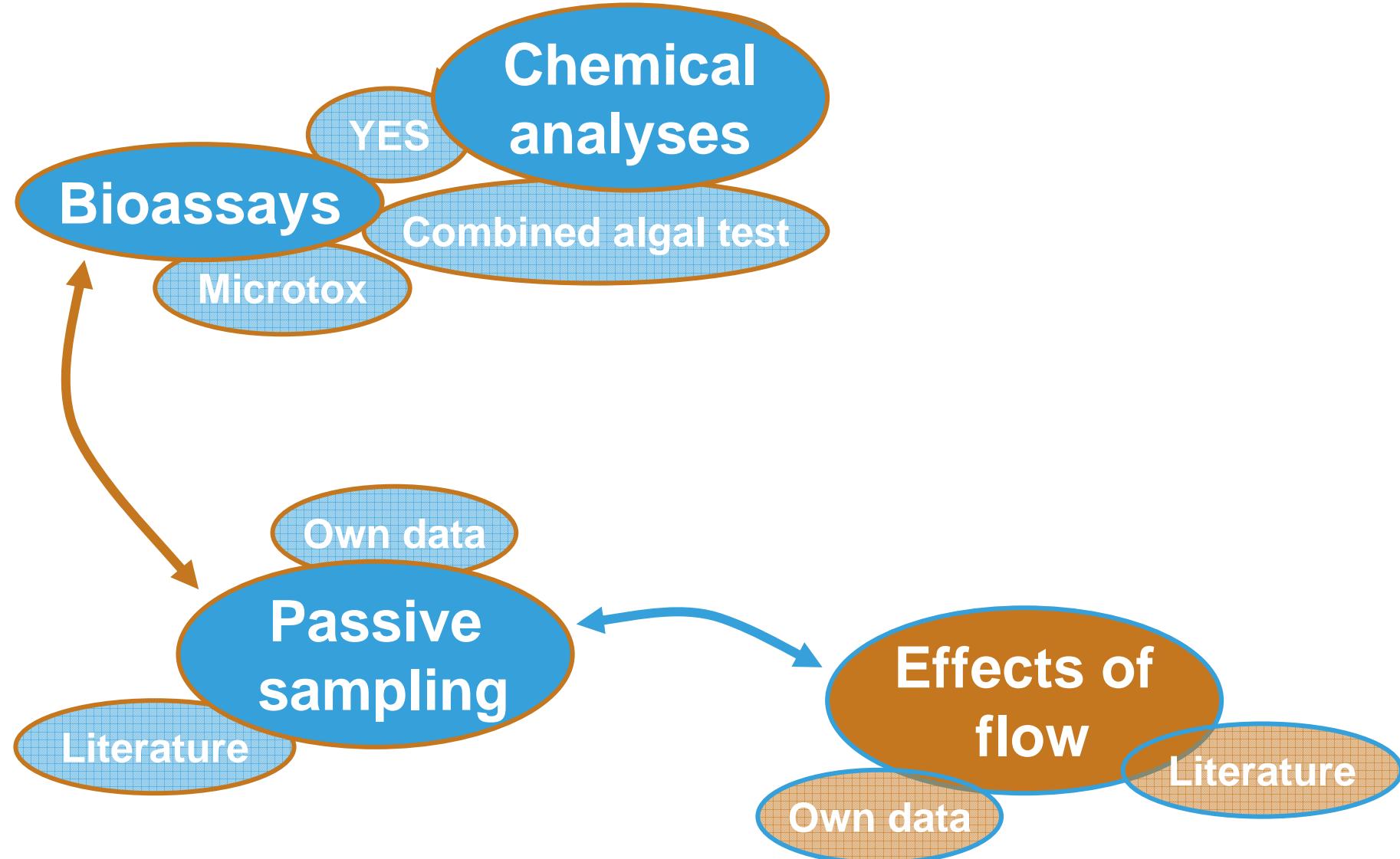


Combining passive sampling with bioassays

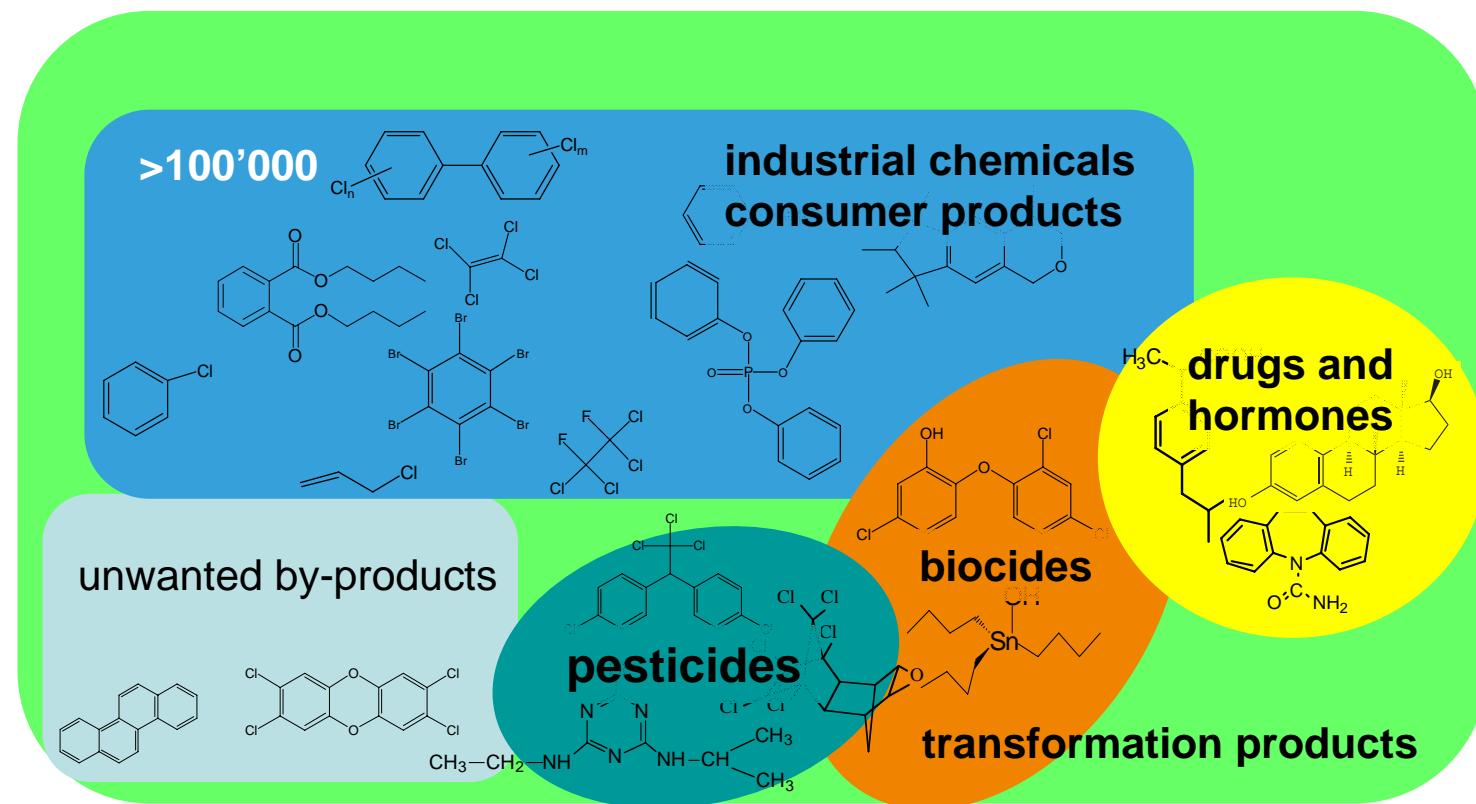
Evaluating effects of flow under environmental conditions

Contents



A view of the chemical world

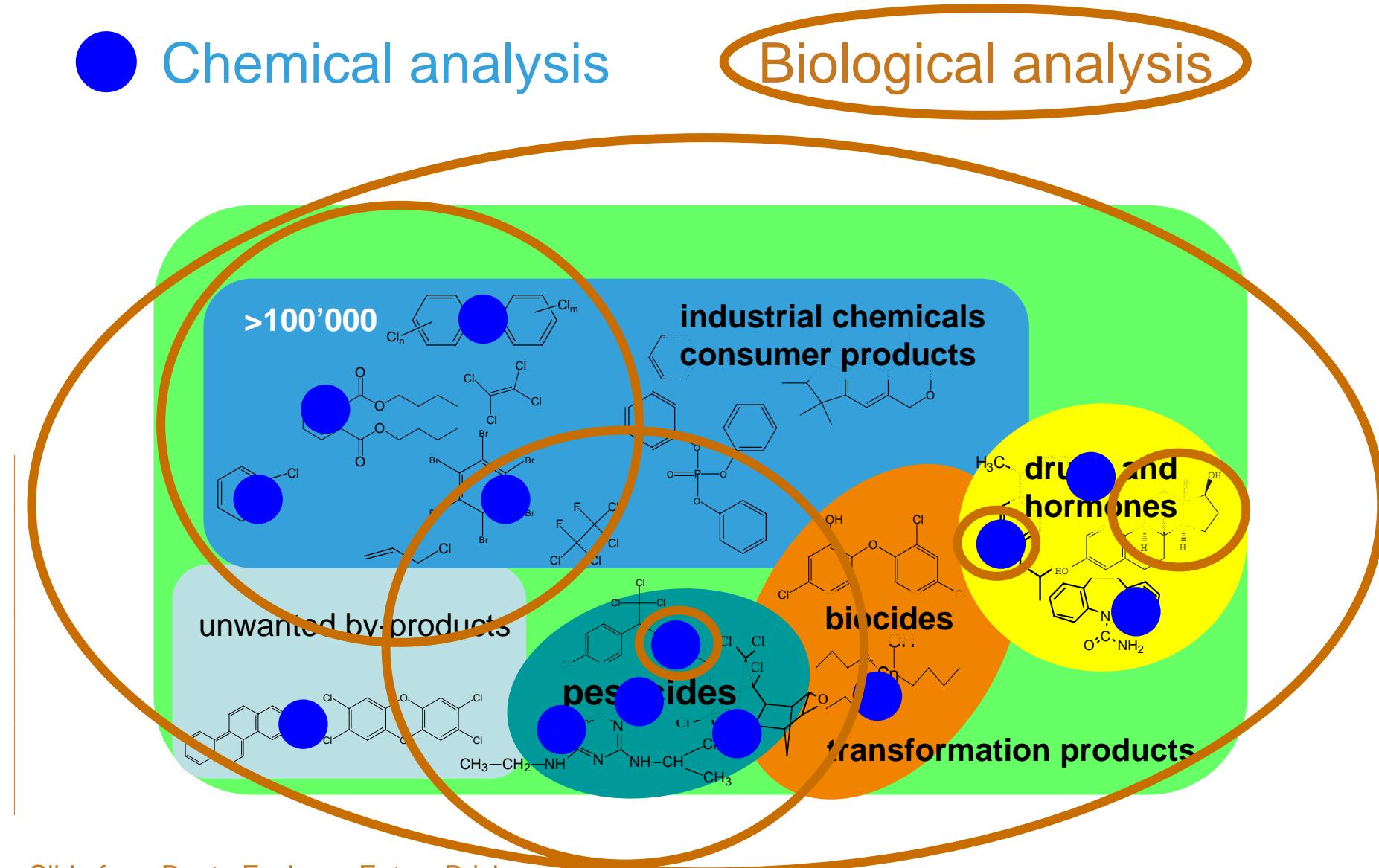
© eawag, Swiss Federal Institute for Water Research



A rationale for bioassays

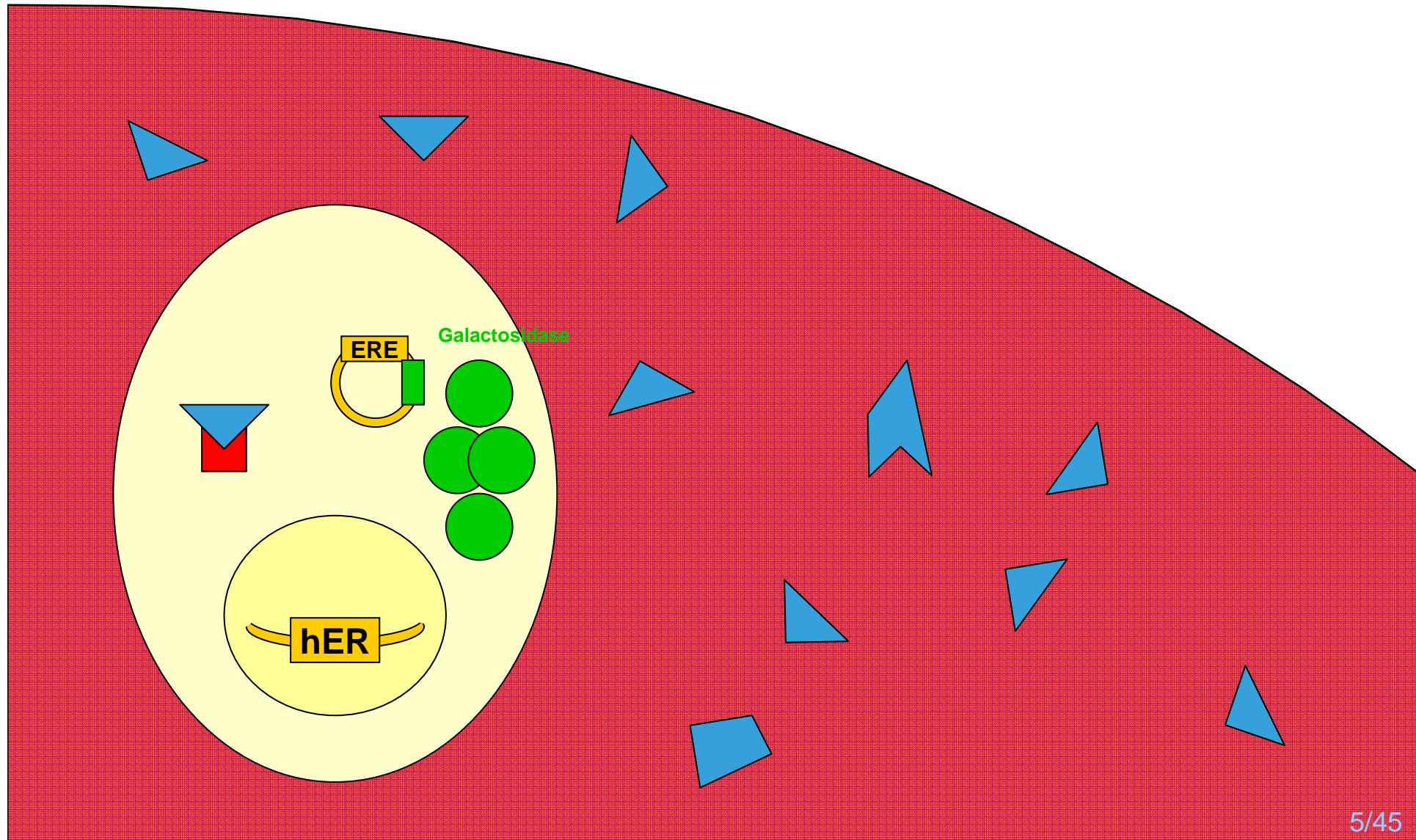
Chemical analysis

Biological analysis



Yeast estrogen screen

Routledge and Sumpter, 1996 ET&C 15

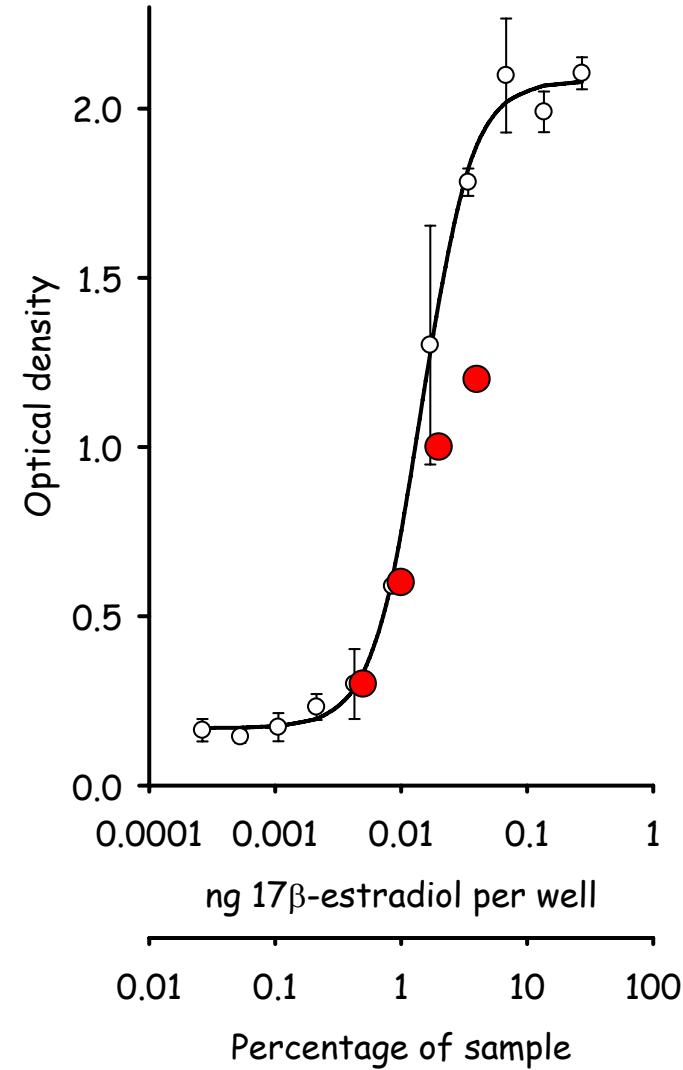
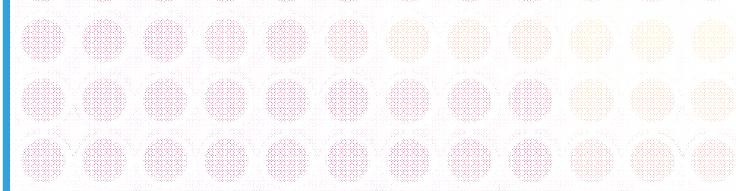


Toxicity equivalents

Data evaluation along:
“toxicity equivalent concentrations”

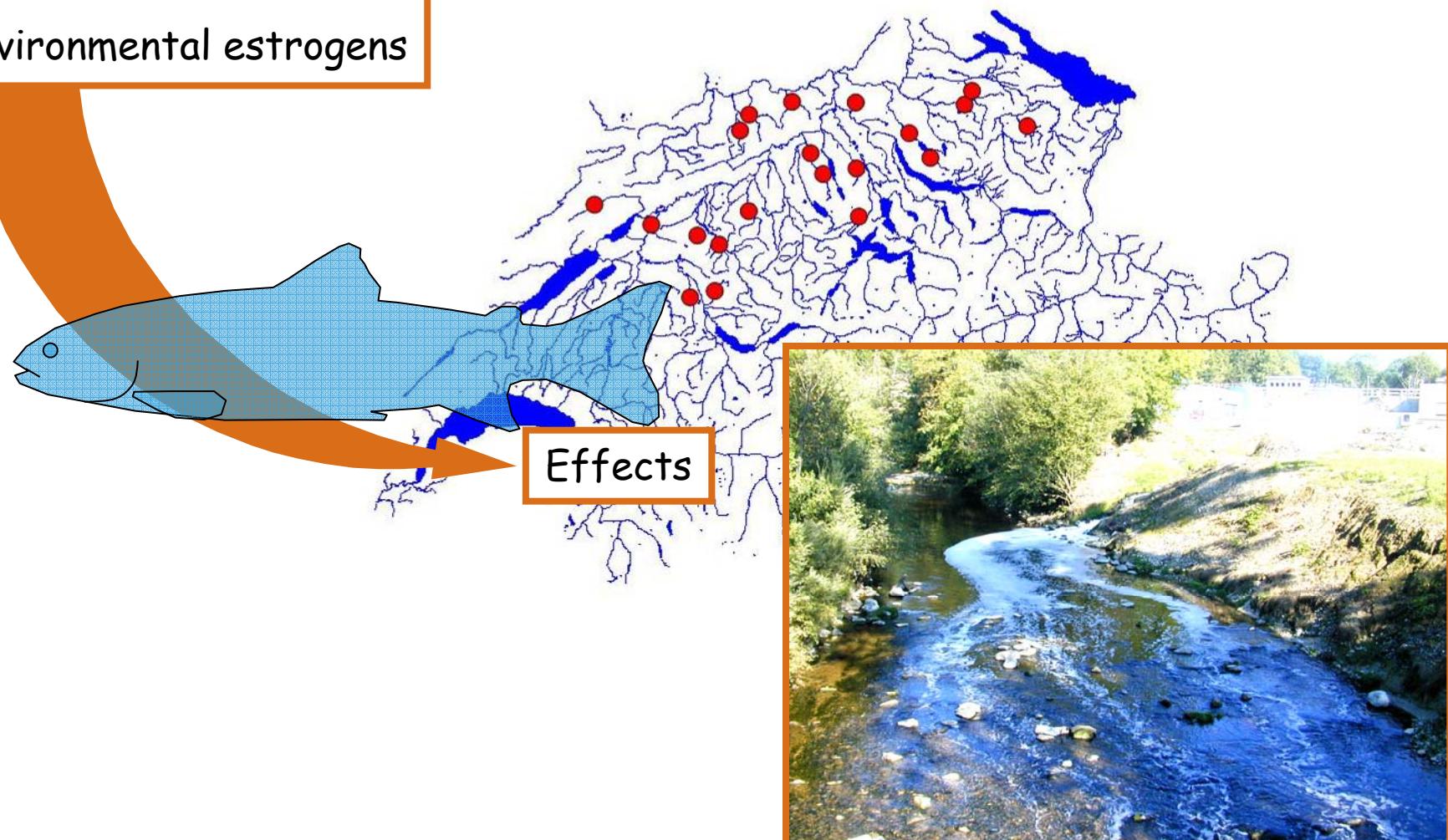


EEQ: 17β -estradiol equivalents



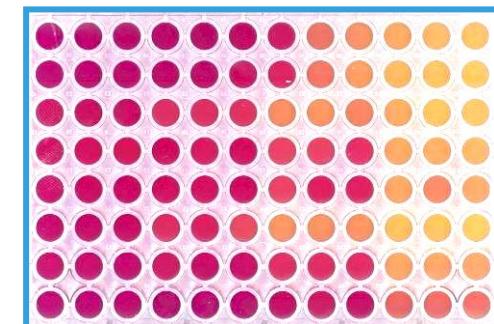
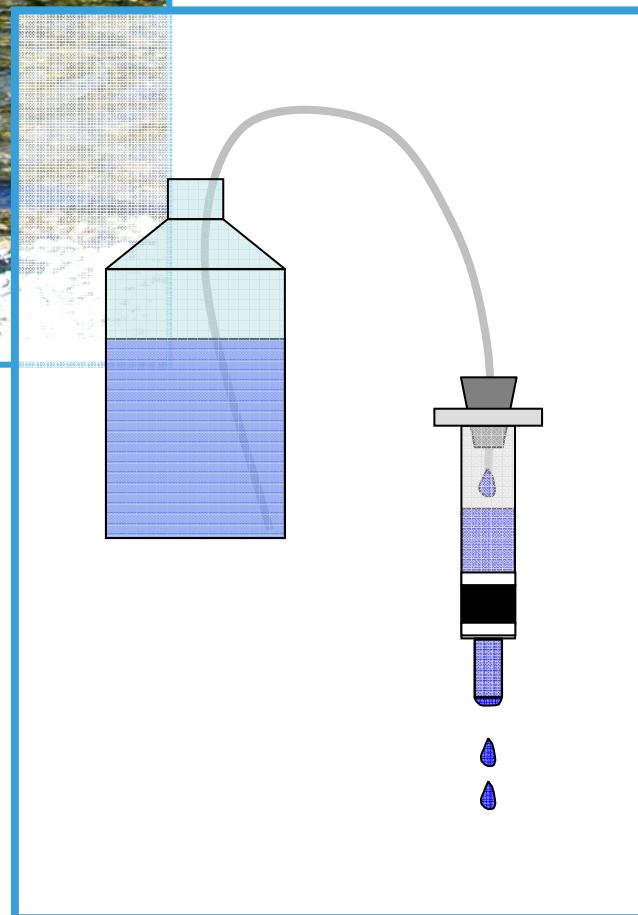
From grab sampling to passive sampling

Environmental estrogens



Effects

From grab sampling to passive sampling



From grab sampling to passive sampling

“Estrogens in Swiss rivers and effluents - sampling matters” Vermeirssen et al. (2008) Chimia 62

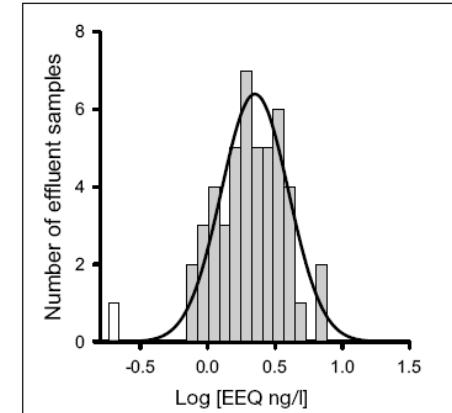


Fig. 4. Histogram of effluent estrogenic activity (EEQ). The normal distribution plot was fitted to 47 data (grey); one EEQ (open bar) was excluded.^[20]

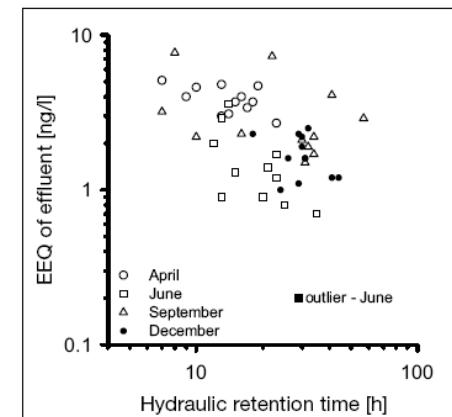
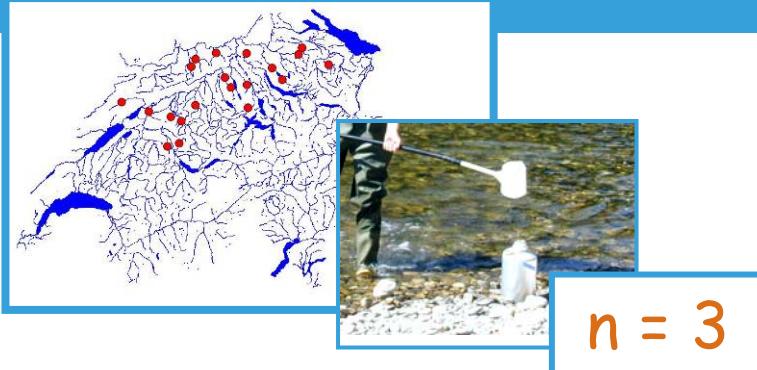
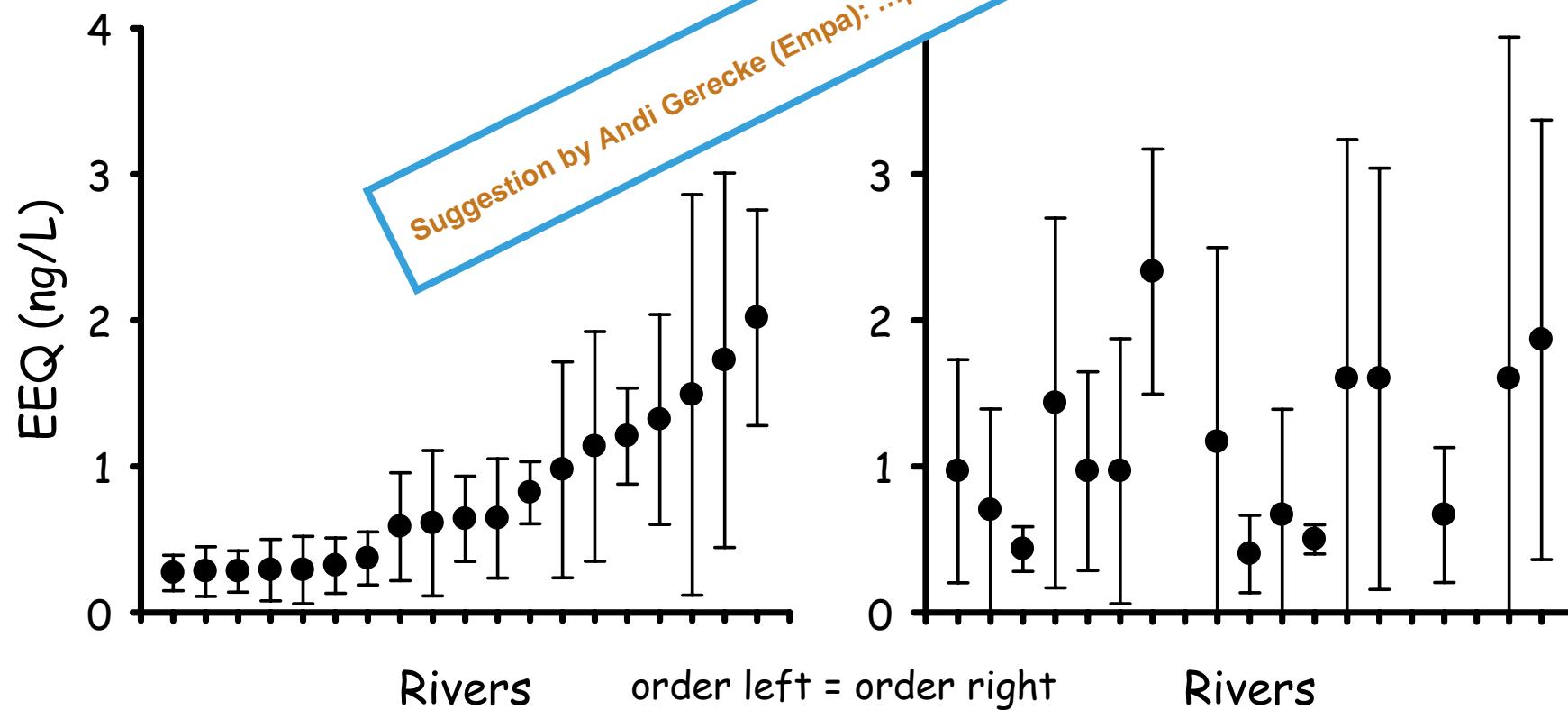


Fig. 5. Relationship between the hydraulic retention time (HRT) and the estrogenic activity (EEQ) of effluent over four 12-day sampling periods. The association between HRT and EEQ was negative in all but one sampling period (small filled circles).^[20]

From grab sampling to passive sampling



$n = 3$



Nov 2003: passive sampling and bioassays

eawag
aquatic research...



CHEMOSPHERE

ELSEVIER

Chemosphere 54 (2004) 1217–1224

www.elsevier.com/locate/chemosphere

Purification of triolein for use in semipermeable membrane devices (SPMDs)

J.A. Lebo ^{a,*}, F.V. Almeida ^b, W.L. Cranor ^a, J.D. Petty ^a, J.N. Huckins ^a,
A. Rastall ^c, D.A. Alvarez ^a, B.B. Mogensen ^d, B.T. Johnson ^a

CHEMOSPHERE

Chemosphere 54 (2004) 695–705

www.elsevier.com/locate/chemosphere

A holistic passive integrative sampling approach for assessing the presence and potential impacts of waterborne environmental contaminants

J.D. Petty ^{a,*}, J.N. Huckins ^a, D.A. Alvarez ^a, W.G. Brumbaugh ^a,
W.L. Cranor ^a, R.W. Gale ^a, A.C. Rastall ^b, T.L. Jones-Lepp ^c,
T.J. Leiker ^d, C.E. Rostad ^d, E.T. Furlong ^d

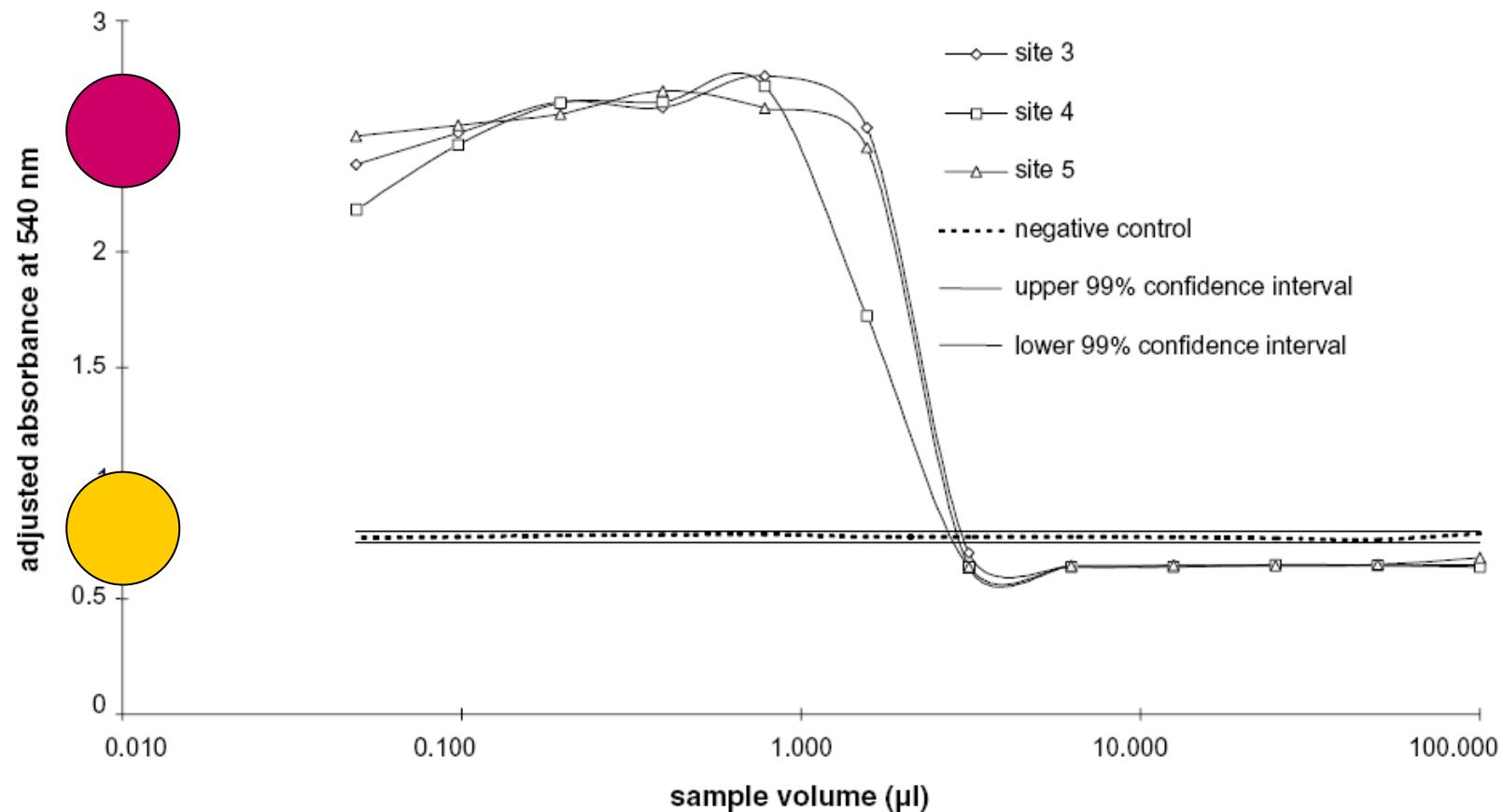
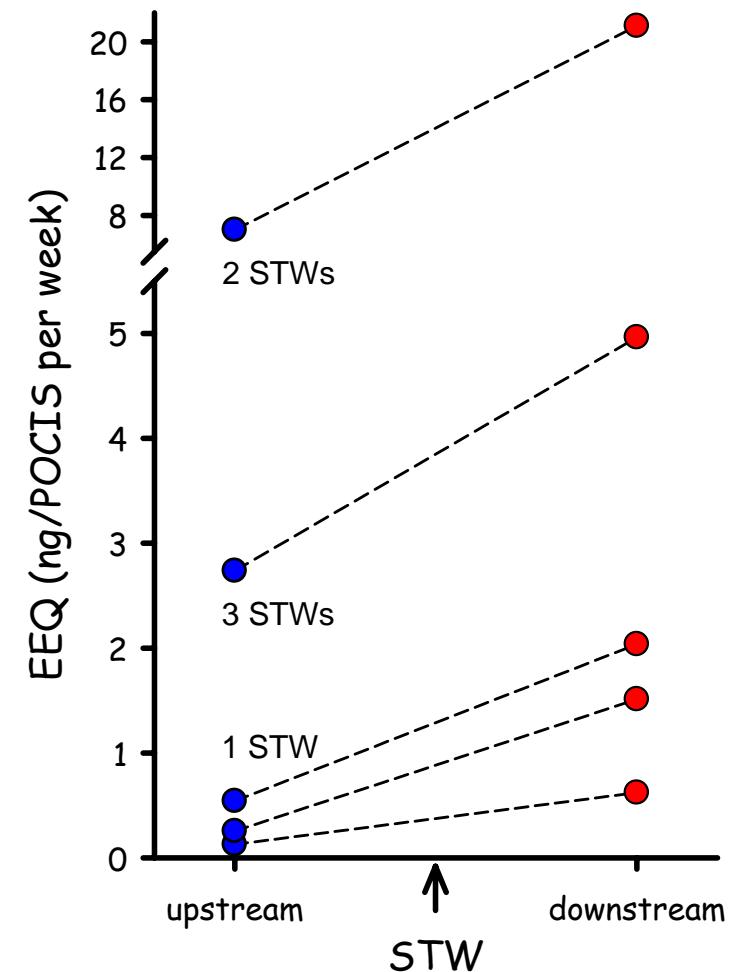
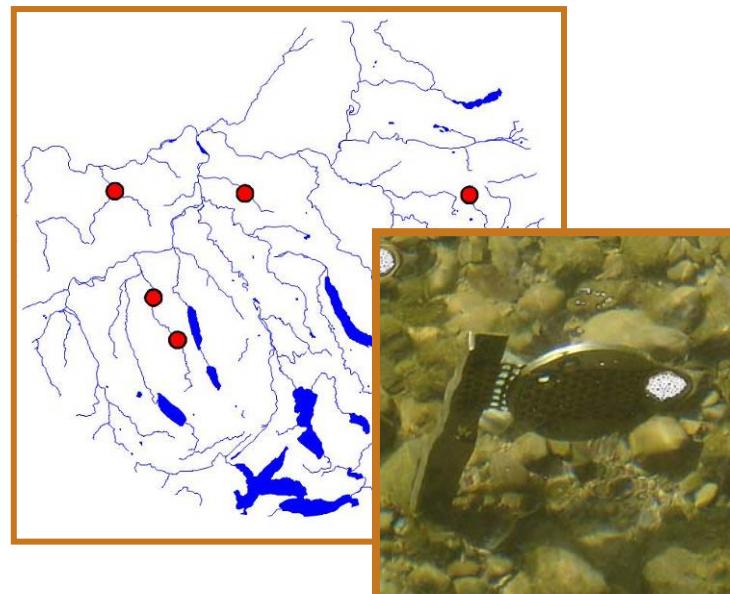


Fig. 2. Results of the YES assay performed on POCIS extracts.

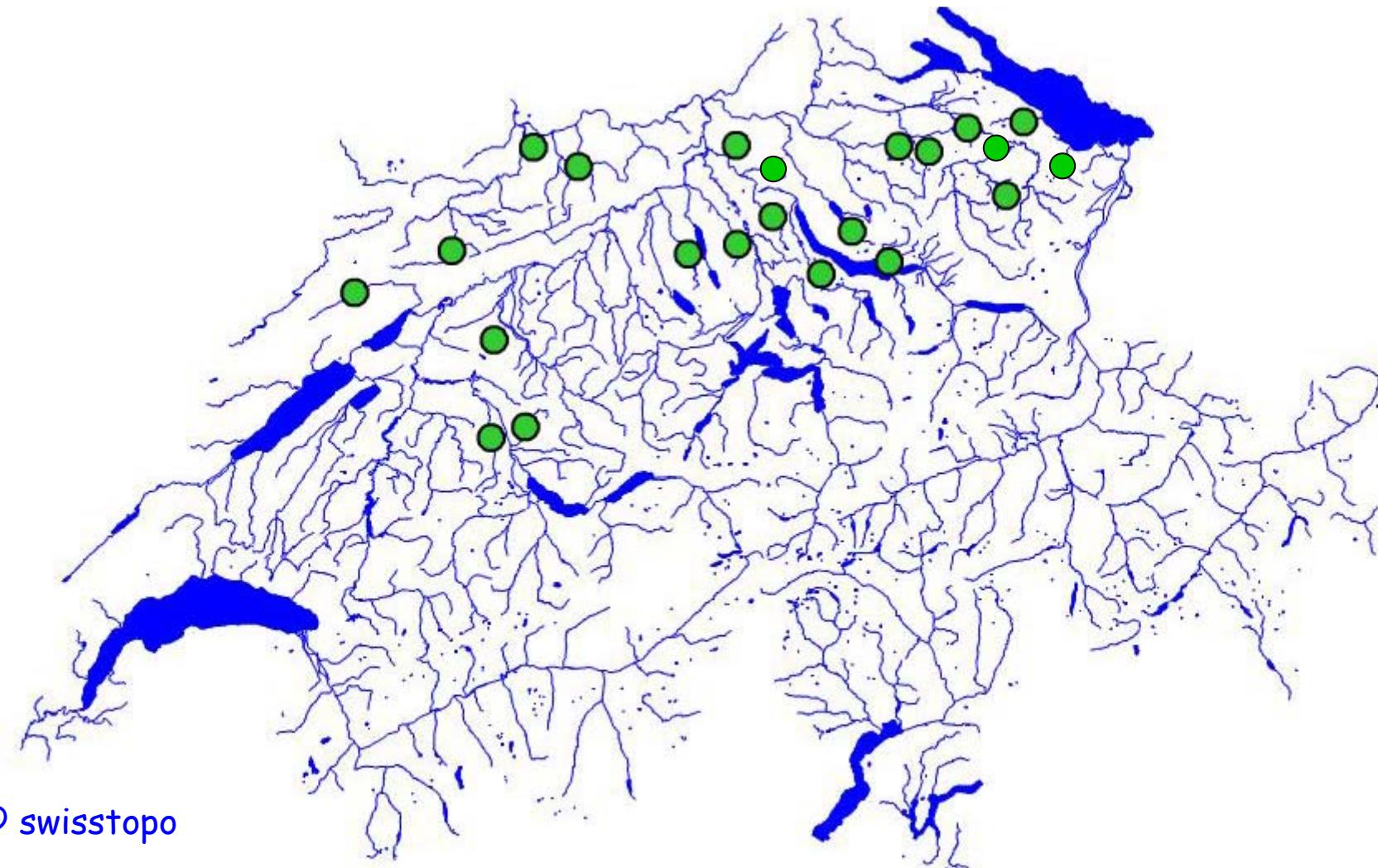
POCIS feasibility test

Vermeirssen et al. 2005 ES&T 39/ 2008 Chimia 62

- Do POCIS see the effluent?
 - Do POCIS data correlate with repeated grab sampling data?
 - Do POCIS compare to bioaccumulation?
 - Do bioassay data relate to LC/MS/MS?



Further evaluation of POCIS – EPSA



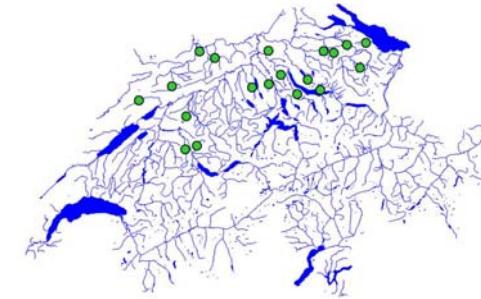
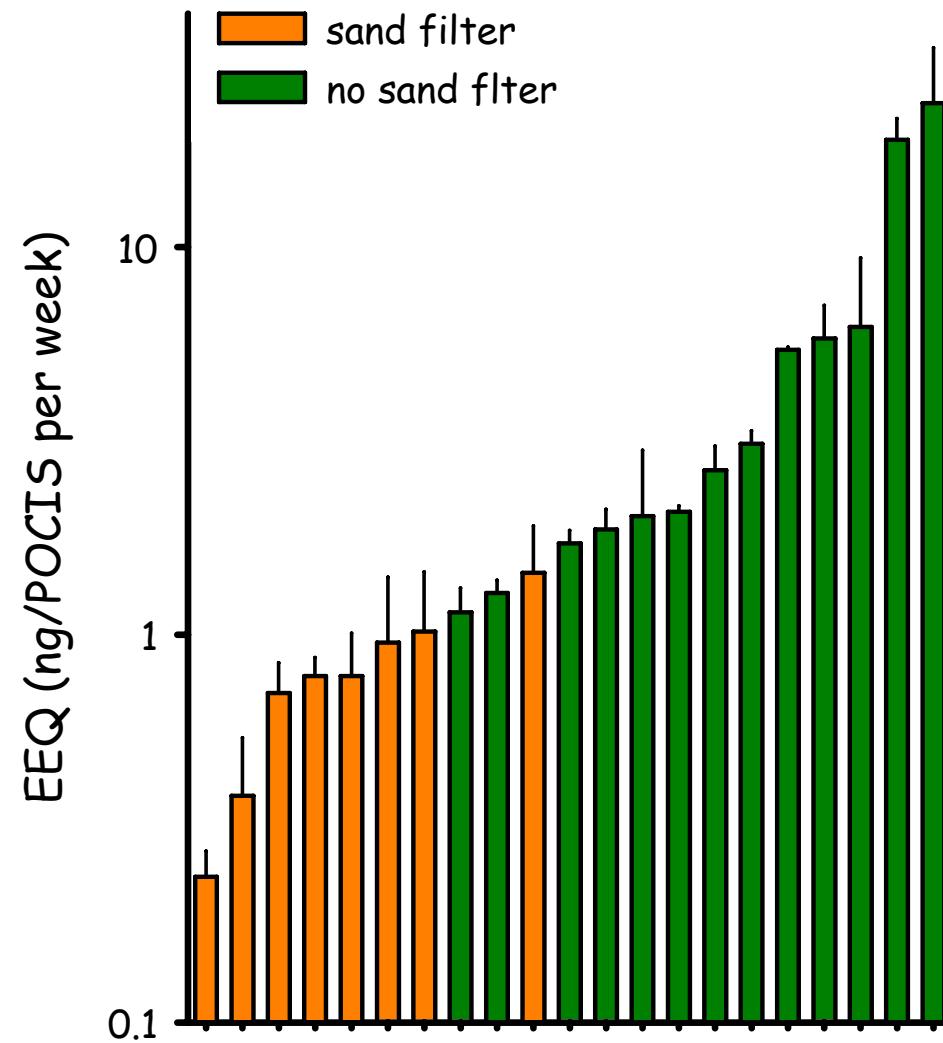
Deployment in effluents (and rivers)



EPSA – results: YES

POCIS in effluent for 5 weeks

Vermeirissen et al. (2008) Chimia 62



Other studies on POCIS estrogens and YES



Available online at www.sciencedirect.com



Science of the Total Environment 367 (2006) 616–630

**Science of the
Total Environment**

An International Journal for Scientific Research
into the Environment and its Relationship with Humanity

www.elsevier.com/locate/scitotenv

Contamination of headwater streams in the United Kingdom by oestrogenic hormones from livestock farms

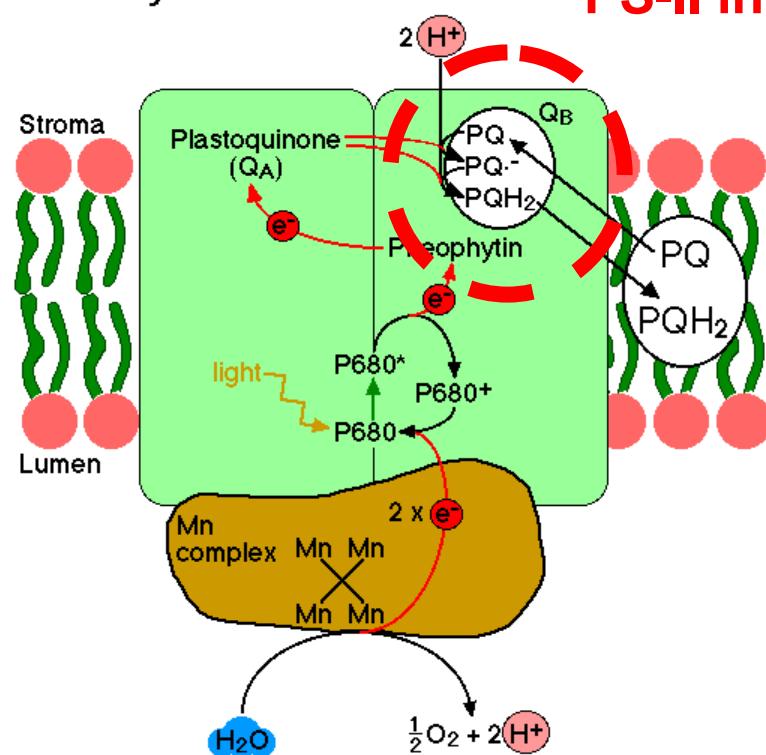
P. Matthiessen ^{a,*}, D. Arnold ^b, A.C. Johnson ^c, T.J. Pepper ^b,
T.G. Pottinger ^a, K.G.T. Pulman ^a

- Calibration of the POCIS
- LC/MS/MS
- YES
- Link between biological and chemical analysis

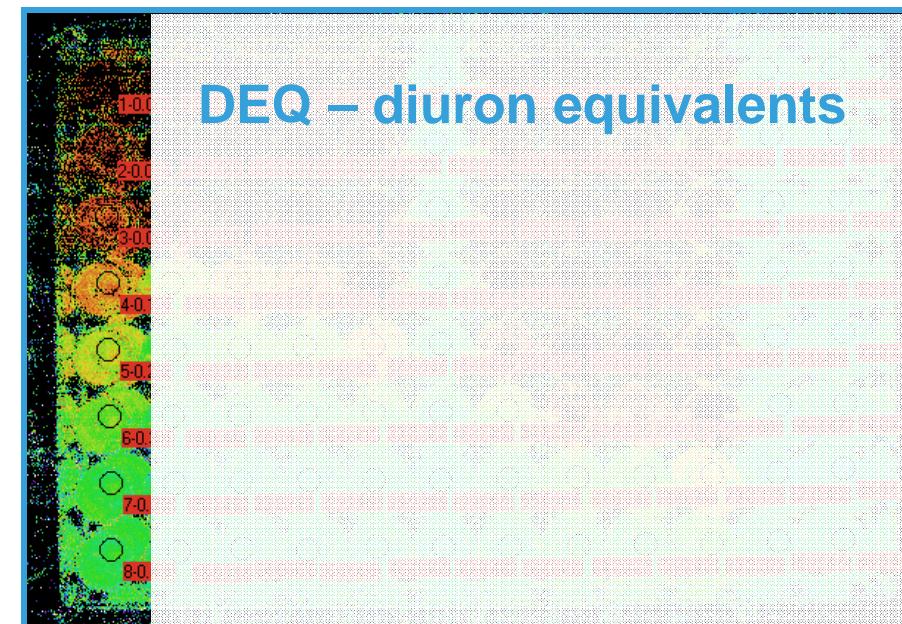
Combined algal test

Escher et al. 2008 J. Environ. Monit. 10

Photosystem II



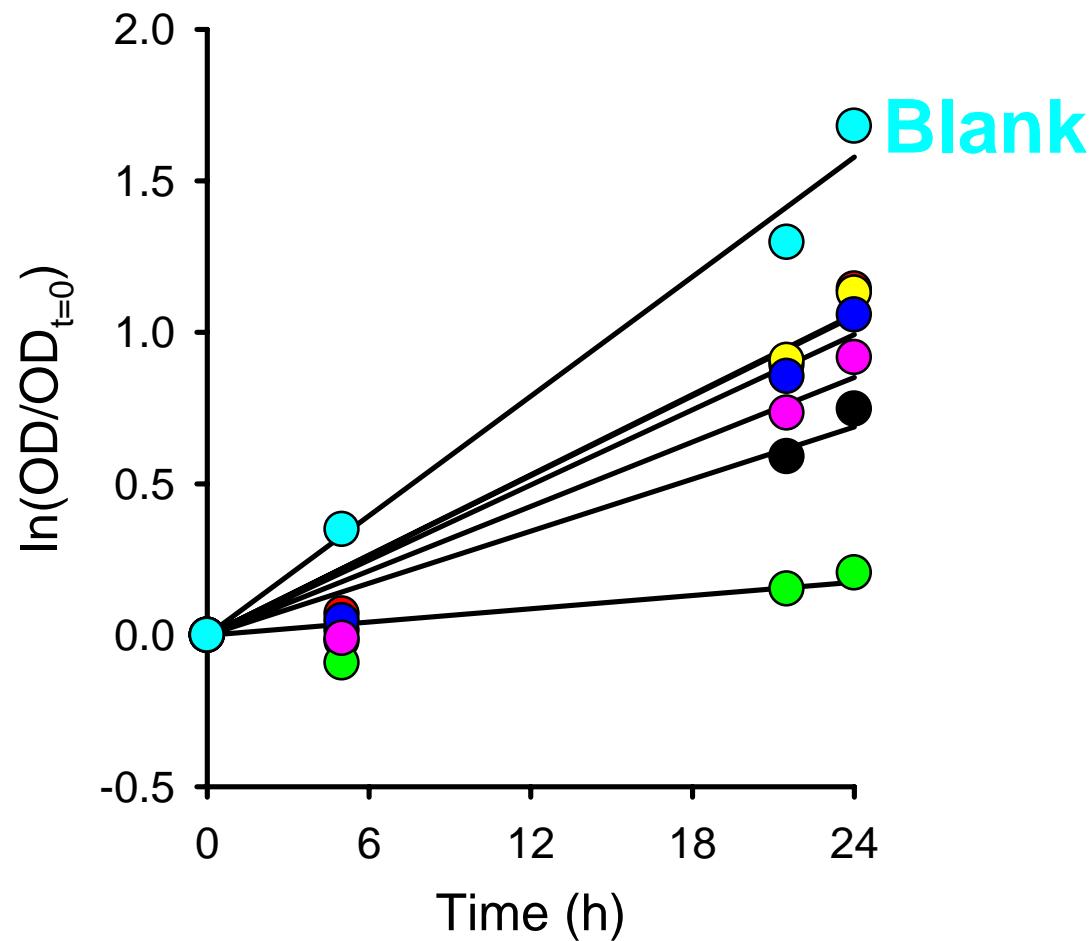
PS-II inhibitors, e.g. diuron



Combined algal test – inhibition of algal growth

Escher et al. 2008 J. Environ. Monit. 10

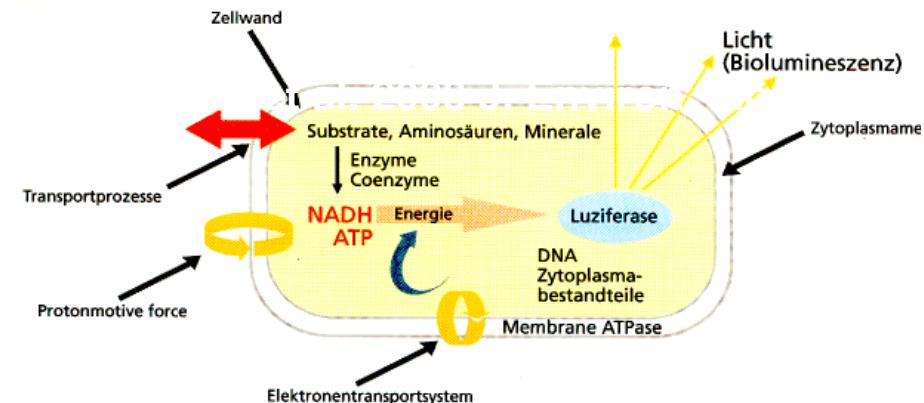
Output: TEQ – toxicity equivalent concentration



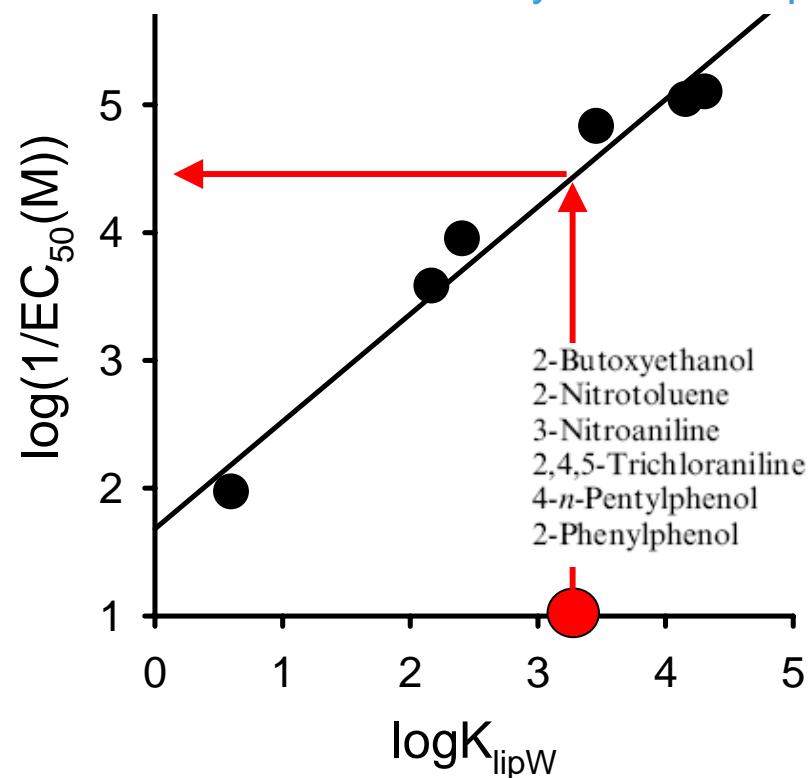
Bacterial bioluminescence



Escher et al. 2008 J. Environ. Monit. 10



QSAR:
quantitative structure-activity relationship



Output: TEQ

Virtual toxicant: logK_{OW} = 3

$$\log K_{\text{lipW}} = 3.2$$

$$EC_{50} = 4.1 \times 10^{-5} \text{ M}$$

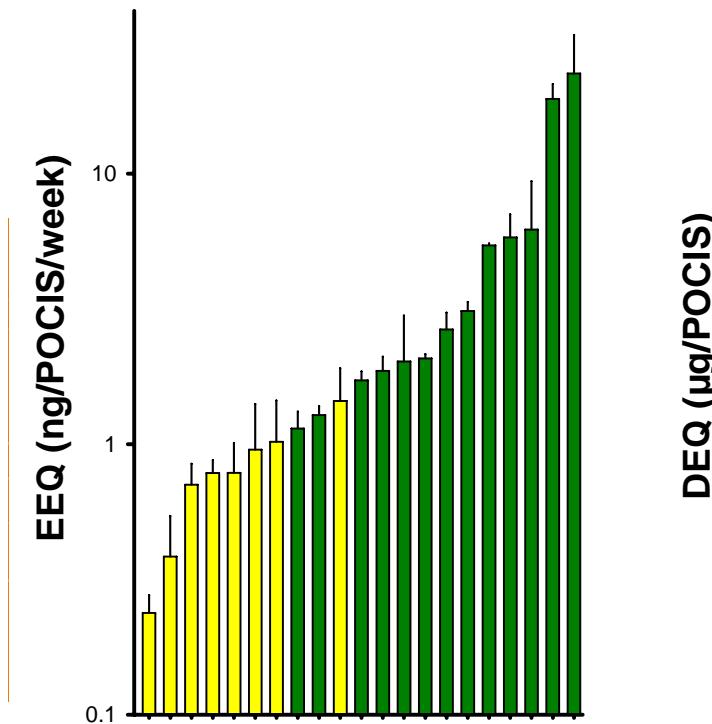
$$MW = 300$$

POCIS: 21 effluents – 3 bioassays

Vermeirssen et al. (in preparation): Micropollutants in effluent-exposed passive samplers – linking toxicity in algal and bacterial assays with chemical analysis

Estrogens

100-fold



PS-II inhibition

160-fold

DEQ ($\mu\text{g}/\text{POCIS}$)

Non-specific

11-fold

TEQ (mg/POCIS)

Linking bioassays with chemical analyses

Diuron equivalents ($\mu\text{g}/\text{POCIS}$)

Calculated diuron equivalents ($\mu\text{g}/\text{POCIS}$)

ng x

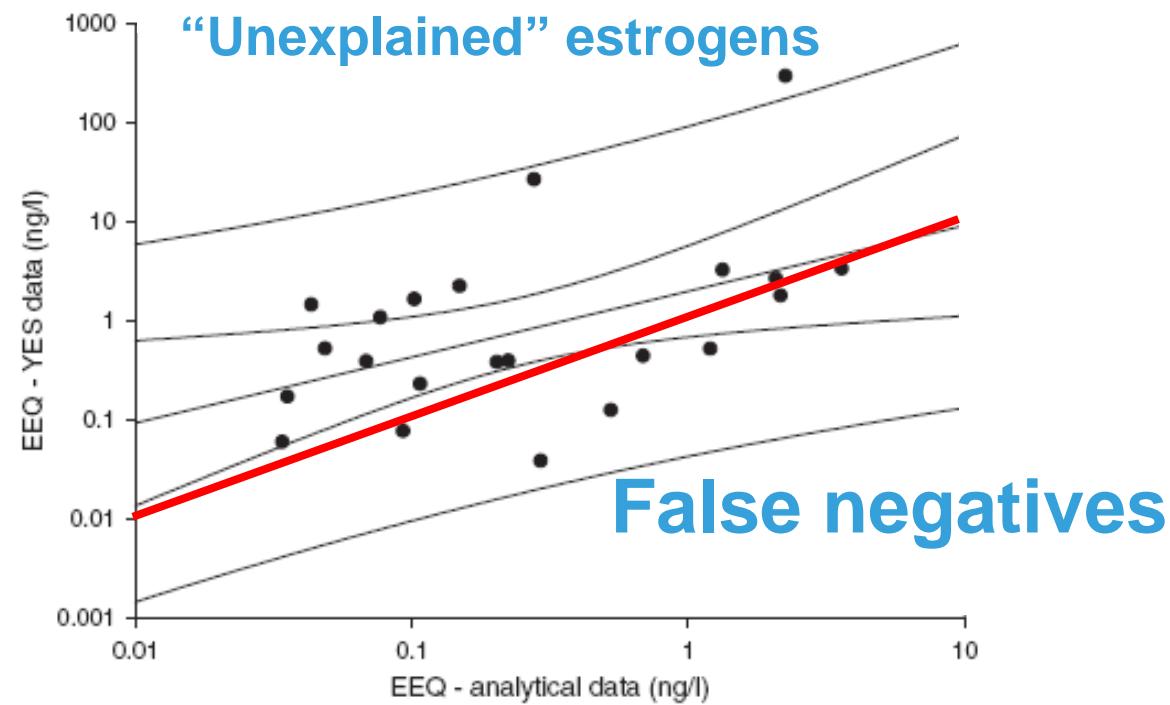
Diuron
Terbutryn
Isoproturon
Atrazine
Irgarol
Terbutylazine

Relative potencies

=	1.00
=	0.78
=	0.16
=	0.11
=	2.51
=	0.46

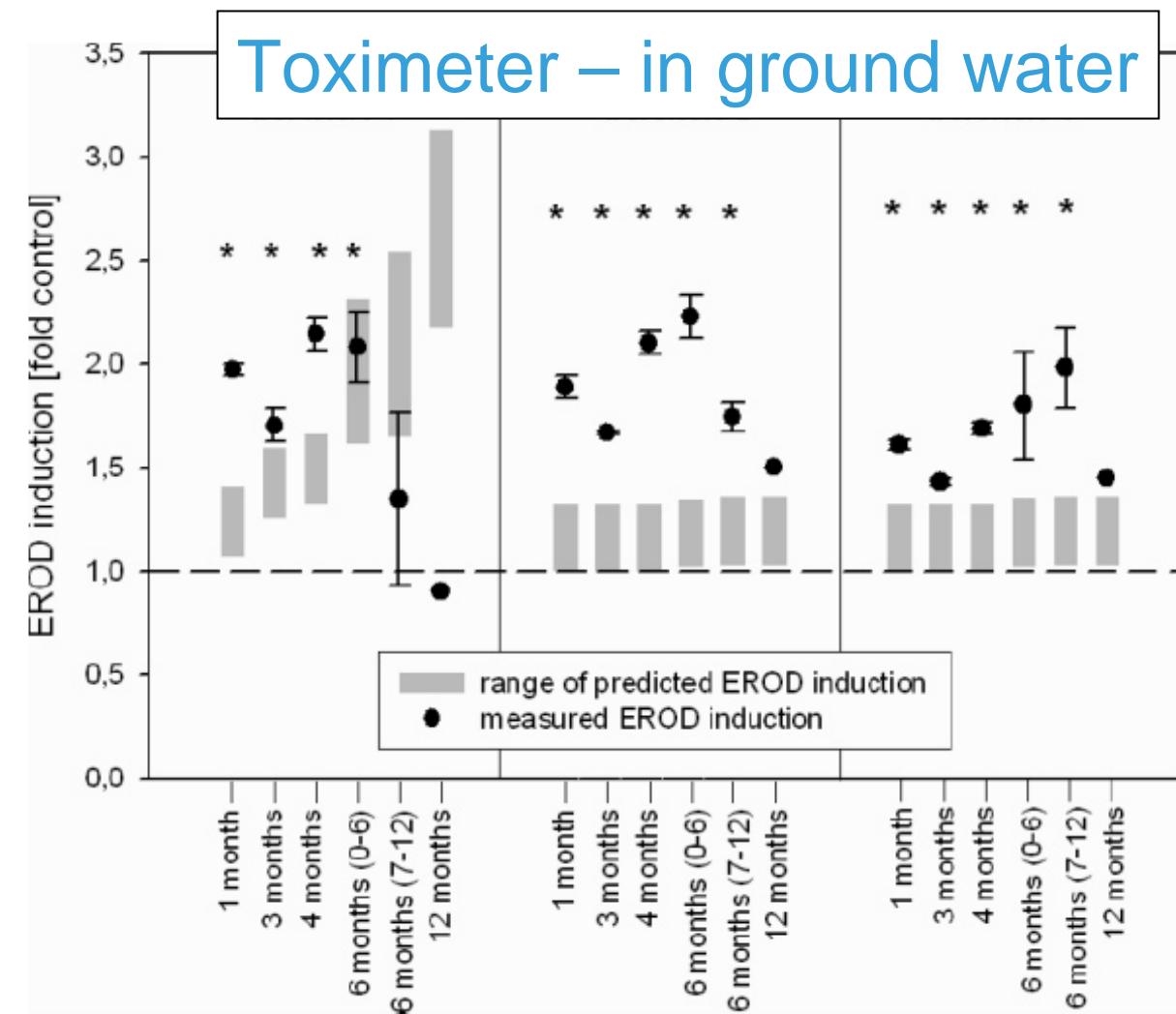
sum of herbicides

Bioassays and chemical analyses – other examples



Bioassays and chemical analyses – other examples

Rastall et al. 2004 ESPR 11: on SPMDs, EROD, YES, AMES test



Passive sampling and flow rate

$$Sh = k_f \frac{L}{D_{AB}} = 0.664 \cdot Re^{0.5} \cdot Sc^{1/3} \cdot [k_b t]$$

$$k_u = k_b K \quad Re = \frac{Lv\rho}{\mu} \quad (V_m)$$

Stephens et al. 2005 ES&T 39

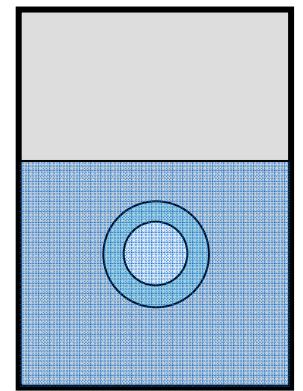
Flow rate

“Channel trials”

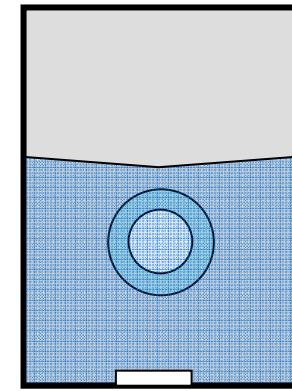


Effects of flow: basic set-up

Alvarez et al. 2004 ETC 23



No stirring

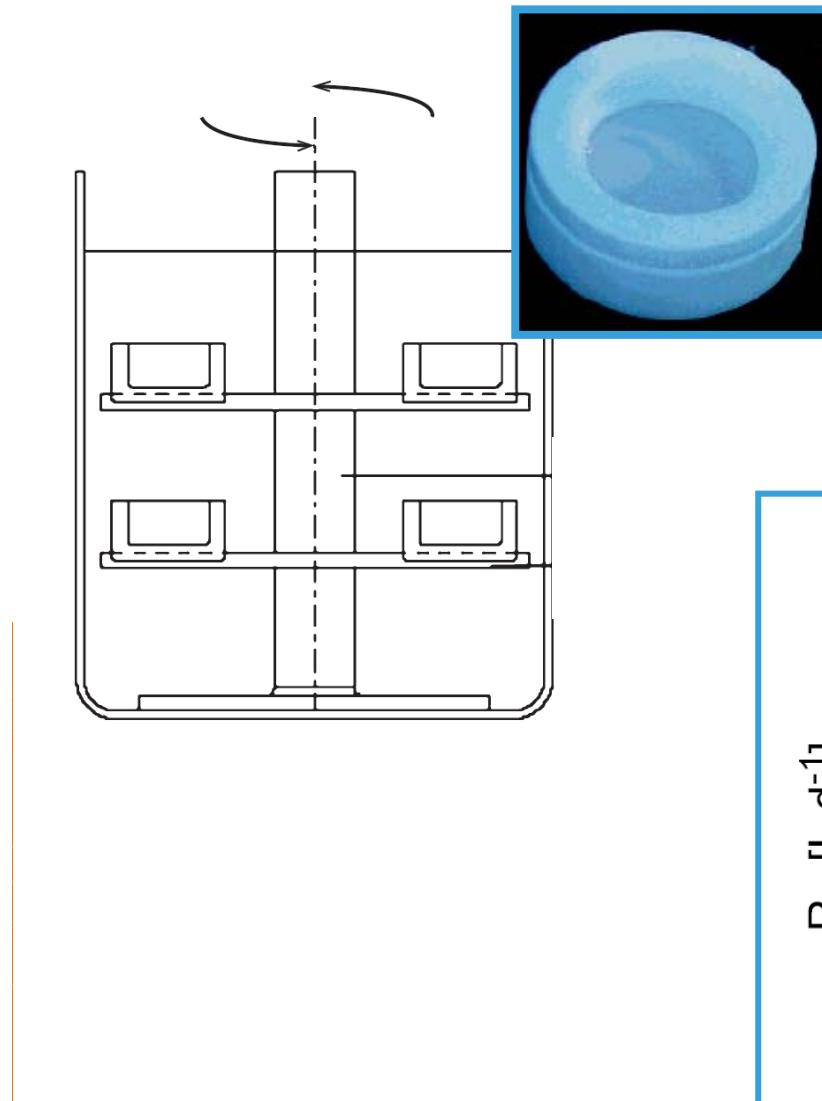


Stirring

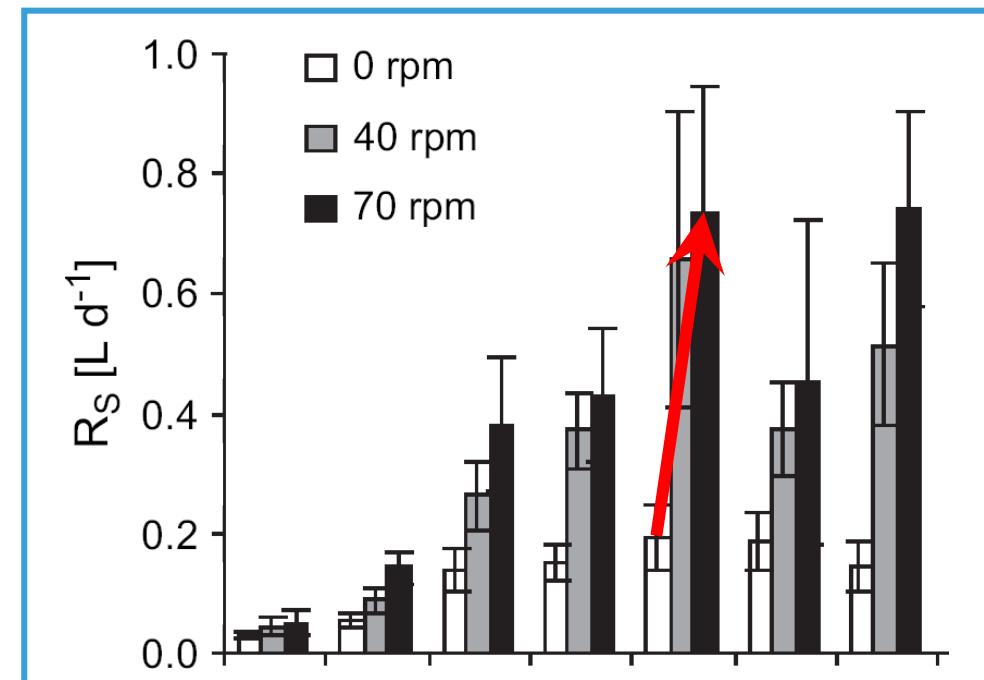
Analyte	R_s from quiescent renewals (L/d)	R_s from turbulent renewals (L/d)
Diuron	0.005 (0.002)	0.045 (0.016)
Isoproturon	0.015 (0.003)	0.086 (0.008)
Azithromycin	0.021 (0.006)	0.120 (0.075)
Fluoxetine	0.012 (0.007)	0.086 (0.023)
Levothyroxine	0.009 (0.008)	0.053 (0.028)
Omeprazole	0.007 (0.004)	0.030 (0.008)

Rotating samplers

Vrana et al. 2006 Environ. Pollut. 142

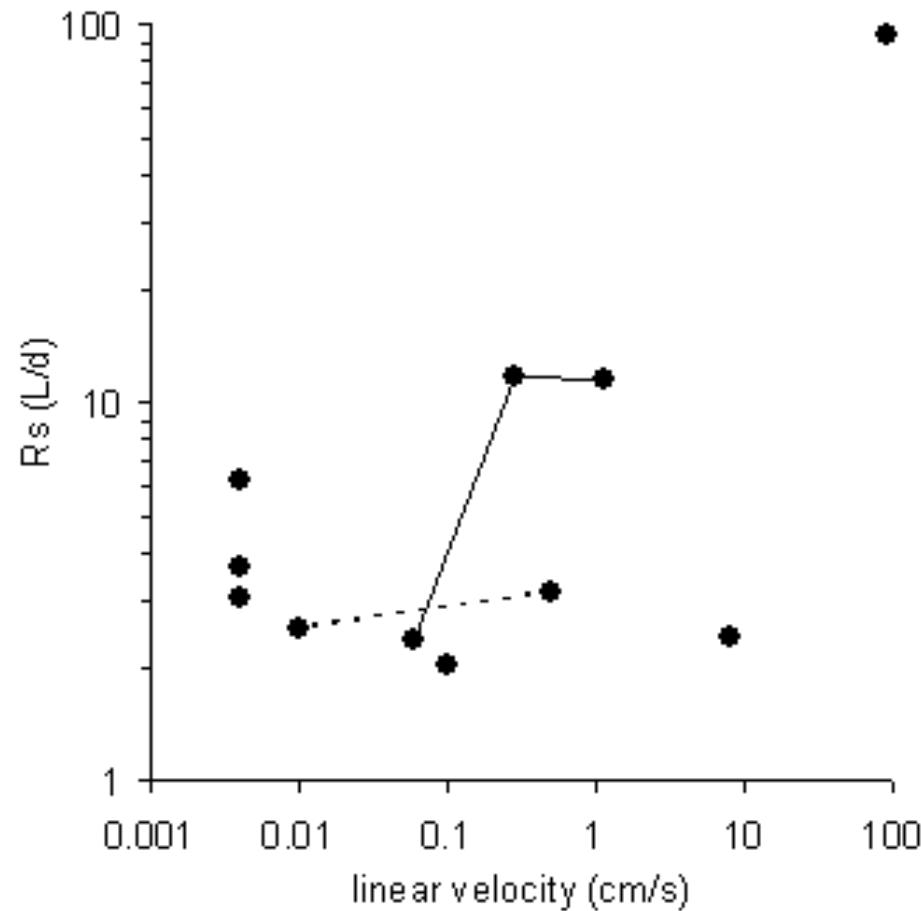


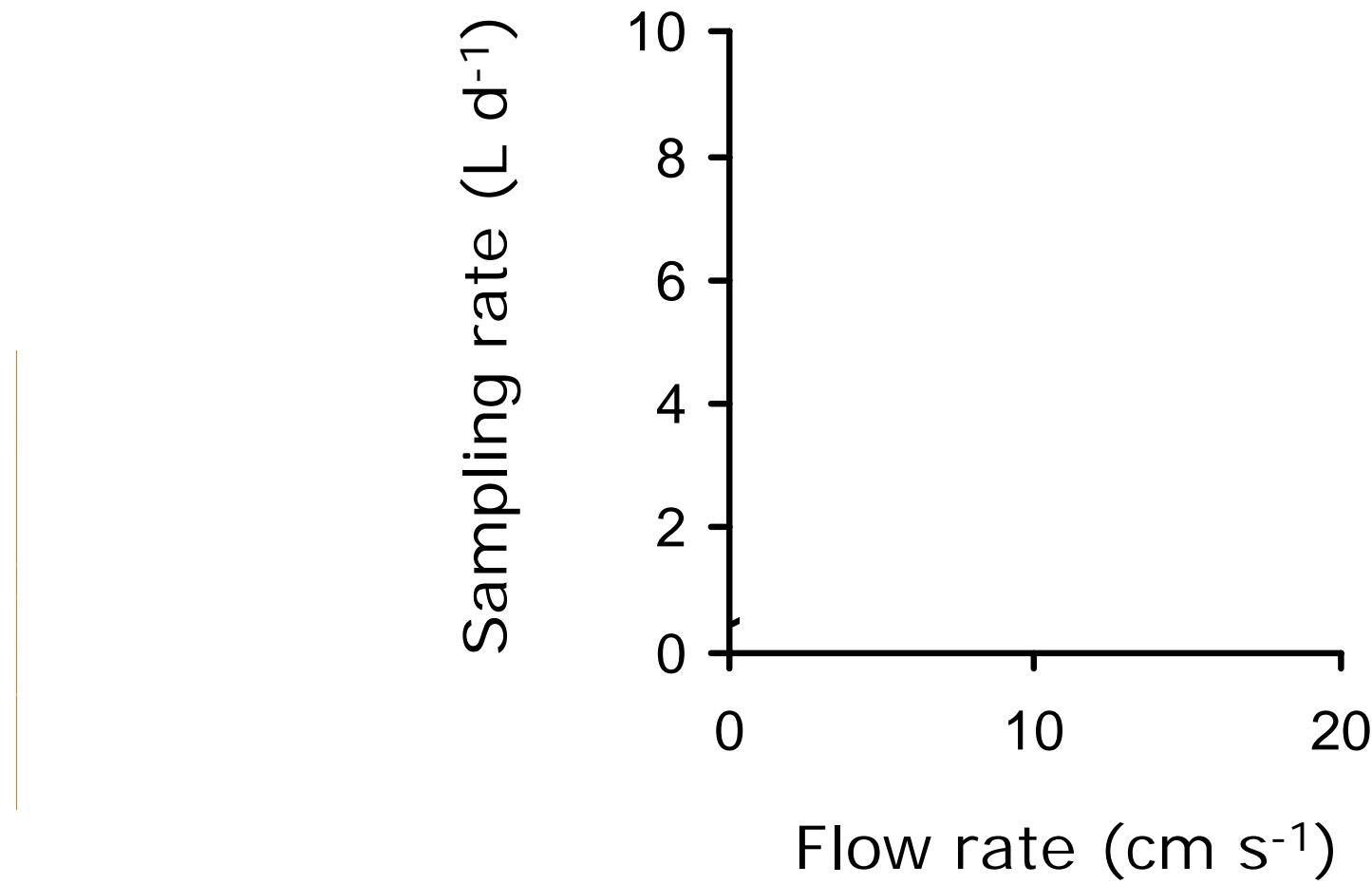
Microcosm running at 2 to 50 cm/s
Mazzella et al. 2008 Chemosphere 73



SPMDs and effects of flow rate

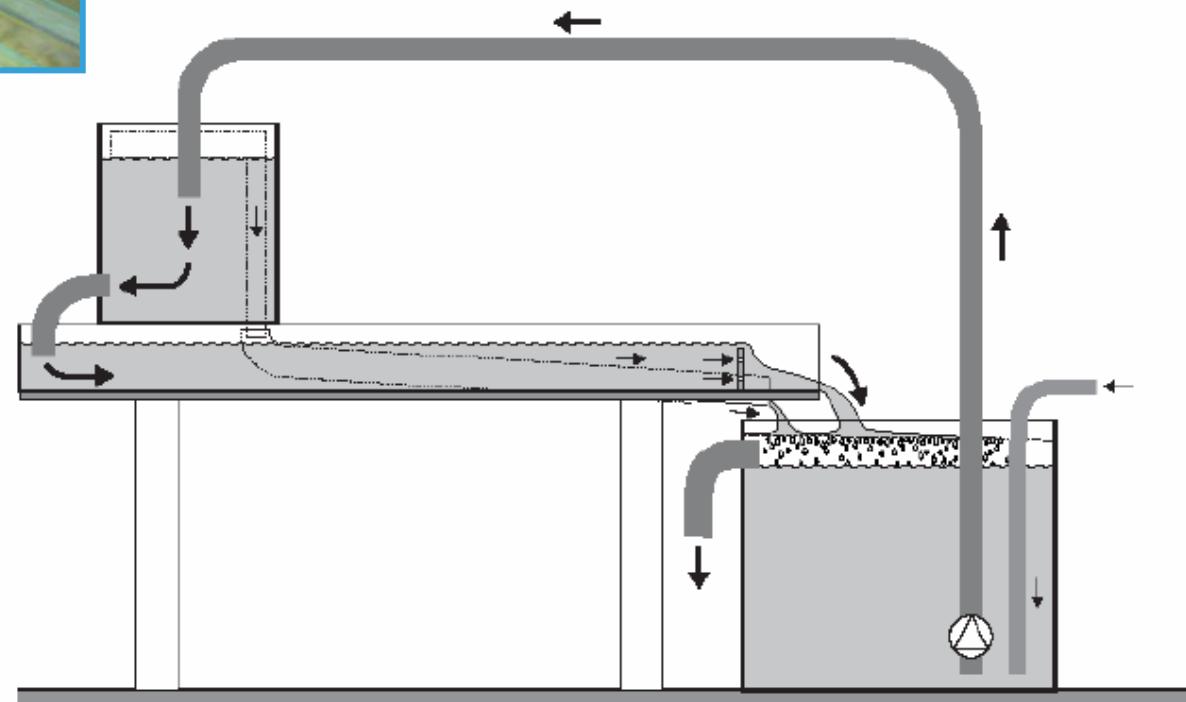
Fig 3.6 from “The SPMD book”: Huckins, Petty and Booij (2006)





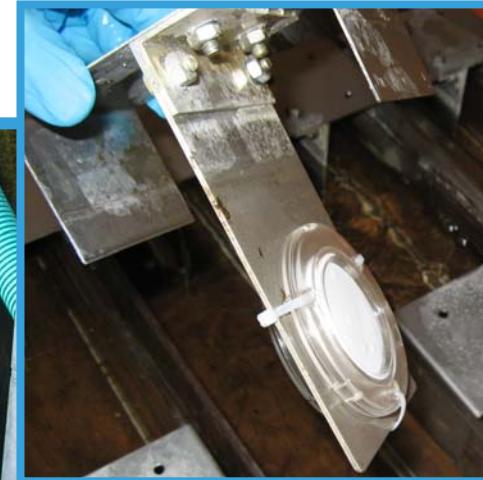
Channel system set-up

Vermeirssen et al. 2008 J. Environ. Monit. 10



Environmental conditions

Vermeirssen et al. 2008 J. Environ. Monit. 10



Channel system running with effluent

Vermeirssen et al. 2008 J. Environ. Monit. 10

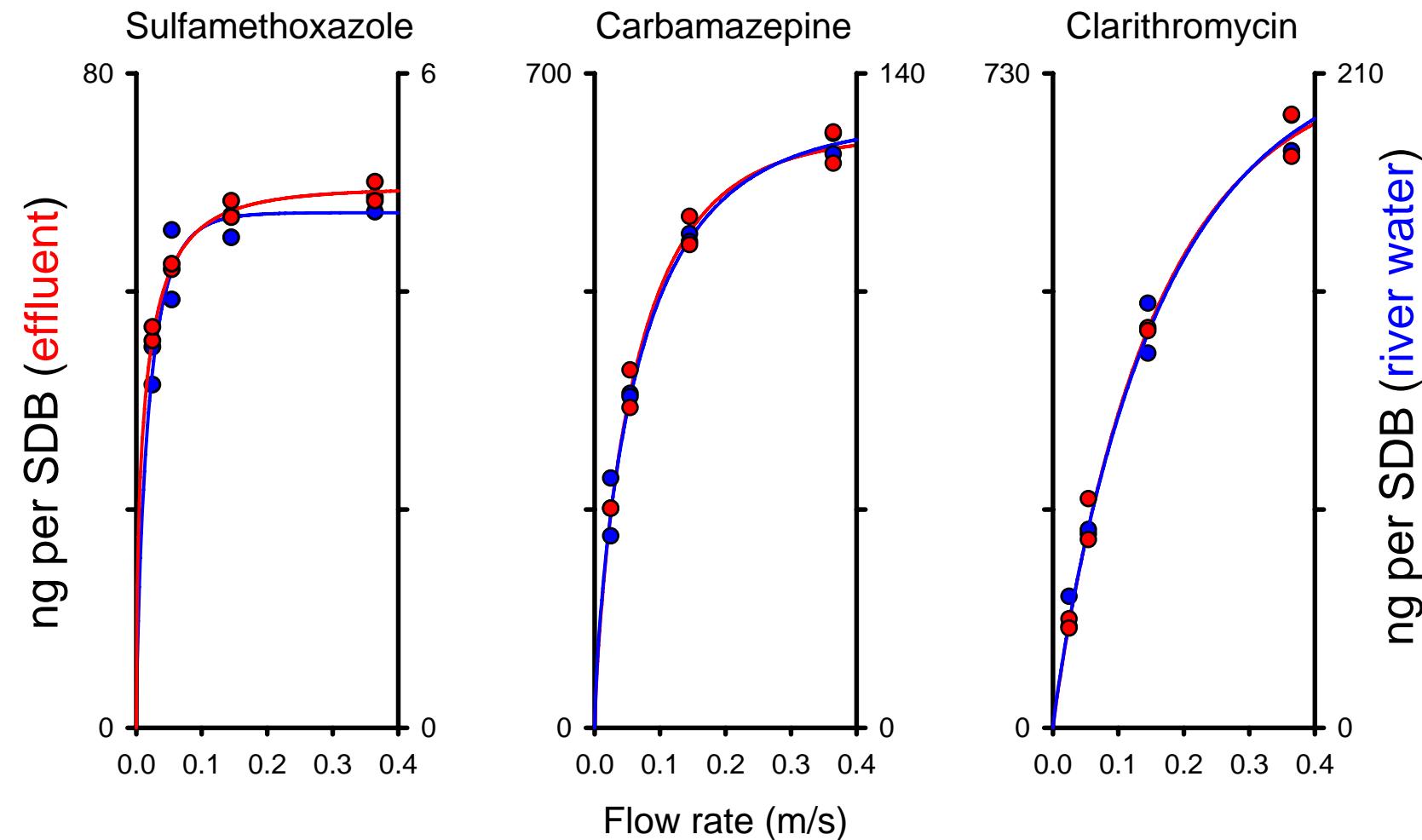


Three pharmaceuticals in two matrices

Empore SDB-RPS disks

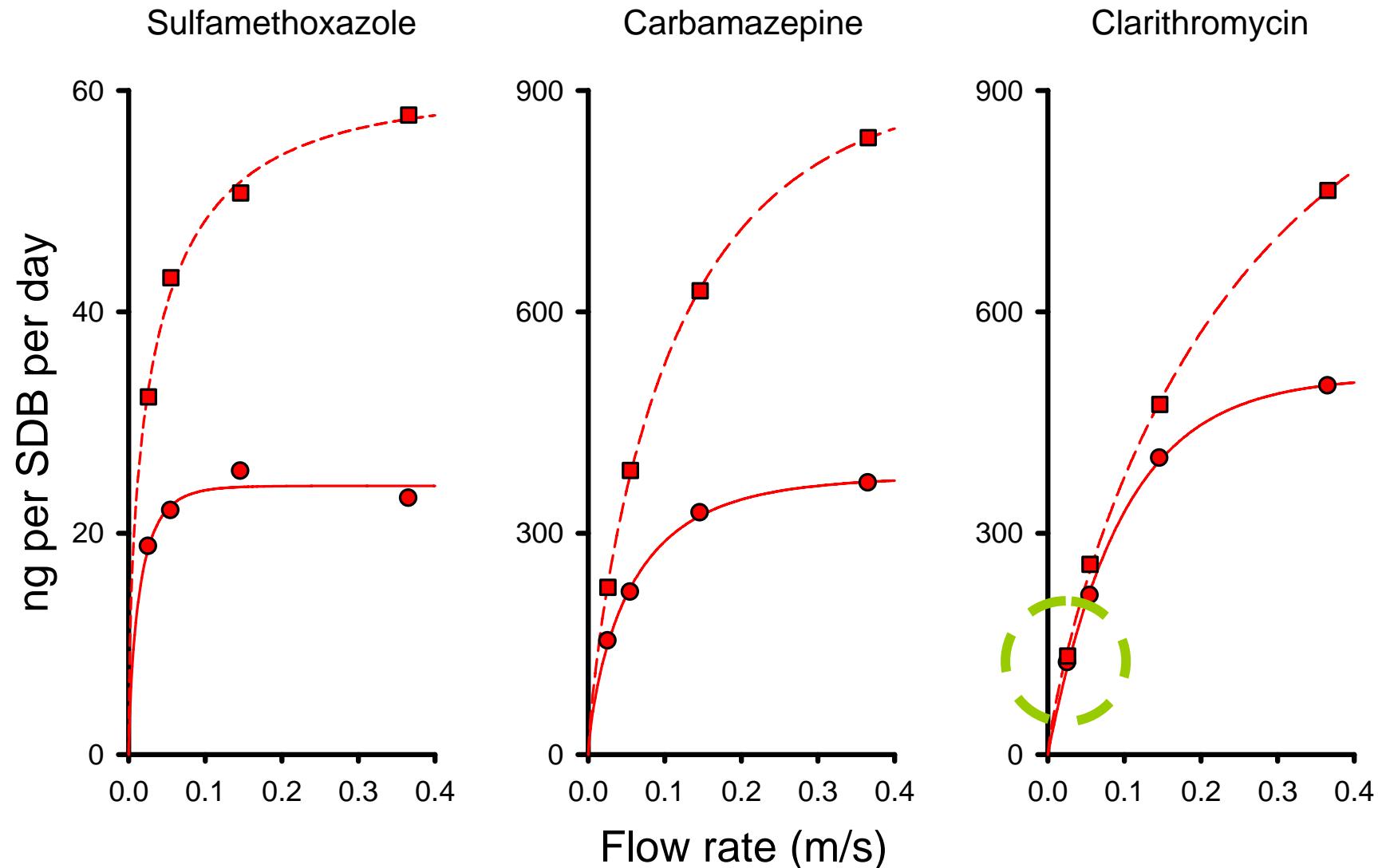
Vermeirssen et al. 2008 J. Environ. Monit. 10

Salinity: Togola & Budzinski 2007 AC 79



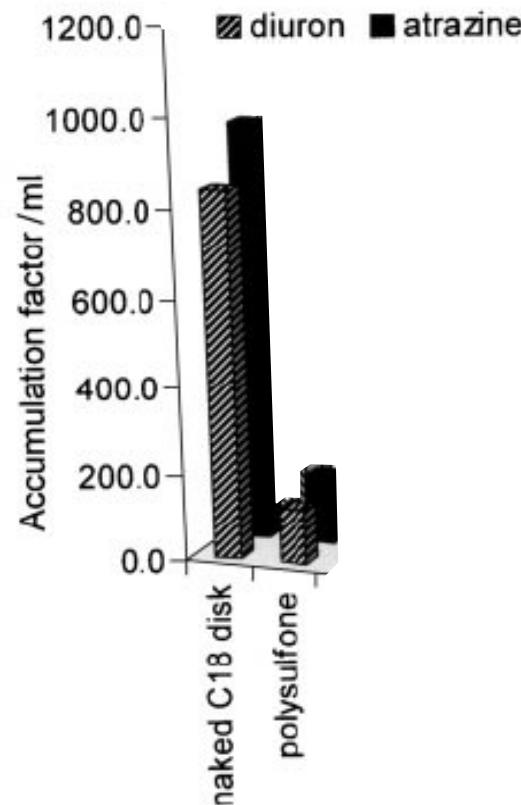
Three pharmaceuticals; overlapping exposure slots

Vermeirssen et al. 2008 J. Environ. Monit. 10



Extending the integrative sampling window

Cover the Empore disk
with a **membrane**



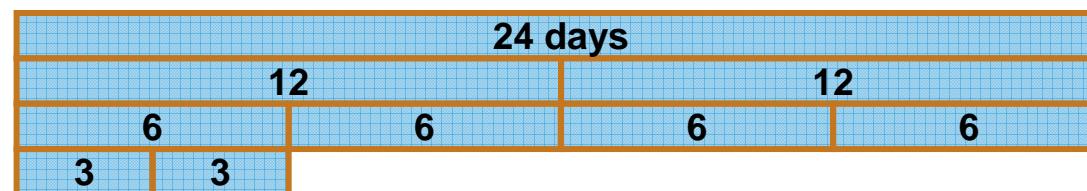
Explore Empore **SDB-XC**

Gunold et al. 2008 Env. Pollut. 155
Approximate linear uptake for 12 days

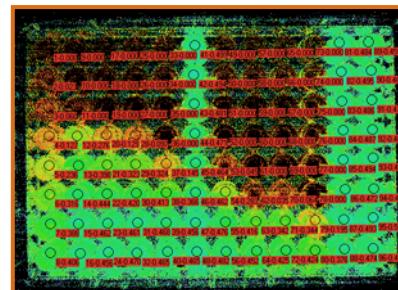
Kingston et al. 2000 J. Env. Monit. 2 Figure 2
see also Tran et al. 2007 ET&C 26

Three experiments

1. 5 day flow rate trials with **SDB-RPS** and **SDB-RPS-PES** (polyethersulfone)
2. 5 day flow rate trials with **SDB-RPS** and **SDB-XC**
3. overlapping sampling blocks with all three configurations at 0.1 m/s

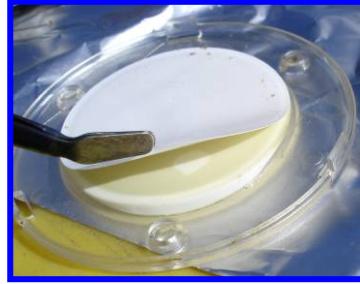
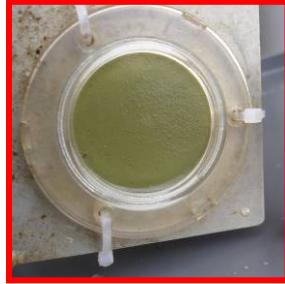


Combined bioanalysis and chemical analysis

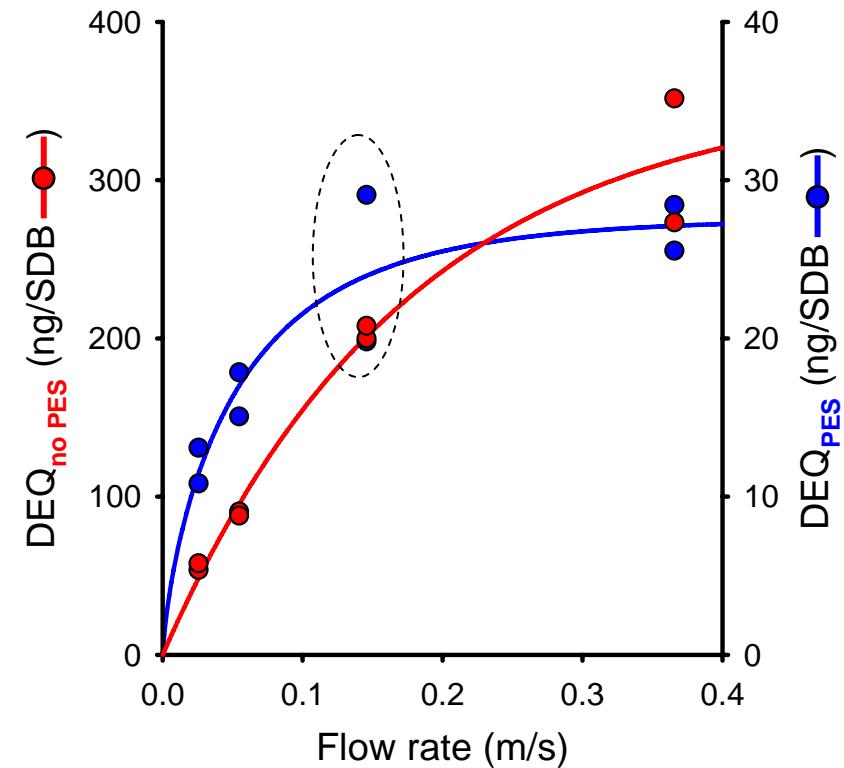


Adding a PES membrane

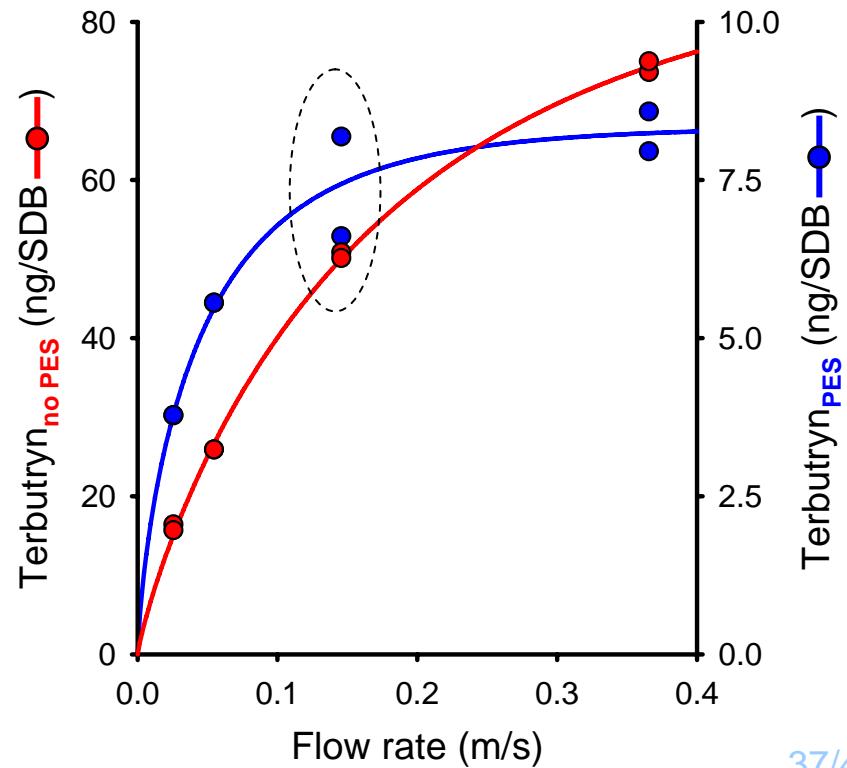
Vermeirssen et al. 2008 Wat. Res. 43



Diuron equivalents

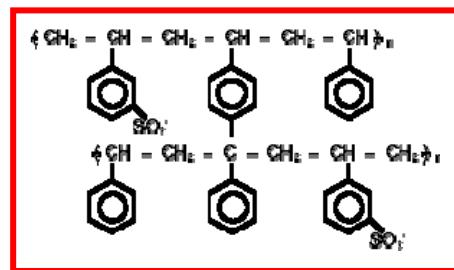


Terbutryn



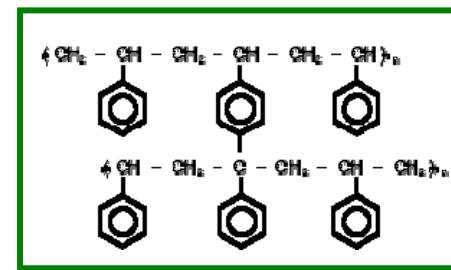
Comparing RPS and XC

Vermeirissen et al. 2008 Wat. Res. 43

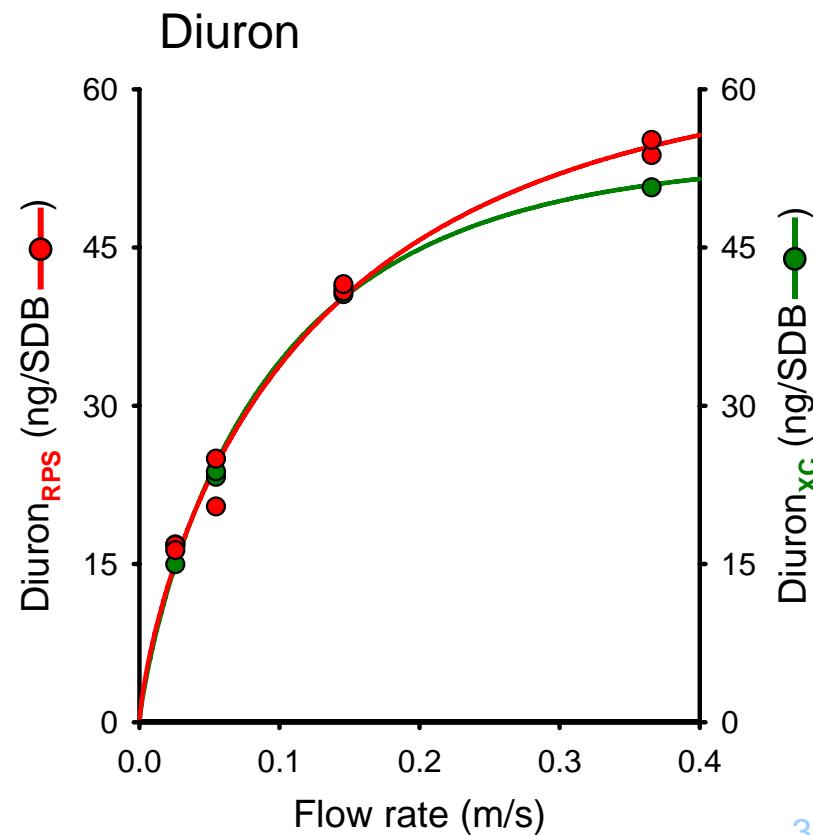
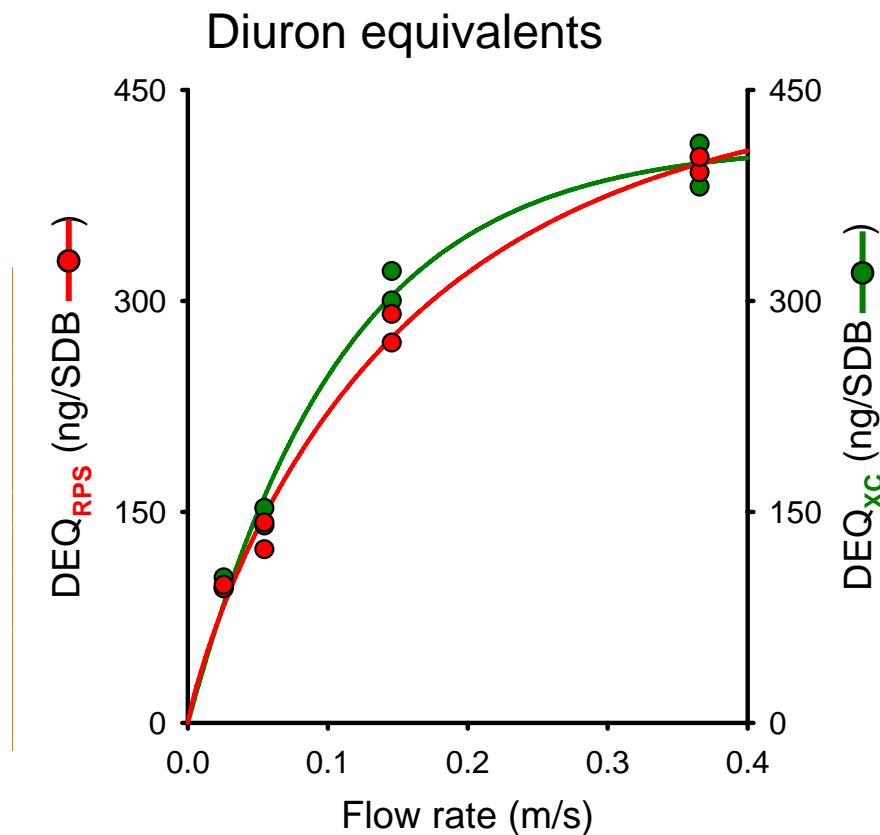


RPS

sulfonic acid groups

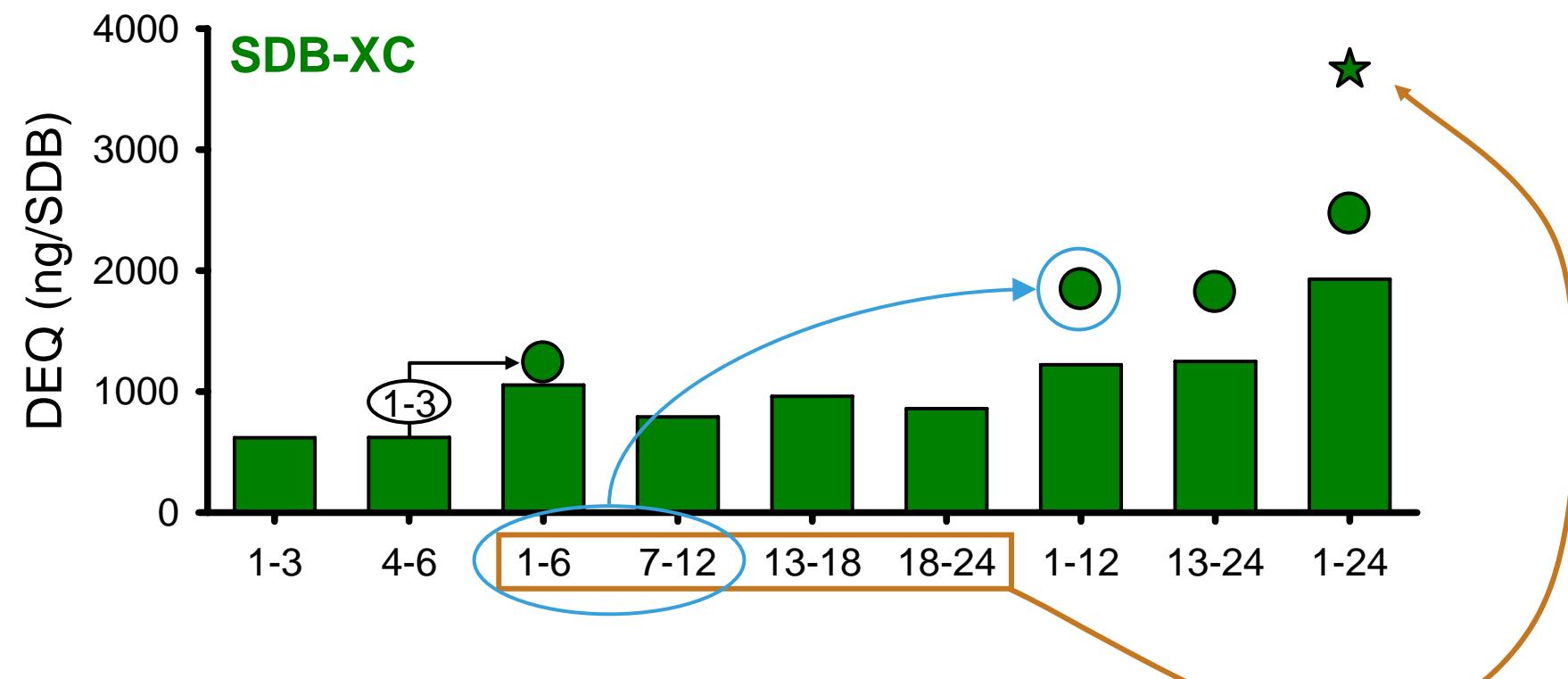


XC



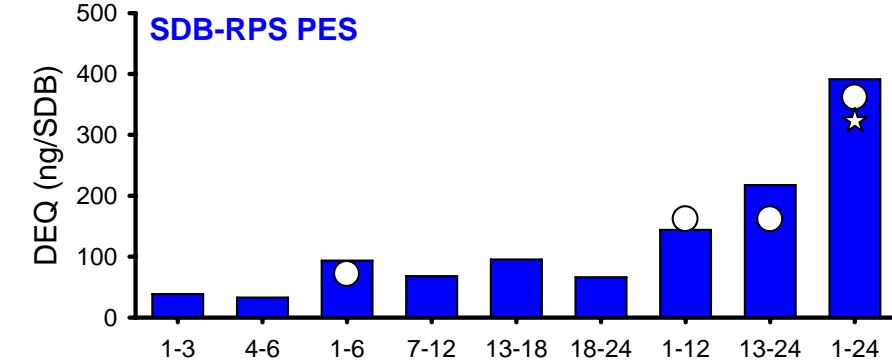
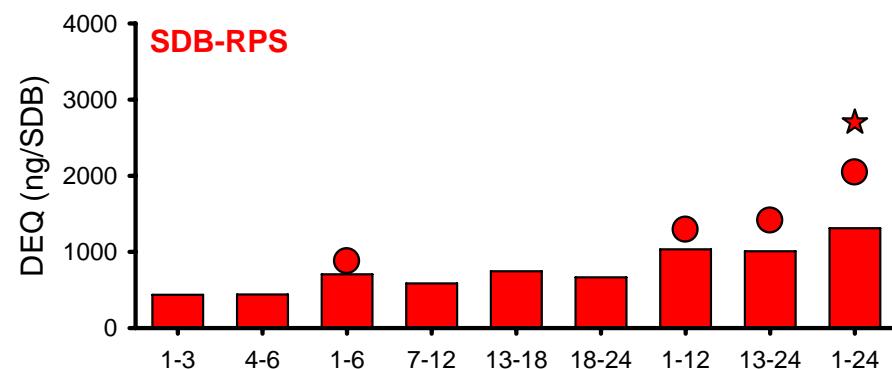
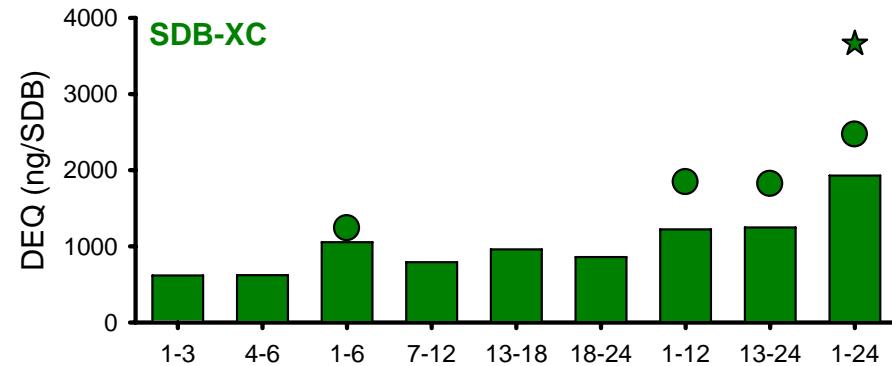
Duration of integrative period

Vermeirssen et al. 2008 Wat. Res. 43

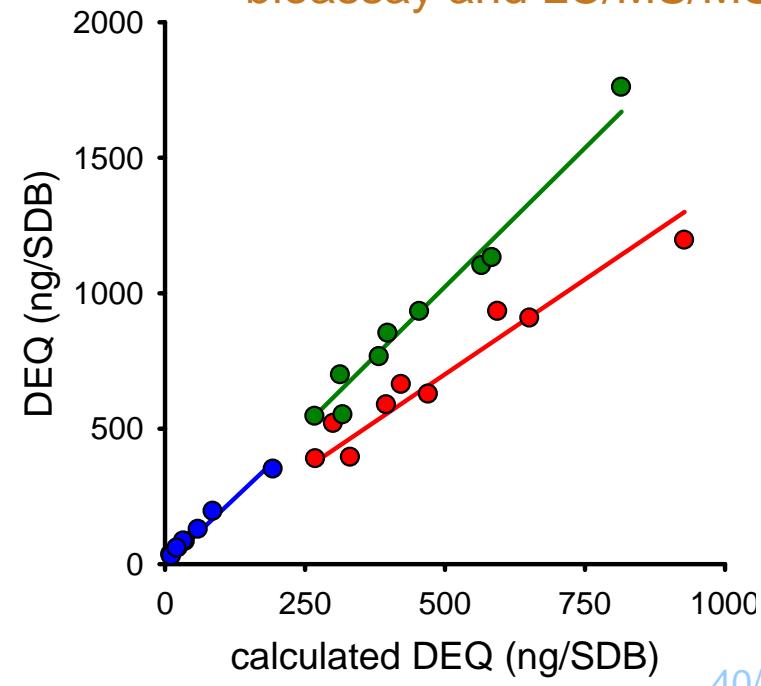


Three Chemcatcher configurations – 24 days

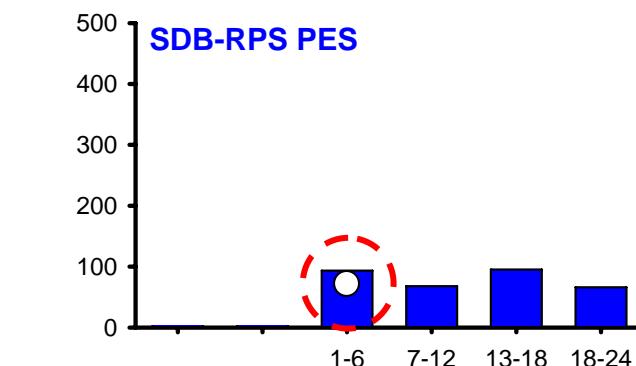
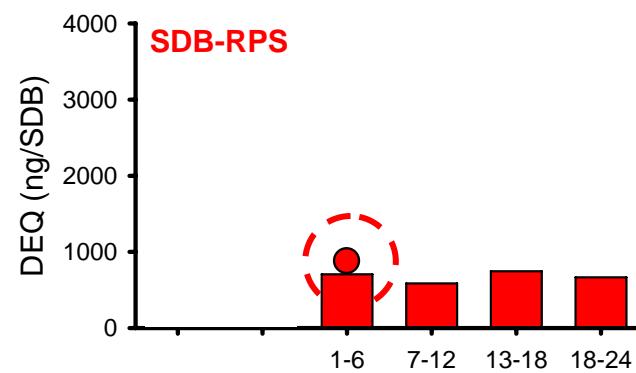
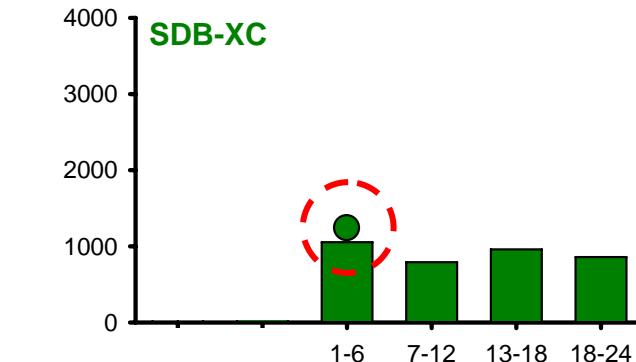
Vermeirssen et al. 2008 Wat. Res. 43



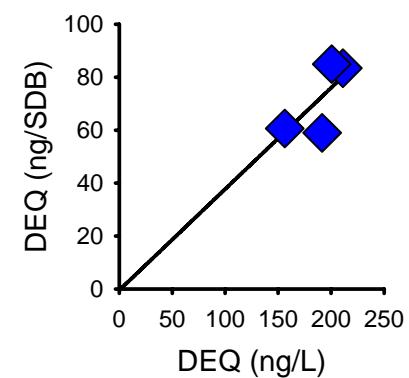
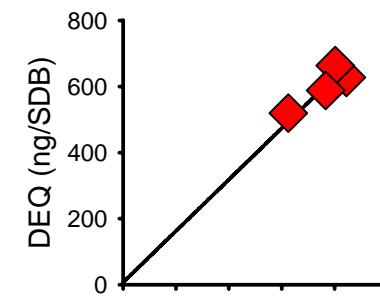
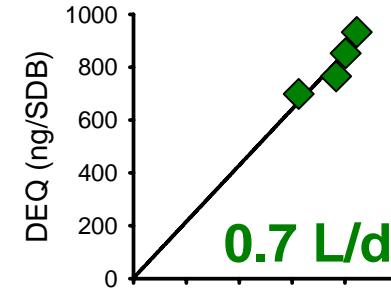
Again, a good association between bioassay and LC/MS/MS



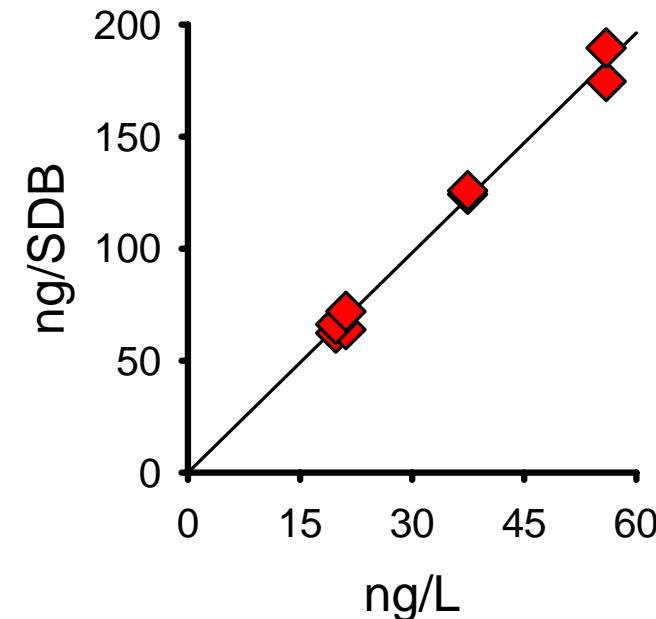
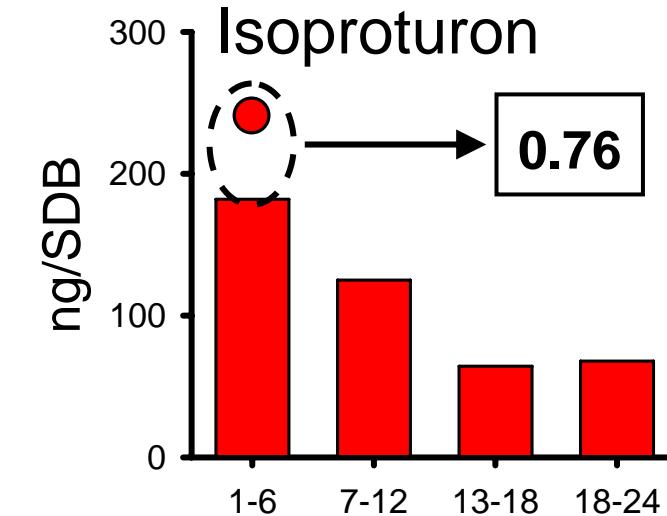
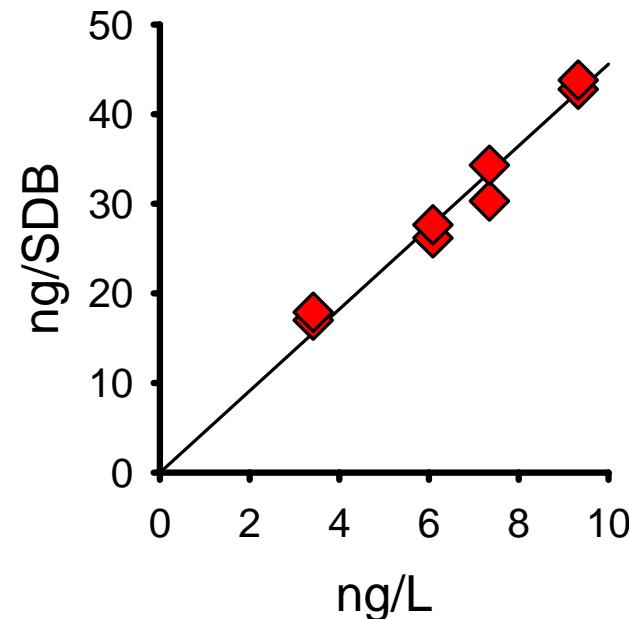
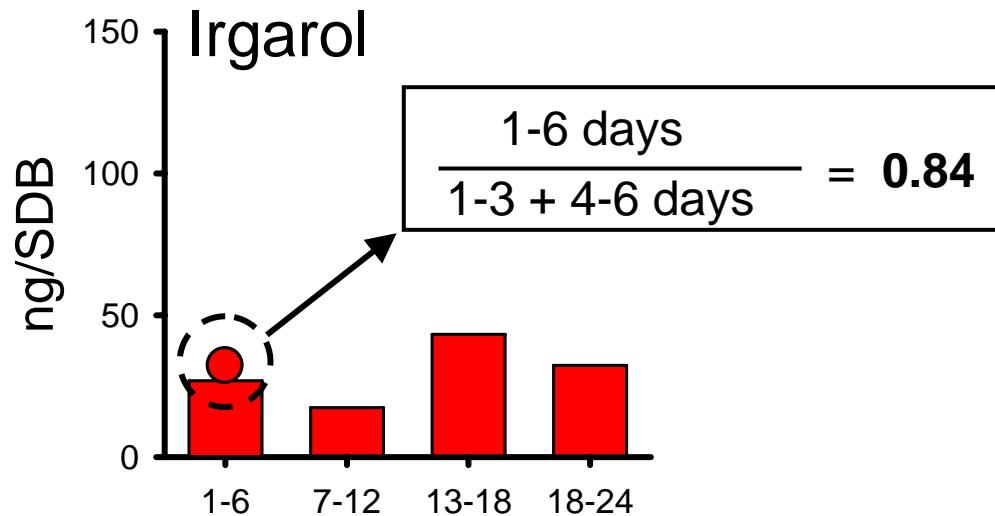
Estimating sampling rates



4 times a 6 day
water sample

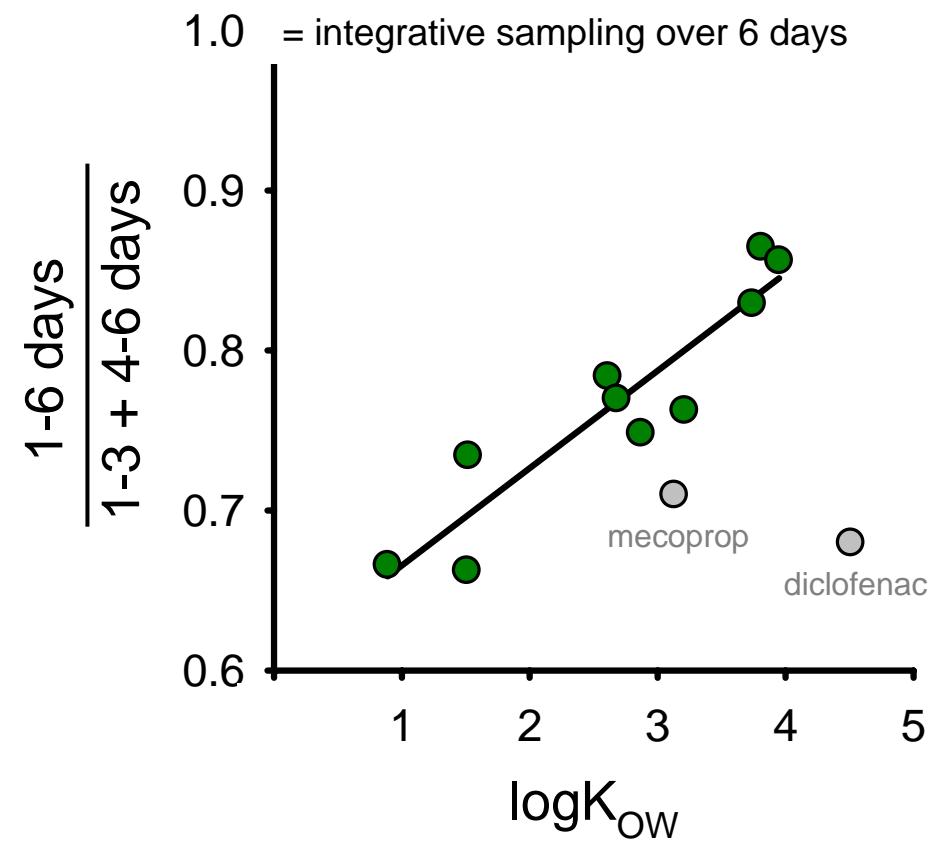
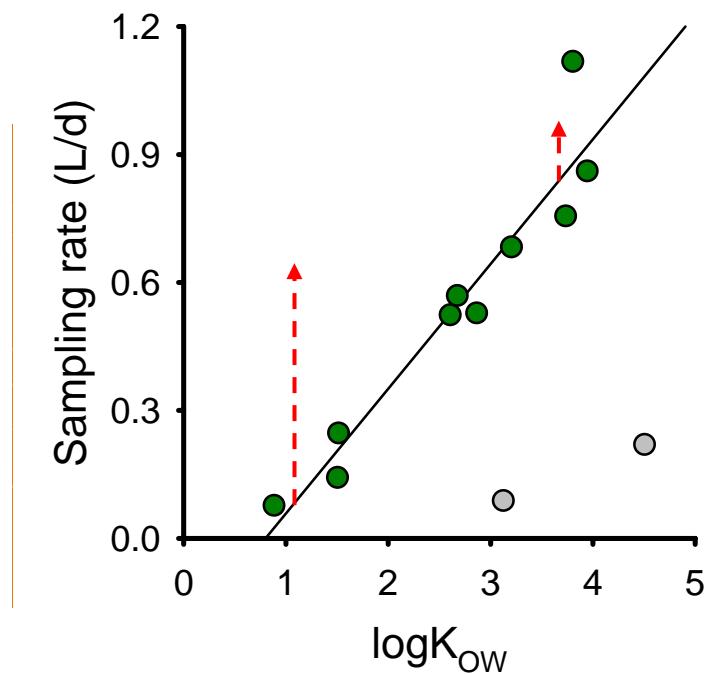


An integrative ratio

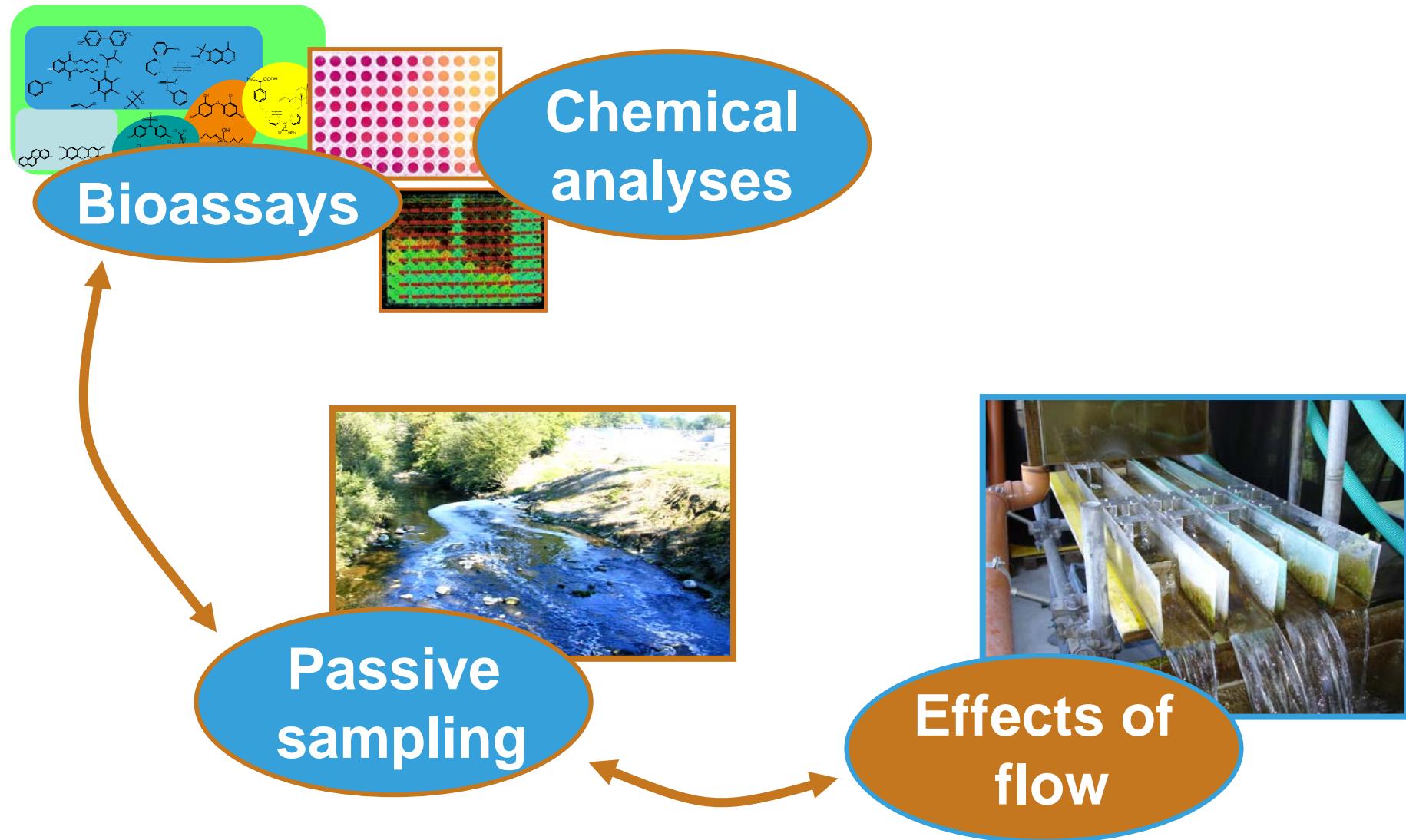


Sampling properties and logK_{OW}

Vermeirssen et al. 2008 Wat. Res. 43



Summary



Acknowledgements

Technicians: Nadine Bramaz
Alfred Lück
Sibylle Rutishauser
Pam Quayle

Students: Jürgen van der Voet
Oliver Körner

Main co-authors: Beate Escher
Juliane Hollender
Jung-Hwan Kwon

Funding: SNF – Bafu – Eawag



Up to 2009 –
Environmental Toxicology –
Group Escher

Since January 2009 –
Environmental Chemistry
Juliane Hollender

Thank you for your
attention !

