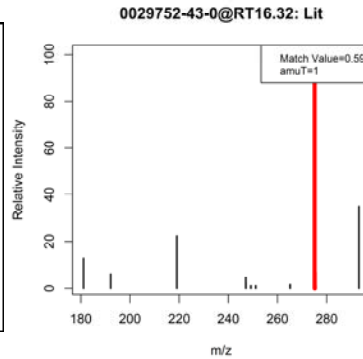
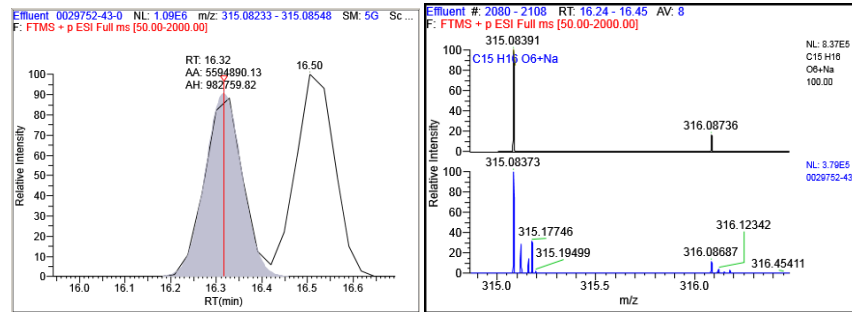
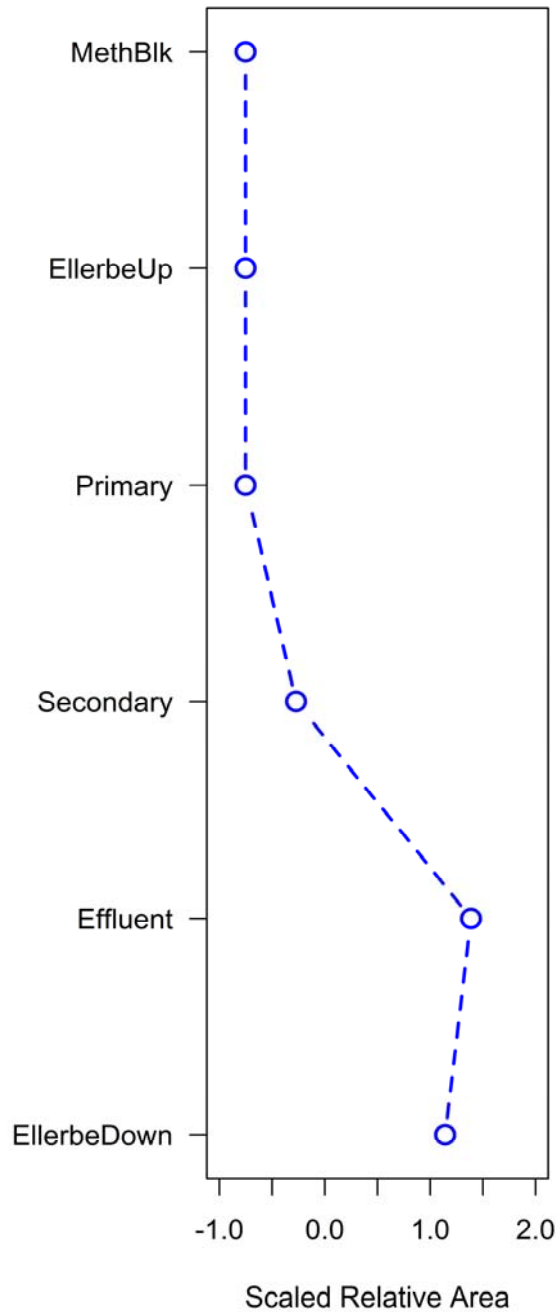
Literature HR/AM MS²:



Helbling, et. al., *Environ. Sci. Technol.* **2010**, 44, 6621-6627

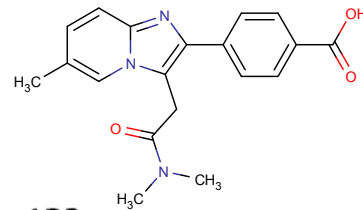
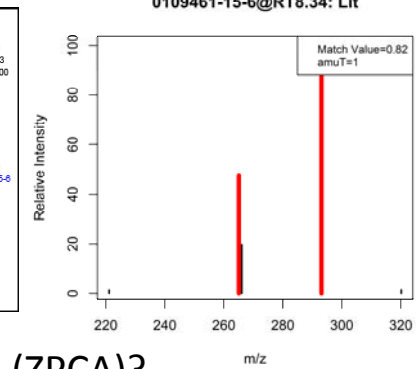
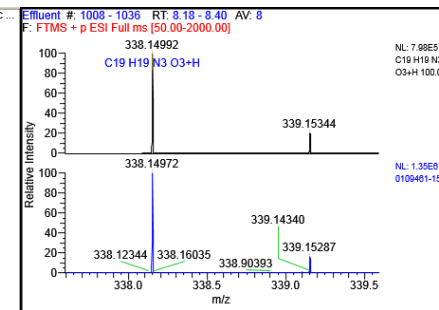
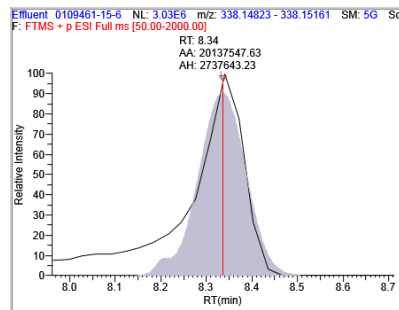
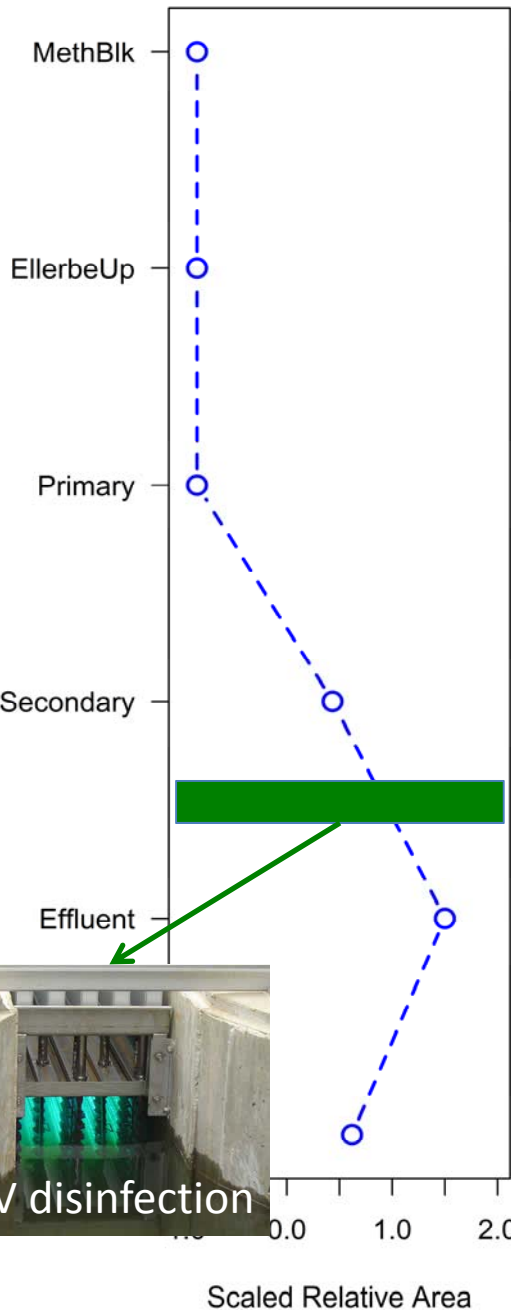
Example of a compound appearing after treatment:

altenuene



What is its source? Mycotoxin detected in foodstuffs...

Identification of a novel TP in the environment



Zolpidem carboxylic acid (ZPCA)?

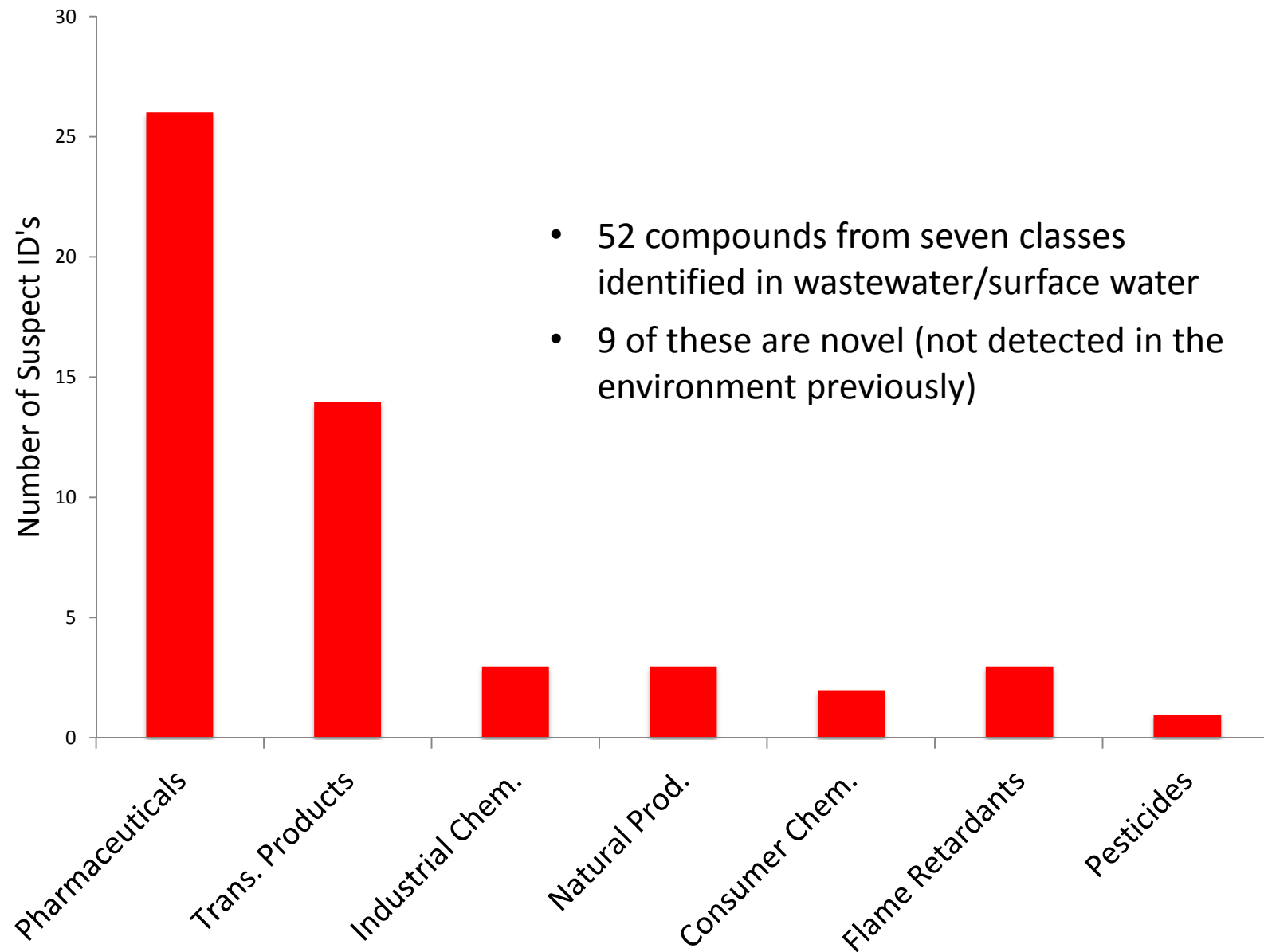
Possible photodegradation product?

Table 4. The Results of the Stress Degradation Tests Using Different Conditions

Stress Test Condition	Solvent	Temperature	Time	% of Zolpidem
Acidic	1 M HCl	95 °C	4 h	90.2
	2 M HCl	70 °C	3 h	97.0
	2 M HCl	95 °C	3 h	75.7
	5 M HCl	95 °C	3 h	62.9
Alkaline	1 M NaOH	95 °C	1 h	35.1
Oxidative	3% H ₂ O ₂	70 °C	1 h	54.6
	10% H ₂ O ₂	70 °C	1 h	37.6
Photolytic				
UV light	Solid form	Room temperature	5 days	90.2
UV light	Water	Room temperature	5 days	45.0
Visible light	Solid form	Room temperature	5 days	84.6
Visible light	Water	Room temperature	5 days	83.0
Heat	Solid form	80 °C	5 days	85.0
	Water	80 °C	5 days	85.7

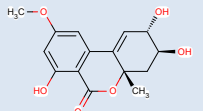
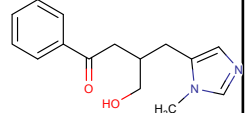
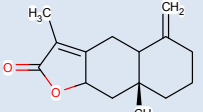
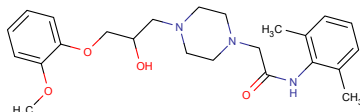
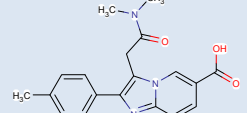
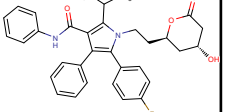
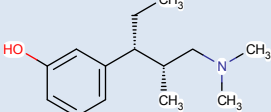
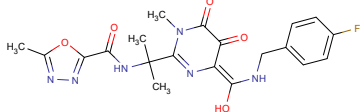
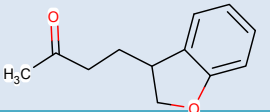


Results: Suspect compounds tentatively identified



- 52 compounds from seven classes identified in wastewater/surface water
- 9 of these are novel (not detected in the environment previously)

Tentative Novel Compound Identifications LF12

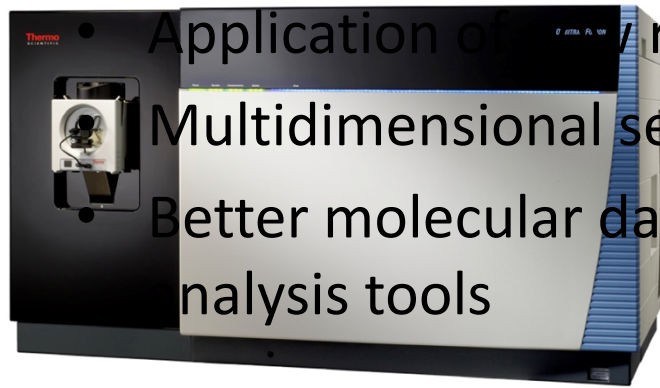
<u>Compound</u>	<u>Class</u>	<u>Structure</u>
Altenuene	Mycotoxin	
1-Butanone, 3-(hydroxymethyl)-4-(1-methyl-1 <i>H</i> -imidazol-5-yl)-1-phenyl-	Transformation product of pilocarpine (glaucoma treatment)	
Atractylenolide II	Sesquiterpene natural product	
Ranolazine	Antianginal	
ZPCA	Transformation product of zolpidem (Ambien)	
Atorvastatin lactone	Transformation product of atorvastatin (Lipitor)	
Tapentadol	Analgesic	
Raltegravir	Antiretroviral HIV treatment	
4-(2,3)-dihydro-3-benzofuranyl)-2-butanone	Transformation product of butylphthalide (celery oil comp.)	

Conclusions

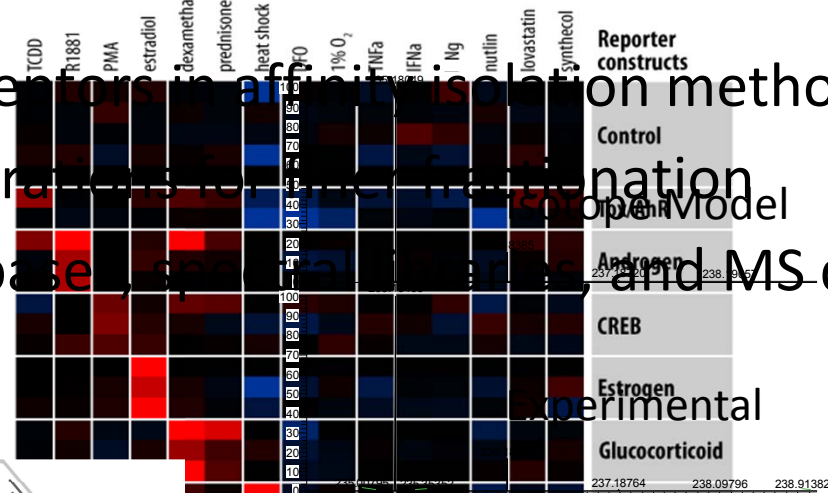
- Systematic effects-directed analysis approaches can solve practical problems in environmental contaminant science.
- Affinity purification approaches based on mode-of-action-specific molecular interactions of toxicants with receptors are well-suited for coupling with analytical detection methods.
- Process-dependent analysis is a powerful tool in combination with non-targeted or suspect screening for prioritizing the most relevant micropollutants in a contaminant mixture.
- New generation non-targeted analysis relies CRITICALLY on both analytical technologies (e.g. mass spectrometry) and high-content bioassays.

What's next for non-targeted analysis??

- New screening assays for determining effects:
 - High throughput fish embryo assays (*in vivo*)
 - Multi-target reporter assays (*in vitro*)
- Ultra-high resolution mass spectrometry

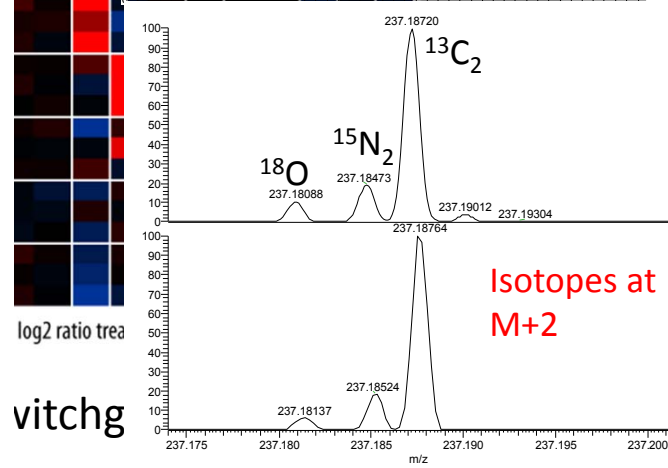
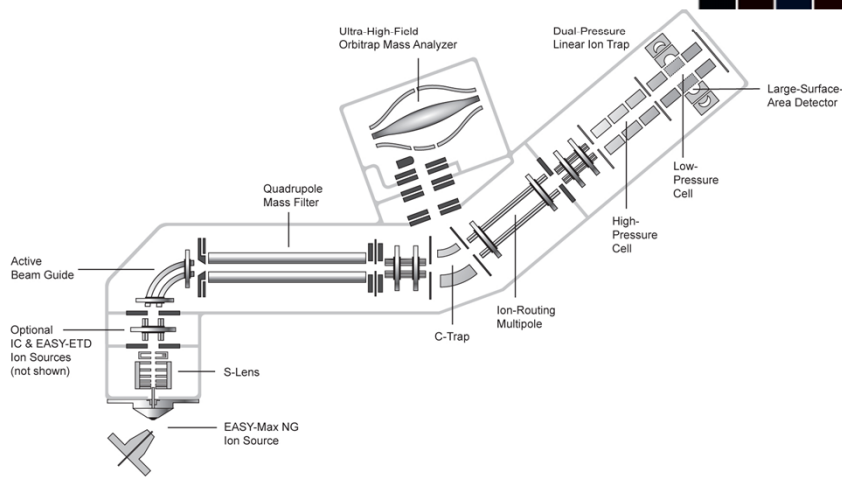


- Application of ^{13}C / receptors in affinity isolation methods
- Multidimensional separations for metabolite identification
- Better molecular database, spectral libraries, and MS data-analysis tools

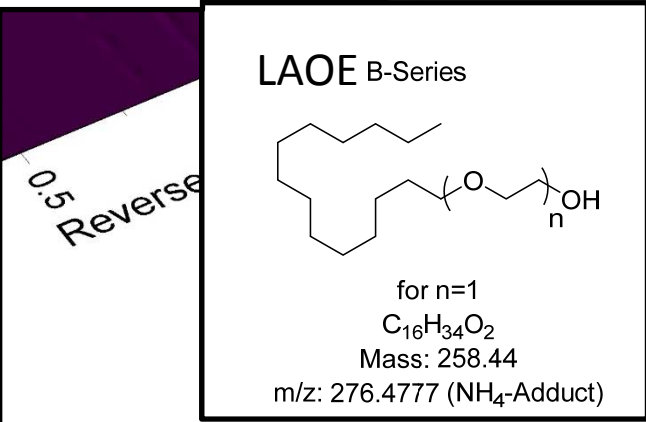
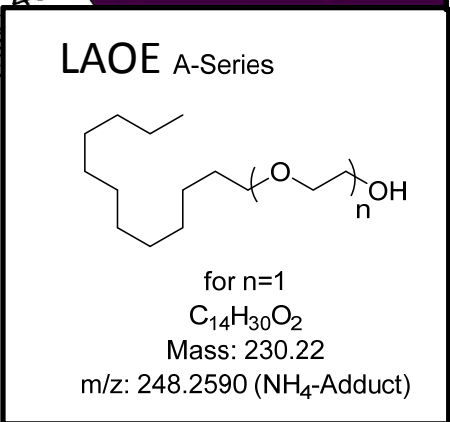
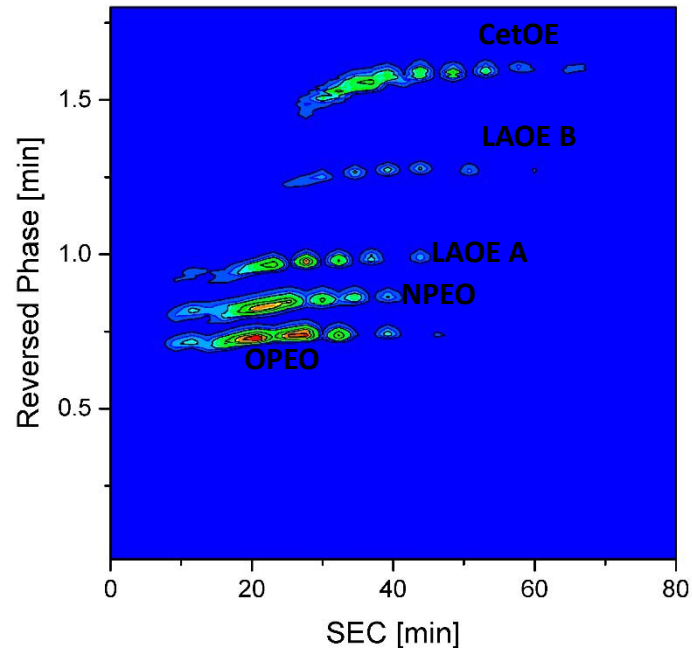
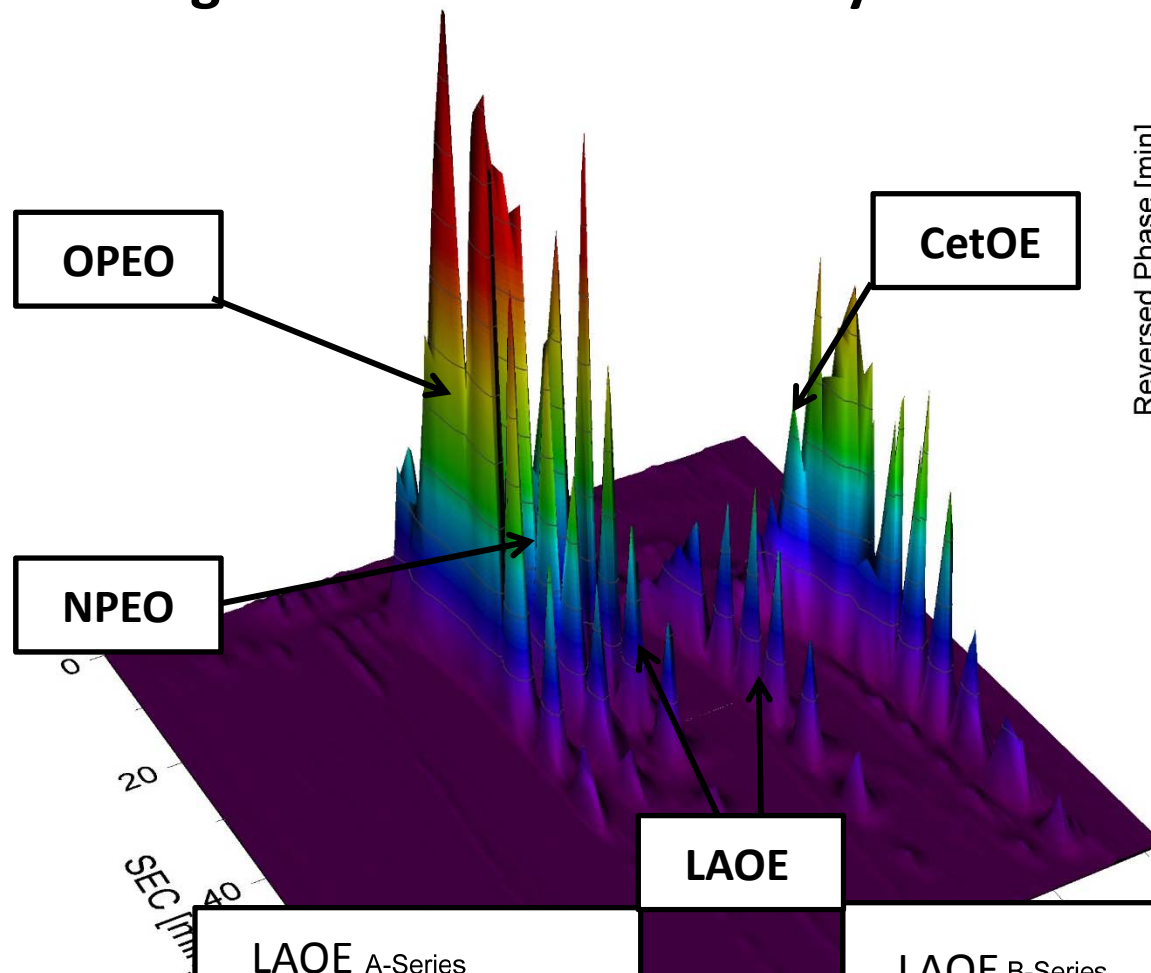


N₂ 9.9714
 C₁₄H₂₃O₁N₂
 C₁₄H₂₃O₁N₂
 p (gs, s, p, 8) Chg 1
 R 250000 Res .Pwr . @FWHM

 9.29E6
 NDurhCompHLB3#1386 RT:
 5.95 AV: 1 F: FTMS + pESI
 Full ms [50.00-1000.00]



Resolving nonionic Surfactants by 2D-HPLC



Acknowledgements

Graduate students

Gordon Getzinger
Bernadette Vogler
Katy Stencel
Audrey Bone
Laura MacCauley
Lauren K. Shaw

Undergraduate students

Katherine C. Hyland
Akshay Save

Postdoctoral researchers

Kimberly Ralston-Hooper
Carin Huset



Collaborators

Tara Sabo-Attwood (U. Florida)
Tom Chandler (U. South Carolina)
Alan Kolok (U. Nebraska)
Norm Shea, Becky Dennis, Brad
French (Kiawah Island)
Steve Corsi (USGS)
Steve Geis and Amy Mager
(WSLH)
Devon Cancilla (Western
Washington U.)

